



# Preparation and Characterization of Fish Protein Isolate from Tiger Tooth Croaker (*Otholithus ruber*) Obtained through PH Shift Method

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## Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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## ABSTRACT

Development of Fish Protein Isolate (FPI) from tiger tooth croaker (*Otholithus ruber*) fish meat using the pH shift method was carried out during this study. Tiger tooth croaker was used as raw material because of their abundance and comparatively low price. During the study, the physical characteristics and proximate composition of the fresh fish were analysed. The average length of fish was 19.95 cm and weighed 94.6 g. respectively. FPI treated at different pH treatments (2.5, 4, 7, 11.5 and 12.5) were analyzed for proximate composition, peroxide value and functional characteristics. In the present work, Tiger tooth croaker (*Otolithes ruber*) fish was found to be suitable for fish protein isolate production using acid or alkali processing and isoelectric

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precipitation. During storage at ambient temperature for 120 days in a 200 gauge LDPE pouch, Fish Protein Isolates treated at different pH treatments (2.5, 4, 7, 11.5 and 12.5) indicated an increase in moisture content, reduction in protein and lipid content and no significant reduction in ash content. The total protein content was specifically high for pH 7 followed by pH 12.5, 11.5, 4 and 2.5. The effect of different pH on PV of fish protein isolates showed increasing content with increasing storage. The functional properties such as water-holding capacity (WHC), oil-holding capacity (OHC), emulsifying capacity (EC) and foam measurements (Foaming capacity (FC) and foam stability (FS) exhibit high values for all the samples of fish protein isolates. The quality attribute of functional properties of all fish protein isolates samples showed a trend of decreasing during the storage of 120 days.

**Keywords:** Fish protein isolate; tiger tooth croaker; pH-shift method.

## 1. INTRODUCTION

“As a source of animal protein, humans are highly dependent on seafood. Fishery by-products have received much consideration as an important protein source because of utilizing animal protein as a functional food ingredient” [1]. “Generally, protein providing energy in terms of calories is not used but its contribution to protein synthesis is of high importance and it plays crucial roles in normal development and maintenance. The sensory and physicochemical characteristics of any protein-rich food contribute to the overall structural behavior of the food” [2]. “Sources of dietary protein can be categorized into functional health-promoting foods Based on their biological characteristics” [3].

Isolates are the most refined form of protein products containing the greatest concentration of protein and concentration contains no dietary fiber. They are very digestible and easily incorporated into different food products. Fish protein isolate is a protein concentrate that is prepared from fish muscle without retaining the original shape of the muscle. It is not generally consumed directly but used as raw material for the production of other value-added products. “Humans are highly dependent on seafood as a source of animal protein. Fishery by-products, which are in huge supply, have received much consideration as a vital protein source as growing interest has been paid to utilizing animal protein as a functional food ingredient” [1].

The pH-shift processing also called acid and/or alkaline solubilization followed by isoelectric precipitation [4], has been successfully recognized as a promising technique to recover direct protein from unconventional complex aquatic raw materials, including gutted fish [5,6] and seafood processing by-products [7,8]. “This

process involves selective isolation of proteins from homogenized raw material using a high (> 10.5) or a low (< 3.5) pH to solubilize the muscle proteins followed by centrifugation to separate the solubilized proteins from high and low-density undissolved material. Then, the recovery of solubilized proteins is done using isoelectric precipitation (usually pH 5.5) and dewatered by centrifugation or filtration. The recovered protein isolate can be mixed with cryoprotectants and then frozen like surimi or minced fish or might be directly dried into a fish protein powder (FPP) for further utilization” Kumarakuru et al. [9].

*Otolithes ruber* commonly known as the tiger tooth croaker, is a fish native to the Indian and Western Pacific Oceans and the Bay of Bengal. It belongs to the family *Sciaenidae* of order Perciformes. In India, It constitutes 10–12% of the demersal catch and is found in the both east and west coasts throughout the year. It is a well-known edible marine fish. Croaker, being a carnivorous Species, diet comprises a wide range of animals, such as crustaceans, polychaetes, mollusks, and small fish. In India, for surimi production, croaker is also one of the major raw materials. Croakers alone contributed 1.36 lakh tons during the 2018-19 marine fish landing. For realizing the conversion of low-value processing discards into high-value byproducts, chemical characterization of croaker discards is important. At present, croaker processing discards are mainly used for the production of fish manure, fish meal, and fish silage. The croaker discards can be used for the recovery of bioactive molecules that are utilized in food, healthcare, pharmaceutical, and nutraceutical industries for improving the economic value of these processing discards as they are one of the important bioresources.

In this study, the alkali solubilization and precipitation technique to isolate proteins from

tiger tooth croaker (*Otolithes ruber*) were used. The proximate composition, peroxide value and functional properties of the protein isolates were also evaluated.

## 2. MATERIALS AND METHODS

### 2.1 Materials

Tiger tooth croaker (*Otolithes ruber*) fish was purchased from the Veraval fish landing center and transported in iced condition (temperature range of 0 to 2°C) to the fish processing laboratory of the College of Fisheries Science, Veraval. It was washed thoroughly in potable chilled water to remove all adhering matters. Proximate analysis was carried out for the raw material. All chemicals and reagents were of analytical grade and were obtained from Central Drug House (CDH) Limited - New Delhi, Ranbaxy laboratories limited - SAS Nagar, Astron Chemical (INDIA), Rankem - New Delhi, Chemdyes Corporation, and Baroda chemical industries (Baroda) limited.

### 2.2 Preparation of Fish Protein Isolates (FPIs)

The extraction of Fish Protein Isolates (FPIs) was done by the method adopted by Hultin and Herbert (2005). Briefly, the fish fillets were ground to mince in a mixer grinder and homogenized with ice-cold deionized water (1:9 ratio) for 3 mins. The pH of the suspension was adjusted to pH 2.5 using 1M HCL, pH 4 using 0.5 N 4C HCL, pH 7 using 0.5 N 4C HCL/NaOH, pH 11.5 using 1N NaOH and pH 12.5 using 1M NaOH. The homogenate was centrifuged at 8000 × g. for 20 mins at 4°C. After centrifugation, three layers were produced; the upper layer and lower layer consist of lipid content and insoluble protein. The middle layer of the supernatant (soluble proteins) was filtered to remove neutral lipids and solid materials, particularly skin, bone and connective tissue. Subsequently, the filtrate pH was adjusted to 5.5 and the filtrate was again centrifuged at 8000 × g. for 15 mins at 4°C. After centrifugation, the obtained supernatant was removed and the precipitate was neutralized, and completely dried in a Hot air oven at 60°C for 24 hours, the product was then ground in powder form, packed and stored at ambient temperature until analysis. The samples were named as protein isolates at pH 2.5 (T1), pH 4 (T2), pH 7 (T3), pH 11.5 (T4) and pH 12.5 (T5).

### 2.3 Proximate Composition

The Proximate composition such as moisture, protein, lipid and ash contents of FPIs, was analyzed using standard AOAC methods (AOAC 1990).

### 2.4 Functional Characteristics

#### 2.4.1 Water-Holding Capacity (WHC)

The water-holding capacity (WHC) of FPIs was analyzed following the procedure of Ozyurt et al. (2015). 2 g of the sample was dispersed in 20 ml of deionized water, and stirred for 20 mins. at 30°C, and centrifuged at 3000 × g. for 15 mins. The WHC is expressed as ml of water absorbed/g of the sample.

#### 2.4.2 Oil-Holding Capacity (OHC)

The oil-holding capacity (OHC) of FPIs was analyzed following the procedure described by Ozyurt et al. (2015). 1 g. of the sample was dispersed in 10 ml of vegetable oil, stirred well for 5 mins. and centrifuged at 3000 × g. for 15 mins. The OHC was displayed as the weight difference.

#### 2.4.3 Emulsifying Capacity (EC)

The emulsifying capacity (EC) of FPIs were determined according to the procedure of Ozyurt et al. (2015). 0.5 g of the sample was added to 50 ml of 0.1 M NaCl and stirred well, and 10 ml of vegetable oil was added. The suspension was homogenized for 5 min, and centrifuged at 5000 × g. for 10 mins. and then poured into a 50 ml graduated measuring cylinder and allowed to stand for few mins until the emulsified layer was stable. The EC was calculated as  $EC (ml/100g.) = (\text{Height of emulsifier layer} / \text{Height of total volume}) * 100$ .

### 2.5 Foam Measurements

The foaming Capacity (FC) and Foam Stability (FS) of FPIs were analyzed according to the method of Foh et al. [2]. 1 g of FPIs was added to 50 ml of distilled water in a 100 ml graduated cylinder. The mixture was stirred for 3 mins. and the generated foam volume was noted and considered as FC. Furthermore, the foam volume noted after 15, 20 and 30 mins. was considered the percentage of FS.

## 2.6 Lipid Oxidation

The Peroxide Value (PV) of lipid was determined from the lipid extract according to Jacobs (1958) odometrically. 10 g. of the sample was taken and ground well with 15 g anhydrous sodium sulphate. Then transferred to a 100 ml stoppered flask and 30-40 ml chloroform was added and placed in a dark place for about 15-20 mins. with occasionally shaking. 10 ml of chloroform extract, and 25 ml of solvent (2 volumes of glacial acetic acid and 35 ml of water) were added. The liberated iodine was titrated against standard sodium thiosulphate solution and explained as milli equivalent of peroxide/ kg of lipid.

## 2.7 Data Analysis

Data was statistically analyzed as per a factorial Completely Randomized Design. Analysis of variance was used to find out significant differences in the sample between the treatments as per the standard statistical methods described by Snedecor & Cochran [10].

## 3. RESULTS AND DISCUSSION

### 3.1 Characteristics of Raw Materials

The physical characteristics and proximate composition of fresh fish is shown are in Table 1. The fresh fish measured  $19.95 \pm 0.86$  cm on average. The standard length of fish was  $17 \pm 0.74$  cm. whereas, the mean weight of fish was  $94.6 \pm 7.22$  g. Similar range of length and weight of tiger tooth croaker (*Otolithes ruber*) was recorded by Vijayakumar et al. [11]. The yield of picked meat was 34% from whole fish.

The fish fillets were used for proximate composition analysis; moisture content was about  $78.02 \pm 1.21\%$ , protein content  $17.75 \pm 0.61\%$ , lipid content  $2.39 \pm 0.06\%$  and ash content was  $1.37 \pm 0.08\%$  respectively. The results of the proximate composition compare well with the results obtained by Zynudheen et al. [12]. The fish meat had a Protein content of 17.36%, lipid of 4.74%, moisture of 77.28% and ash content was found to be 1.14 % respectively.

### 3.2 Characteristics Change in Fish Protein Isolates During the Period of Storage

#### 3.2.1 Changes in proximate composition during the period of storage

**Moisture:** The moisture content in fish protein isolates at different pH (2.5, 4, 7, 11.5 and 12.5) showed increasing trends with increasing storage periods (Table 2). The interaction effect of treatments and storage period (days) was reported to be significant with a CV (%) of 7.537. the initial moisture content of pH 2.5, 4, 7, 11.5 and 12.5 were  $3.17 \pm 0.18$ ,  $3.18 \pm 0.21$ ,  $3.18 \pm 0.17$ ,  $3.02 \pm 0.36$  and  $3.15 \pm 0.19$ . At the end of the storage period moisture was found to be  $3.60 \pm 0.11$ ,  $3.60 \pm 0.14$ ,  $3.59 \pm 0.29$ ,  $3.44 \pm 0.39$  and  $3.55 \pm 0.15$  for pH 2.5, 4, 7, 11.5 and 12.5 respectively (mean  $\pm$  SD). It is due to an increase in relative humidity of more than 70%. In a similar trend, Lone *et al.* (2015) reported a moisture content of RTFPI was 3.5%. Foh *et al.* (2010) reported moisture content of 3.7% for FMMC (Freshly minced meat concentrate) of tilapia fish. The moisture content of FPIC (5.86 g/100 g), FPIIM (5.71 g/100 g), FPIP (5.65 g/ 100 g) and FPIS (6.05 g/100 g) [9].

**Table 1. Characteristics of raw material**

A.	Physical characteristics	Mean $\pm$ S.D.
1	Total Length(cm)	$19.95 \pm 0.86$
2	Standard Length (cm)	$17 \pm 0.74$
3	Weight of Fish (g)	$94.6 \pm 7.22$
4	Yield of picked meat (from whole fish)	34%
B.	Proximate composition	
1	Moisture (%)	$78.02 \pm 1.21$
2	Total Protein (%)	$17.75 \pm 0.61$
3	Total Lipid (%)	$2.39 \pm 0.06$
4	Total Ash (%)	$1.37 \pm 0.08$

**Table 2. Changes in Moisture (%) in Fish Protein Isolate During Storage at Ambient Temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	3.17 ± 0.18	3.18 ± 0.21	3.18 ± 0.17	3.02 ± 0.36	3.15 ± 0.19	3.14
30	3.26 ± 0.14	3.27 ± 0.17	3.26 ± 0.30	3.11 ± 0.42	3.23 ± 0.16	3.22
60	3.34 ± 0.13	3.36 ± 0.16	3.35 ± 0.25	3.20 ± 0.44	3.31 ± 0.13	3.31
90	3.48 ± 0.15	3.48 ± 0.15	3.46 ± 0.27	3.32 ± 0.38	3.42 ± 0.12	3.43
120	3.60 ± 0.11	3.60 ± 0.14	3.59 ± 0.29	3.44 ± 0.39	3.55 ± 0.15	3.55
TX	3.37	3.37	3.36	3.21	3.33	

Each value is represented by dry weight based on the mean ± SD of n=4.

**Table 3. Changes in total protein (%) in fish protein isolate during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	80.90±1.95	82.30±2.20	87.42±0.74	83.15±0.82	84.00±0.95	83.55
30	80.84±1.87	82.24±2.24	87.37±0.76	83.09±0.84	83.94±0.93	83.49
60	80.75±1.92	82.15±2.19	87.28±0.78	83.00±0.86	83.85±0.91	83.40
90	80.65±1.84	82.05±2.16	87.18±0.75	82.90±0.88	83.75±0.97	83.30
120	80.53±1.91	81.93±2.22	87.06±0.71	82.78±0.91	83.63±0.99	83.18
TX	80.73	82.13	87.26	82.98	83.83	

Each value is represented by dry weight based on the mean ± SD of n=4

**Total protein:** The total protein content in fish protein isolates at different pH (2.5, 4, 7, 11.5 and 12.5) showed decreasing trends with increasing storage periods (Table 3). The interaction effect of treatments and storage period (days) was reported to be significant with a CV (%) of 1.766. The initial protein content of fish protein isolates at pH 2.5, 4, 7, 11.5 and 12.5 was found to be 80.90 ± 1.95, 82.30 ± 2.20, 87.42 ± 0.74, 83.15 ± 0.82 and 84.00 ± 0.95. At the end of the storage period total protein was found to be 80.53 ± 1.91, 81.93 ± 2.22, 87.06 ± 0.71, 82.78 ± 0.91 and 83.63 ± 0.99 respectively (mean ± SD). The highest protein value was observed at pH 7 followed by pH 12.5, 11.5, 4 and 2.5. Kumarakuru et al. [9] reported the protein content in FPIP (89.70 g/100g), FPIIM (87.27 g/100 g), FPIC (86.47 g/100 g) and FPIS (84.74 g/100 g). A protein content of 82.39-94.7% has been reported in fish protein isolates (Foh et al., 2010 Liu et al., 2009). The protein content of RTFPI was 75.61% [13]. The decline in protein value in all samples is due to the denaturation of protein during storage at ambient temperature.

**Total Lipid:** The total lipid content in fish protein isolates at different pH (2.5, 4, 7, 11.5 and 12.5) showed decreasing trends with increasing storage periods (Table 4). The interaction effect of treatments and storage period (days) was reported to be significant with CV (%) 6.922. The

initial lipid content of fish protein isolates at pH 2.5, 4, 7, 11.5 and 12.5 was found to be 2.45 ± 0.12, 2.37 ± 0.14, 2.07 ± 0.11, 2.16 ± 0.18 and 2.14 ± 0.19. At the end of the storage period total lipid were found to be 2.09 ± 0.18, 2.01 ± 0.14, 1.71 ± 0.19, 1.80 ± 0.17 and 1.81 ± 0.15 respectively. The lipid content of FMMC of tilapia fish as reported by Foh et al. (2010) was 1.81%. Tongnuanchan et al. [14] reported 0.12% lipid content in fish protein isolate from red tilapia. The lipid content of RTFPI was 2.35% respectively.

**Total Ash:** The total ash content in fish protein isolates at different pH (2.5, 4, 7, 11.5 and 12.5) showed increasing trends with increasing storage periods (Table 5). The interaction effect of treatments and storage period (days) was reported to be not significant with CV (%) 3.434. The initial ash content of fish protein isolates at pH 2.5, 4, 7, 11.5 and 12.5 was found to be 3.57 ± 0.13, 3.59 ± 0.14, 3.45 ± 0.11, 3.52 ± 0.06 and 3.50 ± 0.07. At the end of the storage period total lipids were found to be 3.72 ± 0.21, 3.74 ± 0.15, 3.60 ± 0.14, 3.67 ± 0.09 and 3.65 ± 0.09 respectively.

Lone et al. [13] reported the ash content of RTFPI was 4%. The ash content of FPIIM (0.91 g/100 g), FPIP (0.90 g/100 g), FPIC (0.83 g/100 g) and FPIS (0.88 g/100 g) [9]. The non-significant increase in ash content could be due to the bones present during the mincing operation.

**Table 4. Changes in Total Lipid (%) in Fish Protein Isolate During Storage at Ambient Temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	2.45± 0.12	2.37± 0.14	2.07± 0.11	2.16± 0.18	2.14± 0.19	2.243
30	2.37± 0.15	2.29± 0.13	1.99± 0.15	2.08± 0.15	2.07± 0.21	2.165
60	2.28± 0.13	2.20± 0.15	1.90± 0.13	1.99± 0.21	1.98± 0.17	2.075
90	2.19± 0.17	2.11± 0.19	1.81± 0.17	1.90± 0.19	1.91± 0.18	1.988
120	2.09± 0.18	2.01± 0.14	1.71± 0.19	1.80± 0.17	1.81± 0.15	1.888
TX	2.276	2.201	1.901	1.994	1.986	

Each value is represented by dry weight based on the mean ± SD of n=4

**Table 5. Changes in Ash (%) in fish protein isolate during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	3.57±0.13	3.59±0.14	3.45±0.11	3.52±0.06	3.50±0.07	3.52
30	3.61±0.15	3.63±0.16	3.49±0.13	3.56±0.05	3.54±0.06	3.56
60	3.64±0.17	3.66±0.17	3.52±0.17	3.59±0.08	3.57±0.10	3.59
90	3.69±0.19	3.71±0.19	3.57±0.19	3.64±0.04	3.62±0.12	3.64
120	3.72±0.21	3.74±0.15	3.60±0.14	3.67±0.09	3.65±0.09	3.67
TX	3.64	3.66	3.52	3.59	3.57	

Each value is represented by dry weight based on the mean ± SD of n=4

**Table 6. Changes in peroxide value (mEq/kg) in fish protein isolate during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	3.63±0.18	3.54±0.16	3.41±0.12	3.48±0.14	3.51±0.19	3.51
30	4.09±0.21	4.01±0.13	3.78±0.16	3.87±0.06	3.89±0.06	4.36
60	4.46±0.15	4.42±0.17	4.25±0.19	4.34±0.11	4.36±0.10	3.93
90	4.79±0.13	4.78±0.21	4.68±0.09	4.72±0.03	4.73±0.05	4.74
120	5.36±0.23	5.33±0.25	5.11±0.13	5.24±0.08	5.29±0.13	5.27
TX	4.46	4.41	4.24	4.33	4.35	

Each value is represented by dry weight based on the mean ± SD of n=4

The result indicated that all the parameters were within the prescribed limit signifying the freshness of the fish used in the study.

### 3.2.2 Changes in peroxide value during the period of storage

Lipid oxidation in muscle foods is predominantly detrimental to overall quality and storage stability respectively. Peroxide value (PV) is used to express the oxidative state of lipid-containing foods. It measures the first stage of oxidative rancidity [15]. The effect of different pH on the PV of fish protein isolates is depicted in Table 6 showing increasing trends with increasing storage. At the end of the storage period PV values were found to be 5.36 ± 0.23(meq/kg), 5.33 ± 0.25 (meq/kg), 5.11 ± 0.13 (meq./kg), 5.24 ± 0.08(meq./kg), and 5.29 ± 0.13 (meq./kg) at pH

2.5, 4, 7, 11.5 and 12.5 respectively (mean ± SD). The lowest value was recorded for the pH 7 sample followed by pH 11.5, 12.5, 4 and 2.5 samples. The interaction effect of treatments and storage period (days) was found to be not significant with CV (%) 4.503. The results are in agreement with the work done by Panpipat and Chaijan [16].

### 3.3 Changes in Functional Properties During Storage

#### 3.3.1 Water-holding capacity

Proteins have both hydrophilic and hydrophobic properties therefore it can interact with water and oil in foods [17]. The water-holding capacity of all fish protein isolates samples showed a trend of decreasing during the storage. The interaction

effect of different pH treatments and storage period (days) were found to be significant with CV (%) of 1.204. Initially water-holding capacity at pH 2.5, 4, 7, 11.5 and 12.5 was  $2.18 \pm 0.02$ ,  $2.33 \pm 0.04$ ,  $2.42 \pm 0.03$ ,  $2.45 \pm 0.12$  and  $2.46 \pm 0.11$  (mean  $\pm$  SD). Which changed to  $1.82 \pm 0.17$ ,  $1.98 \pm 0.21$ ,  $2.08 \pm 0.15$ ,  $2.10 \pm 0.23$  and  $2.10 \pm 0.21$  (mean  $\pm$  SD) for pH 2.5, 4, 7, 11.5 and 12.5 respectively on the last day of the storage period of 120 days (Table 7). The WHC is affected by pH and ionic strength. Similar observations were made by Foh *et al.* (2010) while studying FMMC of tilapia fish and Kumarakuru *et al.* [9] while studying functional properties of protein isolates obtained from four fish species.

### 3.4 Oil-Holding Capacity (OHC)

The OHC determines the capacity of food materials to absorb oil. The OHC of proteins is important functional property as it improves the mouthfeel and retains flavor in food. The high oil absorption is essential in the formulation of food systems like sausage, cake batters and mayonnaise and salad dressing [17]. The oil-holding capacity of all fish protein isolates samples showed a trend of decreasing during the storage. The interaction effect of different pH treatments and storage period (days) were found to be significant with a CV (%) of 0.677. Initially oil-holding capacity at pH 2.5, 4, 7, 11.5 and 12.5 was  $1.43 \pm 0.08$ ,  $1.54 \pm 0.01$ ,  $2.11 \pm 0.02$ ,  $2.42 \pm 0.12$  and  $2.48 \pm 0.09$  (mean  $\pm$  SD). Which changed to  $1.08 \pm 0.21$ ,  $1.19 \pm 0.12$ ,  $1.75 \pm 0.01$ ,  $2.07 \pm 0.26$  and  $2.11 \pm 0.23$  (mean  $\pm$  SD) for pH 2.5, 4, 7, 11.5 and 12.5 respectively on the last day of the storage period of 120 days (Table 8). It is because OHC reflects the extent of denaturation of the protein. As the protein denaturation increased during storage the OHC of fish protein isolates decreased. Similar results were also reported by Kumarakuru *et al.* [9]; Elsohaimy *et al.* [18]; Foh *et al.* [2] (2010) with an

OHC range of 5.32 – 5.83 mL/g, 1.88 mL/g, 2.43 mL/g and 3.38 mL/g respectively.

### 3.5 Emulsifying Capacity (EC)

The Emulsifying Capacity (EC) reveals the capacity of a sample to swiftly adsorb at oil/water interfaces during the formation of an emulsion by avoiding flocculation and coalescence. The emulsifying capacity of all fish protein isolates samples showed a trend of decreasing during the storage. The interaction effect of different pH treatments and storage period (days) were found to be significant with CV (%) of 0.128. Initially emulsifying capacity at pH 2.5, 4, 7, 11.5 and 12.5 was  $76.2 \pm 0.12$ ,  $77.5 \pm 0.09$ ,  $78.1 \pm 0.12$ ,  $81.0 \pm 0.11$  and  $81.6 \pm 0.08$  (mean  $\pm$  SD). Which changed to  $75.7 \pm 0.27$ ,  $77.1 \pm 0.29$ ,  $77.6 \pm 0.31$ ,  $80.61 \pm 0.26$  and  $81.14 \pm 0.28$  (mean  $\pm$  SD) for pH 2.5, 4, 7, 11.5 and 12.5 respectively on the last day of the storage period of 120 days (Table 9). EAI (Emulsifying activity index) of RTFPI was 281.0, 207.3 and 535.0 m<sup>2</sup>/g, while the ESI (emulsion stability index) was 11.30, 4.17 and 7.0 min at pH 3, 5 and 7 respectively [13]. Gulzar *et al.* [19] reported the EC of soy protein (52.5 mL/100 g) and marama protein (53.4 mL/100 g) respectively.

### 3.6 Foam Measurements

#### 3.6.1 Foaming Capacity (FC) and Foam Stability (FS)

During protein foaming, the interfacial area that can be produced by a protein is referred to as FC, whereas FS denotes the capability of a protein to stabilize air bubbles against gravitational stress [20]. The foaming capacity of all fish protein isolates samples showed a trend of decreasing during the storage. The interaction effect of different pH treatments and storage period (days) were found to be significant with a

**Table 7. Changes in water holding capacity (mL g<sup>-1</sup>) in fish protein isolates during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	2.18±0.02	2.33±0.04	2.42±0.03	2.45±0.12	2.46±0.11	2.373
30	2.10±0.06	2.25±0.07	2.35±0.05	2.37±0.09	2.38±0.17	2.295
60	2.01±0.09	2.17±0.13	2.26±0.11	2.28±0.04	2.29±0.09	2.206
90	1.92±0.13	2.08±0.17	2.18±0.09	2.20±0.15	2.20±0.05	2.119
120	1.82±0.17	1.98±0.21	2.08±0.15	2.10±0.23	2.10±0.21	2.019
TX	2.009	2.166	2.260	2.287	2.291	

Each value is represented by dry weight based on the mean  $\pm$  SD of n=4

**Table 8. Changes in oil holding capacity (mL g<sup>-1</sup>) in fish protein isolates during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	1.43±0.08	1.54±0.01	2.11±0.02	2.42±0.12	2.48±0.09	2.001
30	1.35±0.02	1.45±0.05	2.02±0.07	2.33±0.05	2.39±0.13	1.913
60	1.27±0.11	1.38±0.09	1.94±0.13	2.25±0.09	2.30±0.19	1.832
90	1.18±0.17	1.29±0.03	1.85±0.21	2.17±0.21	2.21±0.31	1.745
120	1.08±0.21	1.19±0.12	1.75±0.25	2.07±0.26	2.11±0.23	1.645
TX	1.268	1.375	1.940	2.253	2.298	

Each value is represented by dry weight based on the mean ± SD of n=4

**Table 9. Changes in emulsifying capacity (mL /100 g) in fish protein isolates during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	76.2±0.12	77.5±0.09	78.1±0.12	81.0±0.11	81.6±0.08	78.92
30	76.1±0.17	77.4±0.03	78.0±0.09	80.9±0.21	81.4±0.15	78.81
60	76.0±0.19	77.3±0.14	77.9±0.21	80.8±0.16	81.3±0.19	78.70
90	75.9±0.21	77.2±0.18	77.7±0.28	80.7±0.19	81.2±0.24	78.58
120	75.7±0.27	77.1±0.29	77.6±0.31	80.6±0.26	81.1±0.28	78.46
TX	76.02	77.34	77.89	80.84	81.37	

Each value is represented by dry weight based on the mean ± SD of n=4

**Table 10. Changes in foaming capacity (mL /100 g) in fish protein isolates during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	41.5±0.12	42.9±0.09	45.5±0.13	47.8±0.10	48.5±0.17	45.26
30	41.4±0.17	42.8±0.13	45.4±0.20	47.7±0.16	48.4±0.13	45.20
60	41.3±0.23	42.7±0.08	45.3±0.27	47.6±0.09	48.3±0.21	45.09
90	41.2±0.14	42.6±0.11	45.2±0.15	47.5±0.21	48.2±0.27	44.99
120	40.9±0.39	42.5±0.19	45.1±0.19	47.5±0.27	48.1±0.34	44.85
TX	41.29	42.74	45.36	47.66	48.34	

Each value is represented by dry weight based on the mean ± SD of n=4

CV (%) of 0.291. the highest foaming capacity was at pH 12.5 followed by pH 11.5, 7, 4 and 2.5. Initially foaming capacity at pH 2.5, 4, 7, 11.5 and 12.5 was 41.5 ± 0.12, 42.9 ± 0.09, 45.5 ± 0.13, 47.8 ± 0.10 and 48.5 ± 0.17 (mean ± SD). Which changed to 40.9 ± 0.39, 42.5 ± 0.19, 45.1 ± 0.19, 47.5 ± 0.27 and 48.1 ± 0.34 (mean ± SD) for pH 2.5, 4, 7, 11.5 and 12.5 respectively on the last day of the storage period of 120 days (Table 10) [21-23].

The foam stability of all fish protein isolates samples showed a trend of rapidly decreasing during the storage. The interaction effect of different pH treatments and storage period (days) were found to be significant with CV (%) of 0.549. Initially, foam stability at pH 2.5, 4, 7, 11.5 and 12.5 was 26.3 ± 0.15, 27.3 ± 0.21, 28.1 ± 0.27, 29.2 ± 0.31 and 29.5 ± 0.17 (mean ± SD).

Which changed to 13.9 ± 0.23, 14.7 ± 0.29, 15.5 ± 0.21, 16.9 ± 0.19 and 17.3 ± 0.31 (mean ± SD) for pH 2.5, 4, 7, 11.5 and 12.5 respectively on the last day of the storage period of 120 days (Table 11). The highest foam stability was at pH 12.5 followed by pH 11.5, 7, 4 and 2.5 [24-26].

Kumarakuru et al. [9] reported the FC of FPIC, FPIIM, FPIP and FPIS was up to 46.2 mL/100 g, 45.2 mL/100 g, 47.7 mL/100 g and 44.2 mL/100 g respectively, and FS of FPIC, FPIIM, FPIP and FPIS was ranged between 30.5 and 34.0 mL/100 g, 20.3 and 22.5 mL/100 g and 14.8 and 17.1 mL/100 g for foam intervals at 15, 20 and 30 mins respectively, with a significant difference ( $p < 0.05$ ). The FC value of quinoa protein was 58.37 mL/100 mL [18]. The FS of FMMC of tilapia fish ranged from 90.17 to 52.63% as reported by Foh et al. (2010).



**Table 11. Changes in foam stability (mL /100 g) in fish protein isolates during storage at ambient temperature**

Storage Period (Days)	Treatments					DX
	T1	T2	T3	T4	T5	
0	26.3±0.15	27.3±0.21	28.1±0.27	29.2±0.31	29.5±0.17	28.12
30	23.3±0.04	24.1±0.15	24.9±0.11	26.3±0.27	26.7±0.13	25.11
60	20.2±0.11	21.0±0.19	21.8±0.18	23.2±0.09	23.6±0.21	22.01
90	17.1±0.03	17.9±0.26	18.6±0.05	20.1±0.16	20.5±0.27	18.88
120	13.9±0.23	14.7±0.29	15.5±0.21	16.9±0.19	17.3±0.31	22.06
TX	21.466	22.302	23.091	24.507	24.836	

Each value is represented by dry weight based on the mean  $\pm$  SD of n=4

#### 4. CONCLUSIONS

The fish protein isolates are the most refined form of protein products containing the greatest concentration of protein. This study demonstrated that acid or alkali-aided processing and isoelectric precipitation can be successfully used to extraction of protein isolates from tiger tooth croaker (*Otolithes ruber*) fish. Protein recovery was highest for the alkali-aided method specifically at pH 7. The results revealed that the alkali-aided method exhibited more favourable functional properties than the acid-aided method. Low lipid oxidation of protein isolates prepared through the pH-shift process replicates their functional characteristics. Therefore, the pH-shift process can be used as a powerful tool to recover functional proteins from tiger tooth croaker (*Otolithes ruber*).

Hence the study proved the alkali extraction method in the isolation of fish protein isolates with favourable functional characteristics to be applicable in the development of protein rich food products satisfying the present need for isolation of functional nutrients in the area of functional food.

#### CONFERENCE DISCLAIMER

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#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

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