# Advancements in Breeding Strategies for Enhancing Wilt and Root Rot Resistance in Castor (*Ricinus communis* L.): A Comprehensive Review

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*Review Article*

## ABSTRACT

Castor (*Ricinus communis* L.) is an economically important oilseed crop cultivated worldwide for its versatile uses in various industries. However, the cultivation of castor is challenged by wilt and root rot diseases caused by fungal pathogens, leading to significant yield losses. Enhancing wilt and root rot resistance in castor is crucial for sustainable and productive cultivation. This review explores advancements in breeding strategies for enhancing resistance against these diseases through various approaches along with  introduction to castor cultivation and the challenges posed by wilt and root rot diseases in castor. It emphasizes the importance of breeding for disease

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resistance and discusses the significance of genetic diversity in castor germplasm. Traditional breeding approaches including phenotypic selection, recurrent selection, hybridization, mutation breeding and polyploidy induction are examined for their effectiveness in introducing and accumulating resistance genes and traits. Biotechnological approaches such as marker-assisted selection (MAS), genetic transformation and RNA interference (RNAi) are highlighted as powerful tools for targeted selection and manipulation of specific genes associated with disease resistance. The utilization of physiological and biochemical approaches including understanding host-pathogen interactions, elicitor-induced defense responses and metabolic engineering are discussed for enhancing resistance mechanisms in castor. These advancements offer promising avenues for accelerating the development of wilt and root rot-resistant castor varieties.

*Keywords: Castor; wilt; root rot; marker-assisted selection; transcriptomics.*

## 1. INTRODUCTION

Castor (*Ricinus communis* L.) is an important oilseed crop cultivated worldwide for its valuable oil and versatile applications in various industries, including pharmaceuticals, cosmetics, and biofuel production [1]. However, the cultivation of castor is significantly hampered by wilt and root rot diseases caused by fungal pathogens, such as *Fusarium oxysporum* and *Macrophomina phaseolina.* These diseases result in substantial yield losses and pose a major challenge to castor growers [2]. Enhancing wilt and root rot resistance in castor is of paramount importance to ensure sustainable and productive cultivation [3]. Breeders and researchers have focused their efforts on developing resistant castor varieties through various breeding strategies [4]. Traditional breeding approaches, such as phenotypic selection, hybridization, and recurrent selection, have been successfully employed to introduce and accumulate resistance genes and traits within the castor germplasm [5]. These methods rely on the evaluation of plant phenotypes and the selection of individuals displaying resistance to wilt and root rot diseases. In recent years, biotechnological approaches have revolutionized plant breeding and provided powerful tools to enhance disease resistance [6]. Marker-assisted selection (MAS) enables breeders to select plants carrying specific genes associated with resistance, accelerating the breeding process [7]. Genetic transformation techniques, such as the introduction of foreign genes into castor plants, offer targeted methods to enhance resistance by conferring novel genes or traits [8]. RNA interference (RNAi) technology allows for gene silencing and regulation, enabling the suppression of essential genes in the pathogens, thus inhibiting their growth and virulence [9]. Physiological and biochemical approaches have contributed to a better understanding of the host-

pathogen interactions and defense responses in castor [10]. The elucidation of molecular and biochemical mechanisms involved in disease resistance has opened up opportunities for targeted interventions, such as the application of elicitors to stimulate the plant's defense responses or metabolic engineering to enhance the production of defense-related compounds [11].

## 2. GENETIC DIVERSITY AND DISEASE RESISTANCE

### 2.1 Genetic Diversity in Castor Germplasm

Genetic diversity is a fundamental aspect of plant breeding and plays a vital role in the development of disease-resistant castor varieties [12]. Castor germplasm encompasses a wide range of genetic variability, making it a valuable resource for breeders seeking to improve resistance against wilt and root rot pathogens [13]. Germplasm collections, such as gene-banks, house diverse castor accessions from various geographical regions [14]. These accessions represent different ecotypes, landraces, and wild relatives, each harboring distinct genetic traits, including disease resistance [15]. The genetic diversity within castor germplasm can be attributed to natural genetic variation, historical selection pressures, and human intervention. Assessing and characterizing the genetic diversity in castor germplasm is crucial for identifying and selecting resistant sources for breeding programs [16]. Researchers utilize various techniques to study genetic diversity, such as molecular markers, genotyping platforms, and sequencing technologies. These approaches enable the analysis of DNA polymorphisms, such as single nucleotide polymorphisms (SNPs) and simple sequence repeats (SSRs), providing insights into

the extent and patterns of genetic variation within the germplasm [17]. Studies have revealed substantial genetic diversity among castor accessions in terms of disease resistance [18]. Different accessions exhibit varying levels of resistance to wilt and root rot pathogens, indicating the presence of genes and traits conferring resistance within the germplasm [19]. Systematically screening and evaluating castor accessions for their response to disease infection, breeders can identify resistant sources that can serve as valuable genetic resources for breeding programs [20]. The identification of genetically diverse castor accessions with resistance traits allows for the creation of diverse breeding populations. By crossing different resistant accessions, breeders can generate populations with increased genetic variability, facilitating the selection of superior progeny with enhanced resistance. The use of diverse parental lines is particularly important to avoid the risk of vulnerability to a specific pathogen race or strain [21].

## 2.2 Identification of Resistant Sources and Screening Techniques

Identifying resistant sources within the castor germplasm is a critical step in breeding for disease resistance. It involves screening a diverse range of accessions or genotypes to identify individuals with a high level of resistance against wilt and root rot diseases [22].

Field evaluations are commonly employed to assess the performance of castor accessions under natural infection conditions [23]. Castor plants are grown in disease-prone areas, and the incidence and severity of wilt and root rot symptoms are monitored [24]. Accessions that display a reduced incidence of disease symptoms or a delayed progression of the disease are identified as potential resistant sources [25]. Field evaluations provide valuable insights into the performance of accessions in their natural environment and under realistic

**Table 1. Screening of germplasm for resistance to wilt**

| Disease reaction | Range of PDI (%) | Germplasm accessions |
|---|---|---|
| Highly resistant | 0.0 | Nil |
| Resistant | 0.1 – 20.0 | RG844,1146,1221,1577,1697,1766,2100,2720,2759, 2924, 3225, 3242, 3253, 3292, 3296, 3336,3338, 3352,3352, 3359, 3361, 3378, 3383, 3386, 1714, 2093, 2145, 2161 and 2254 |
| Moderately resistant | 20.1 – 40.0 | RG-1180,1305, 1357, 1621, 1718, 1788, 2082, 2098, 2121, 2155, 2173, 2184, 2860, 3312, 3315, 3330, 3332, 3368, 3378, 3375 and 3390 |
| Moderately susceptible | 40.1 – 50.0 | RG-133, 1582, 1612, 1685, 1709, 1904, 1999, 2076, 2077, 2081, 2111, 2112, 2116, 2137, 2140, 2148,2191, 2253, 2614, 2697, 2761, 2773, 2870, 3314,3328, 3357, 3371, 3380 and 3382 |
| Susceptible | 50.1 – 75.0 | RG-62, 190, 1103, 1125, 1142,1313, 1340, 1353, 1414, 1545, 1548, 1696, 1721, 1772, 1937, 2005,2035, 2062, 2071, 2072, 2073, 2075, 2097, 2099, 2102, 2110, 219, 2129, 2131, 2132, 2138, 2142,2150, 2166, 2171, 2240, 2242, 2243, 2250, 2334, 2422, 2451, 2486, 2681, 2685, 2769, 2775, 2776,2804, 2822, 2833, 3061, 3102, 3116, 3177, 3187, 3188, 3206, 3218, 3223, 3233, 3262, 3297, 3298,3304, 3307, 3309, 3311, 3320, 3322,3325, 3329, 3331, 3334, 3335, 3339, 3340, 3342, 3344, 3346,3347, 3348, 3350, 3351, 3352, 3355, 3364, 3366, 3367, 3370, 3372, 3377 and 3387 |
| Highly susceptible | >75.0 | RG-755, 1139, 1148, 1413, 1523, 1526, 1546, 1916, 1986, 2011, 2064, 2080, 2091, 2118, 2378, 2783, 3240, 3251, 3294, 3302, 3313, 3337, 3341, 3343, 3345, 3349 and 3363 |

*Source: Priya et al. [97]*

**Table 2. Source of resistance to root rot**

| Disease | Resistance source | |
|---|---|---|
| | **Germplasm** | **Breeding lines** |
| Root rot / Charcoal rot | RG-43, 109, 111, 293, 297, 392, 724, 1607, 1624, 1628, 2529, 2706, 2710, 2719, 2722, 2726, 2752, 2758, 2779, 2787, 2799, 2809, 2813, 2816, 2818, 2819, 2822, 2824, 2829, 2847, 2987 | SHB-808, Geeta, SKI-299, JP-89, SKI-283, 304, SKP-217, SHB-825, JP-89xPCS-124, SKP-42, JI-220, 259, 315, 342, JP-93 |

*Source: Anonymous [96]*

disease pressure [26]. Greenhouse assays provide controlled environments to assess the response of castor accessions to wilt and root rot pathogens [27,28]. Plants are grown under controlled conditions, and the pathogens are artificially introduced through inoculation. Symptoms, such as wilting, stunting, and root rot, are evaluated over a specified period. Comparing the responses of different accessions, breeders can identify individuals displaying a high level of resistance [29]. Greenhouse assays allow for the rapid screening of a large number of accessions in a controlled setting, enabling the identification of resistant sources with potential for further evaluation [28].

Laboratory-based screening methods involve the isolation and cultivation of pathogens in a controlled laboratory environment. The pathogens are then used to inoculate castor plants or plant tissues under controlled conditions [30]. This allows for the evaluation of disease symptoms and the quantification of pathogen growth or colonization. Laboratory-based screening methods can provide insights into the underlying mechanisms of resistance, such as pathogen suppression or inhibition of pathogen growth within the plant tissues [31].

In addition to screening techniques, molecular markers linked to disease resistance genes or genomic regions can aid in the identification of resistant sources. Genetic markers associated with resistance can be used to screen large populations or germplasm collections rapidly. Marker-assisted selection (MAS) allows breeders to select individuals carrying resistance alleles based on the presence of specific molecular markers [32]. The identification of resistant sources using these screening techniques and molecular markers enables breeders to select suitable parents for developing resistant cultivars through traditional breeding approaches or biotechnological methods. It provides a valuable resource for introducing and enhancing resistance traits in castor breeding programs.

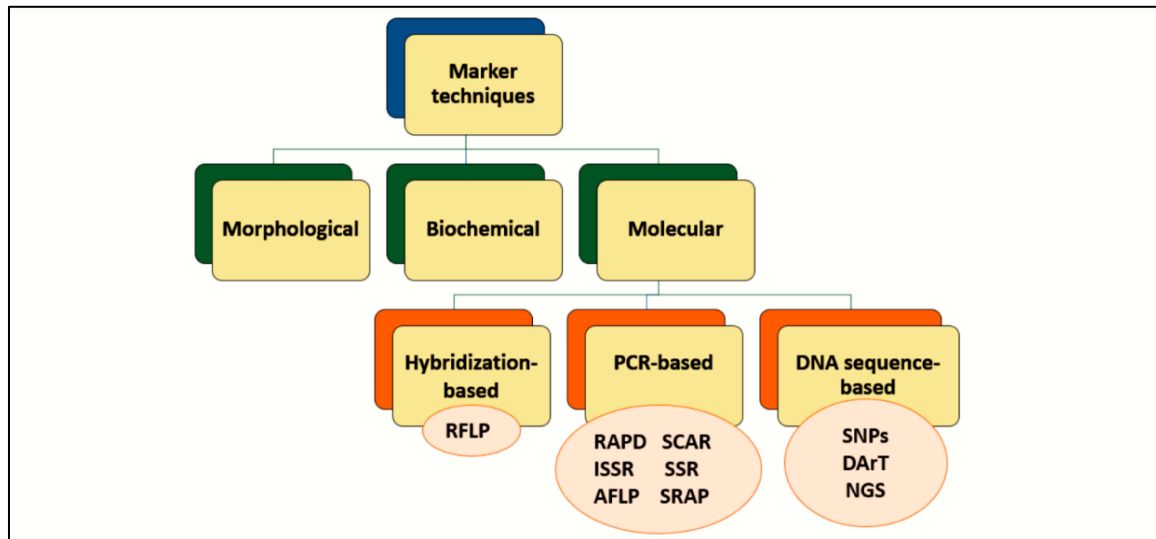## 2.3 Molecular Markers for Disease Resistance

Molecular markers play a crucial role in plant breeding by assisting in the identification and tracking of genes or genomic regions associated with disease resistance [33]. Various molecular marker techniques, such as amplified fragment length polymorphism (AFLP), simple sequence repeats (SSRs), and single nucleotide polymorphisms (SNPs), have been employed to assess the genetic diversity and identify markers linked to wilt and root rot resistance in castor [34]. These markers can facilitate marker-assisted selection (MAS) and accelerate the breeding process by enabling the selection of plants carrying desired resistance traits [35].

## 3. TRADITIONAL BREEDING APPROACHES

Traditional breeding approaches have played a significant role in enhancing disease resistance in castor; including resistance to wilt and root rot diseases [36]. These methods rely on the selection and mating of plants based on their phenotypic characteristics and the desired traits.

## 3.1 Phenotypic Selection and Recurrent Selection

Phenotypic selection is a commonly used traditional breeding approach. Breeders visually evaluate the plants for disease symptoms and select individuals with desirable traits, such as resistance to wilt and root rot diseases. Phenotypic selection requires careful observation and assessment of plant performance in field or greenhouse environments. Resistant individuals showing minimal disease symptoms or delayed disease progression are chosen as parents for the next generation [37].

**Fig. 1. Molecular markers use as disease resistance**
*Source: Guler and Imamoglu [34]*

Recurrent selection is a form of phenotypic selection that involves repeated cycles of selection and mating. Resistant plants are repeatedly crossed with susceptible plants to generate populations with a higher frequency of resistance genes. This approach helps in accumulating favorable genes and traits for disease resistance over multiple breeding cycles. Recurrent selection allows for the gradual improvement of resistance levels in the population and the development of more resilient castor varieties [38].

Hybridization is another traditional breeding approach used to introduce genetic variability and create new combinations of genes. Breeders' cross different castor varieties or genotypes with complementary traits, including resistance to wilt and root rot diseases. The resulting hybrids can exhibit enhanced resistance compared to their parental lines. Hybridization allows for the combination of favorable traits from different parents, potentially leading to improved resistance against specific pathogens [39].

Mutation breeding is a traditional approach that involves the induction of random genetic mutations using physical or chemical mutagens. Breeders treat castor seeds or plants with mutagens to create a diverse population with a wide range of genetic variations. Screening the mutated population for disease resistance helps identify individuals with improved resistance to wilt and root rot pathogens. These resistant mutants can be selected and further bred to develop new castor varieties with enhanced disease resistance [40].

Polyploidy induction is another traditional breeding technique used to increase genetic variation. Polyploidy refers to the multiplication of the chromosome sets within a plant. Inducing polyploidy in castor plants can result in changes in plant physiology and morphology, including enhanced disease resistance. Polyploid individuals may exhibit altered traits that contribute to resistance against wilt and root rot diseases [41].

## 3.2 Hybridization and Selection for Disease Resistance

Hybridization is a fundamental traditional breeding approach that enables the combination of desirable traits from different castor varieties or genotypes, including resistance to wilt and root rot diseases. The process involves crossing two or more parental lines that possess complementary traits, such as resistance to different pathogens or different mechanisms of resistance [42]. Breeders carefully select the parental lines based on their phenotypic characteristics, including disease resistance, growth habits, yield potential, and other agronomic traits. The selected parents should possess contrasting but complementary traits to maximize the chances of obtaining improved resistance in the offspring [43]. The cross between the selected parents leads to the development of a population known as the F1 generation. The F1 generation represents the

first generation of hybrids and exhibits a combination of traits inherited from both parents. These hybrids often display increased vigor and heterosis (hybrid vigor), which can contribute to improved resistance to wilt and root rot diseases [44]. The F1 hybrids are then subjected to rigorous evaluation and selection for disease resistance. This process involves exposing the hybrid plants to the pathogens causing wilt and root rot diseases and assessing their response. The evaluation can be conducted in the field, greenhouse, or controlled laboratory conditions, depending on the specific requirements of the pathogens and the breeding objectives [45]. The selection process where breeders can identify the individuals with enhanced resistance which can manifest as reduced disease symptoms, delayed disease progression or a lower pathogen load. The selected hybrids exhibiting the desired resistance traits are further propagated through successive generations to stabilize and fix the resistance in the population [46].

### 3.3 Mutation Breeding and Polyploidy Induction

Mutation breeding is a traditional breeding approach used to introduce genetic variation by inducing mutations in the plant genome. Chemical mutagens, such as ethyl methanesulfonate (EMS) or sodium azide, or ionizing radiation, such as gamma rays or X-rays, are commonly employed to induce mutations in castor plants. These mutagens cause random changes in the DNA sequence, leading to genetic variations in the population [47]. After mutagenesis, mutagenized populations are screened for disease resistance against wilt and root rot pathogens. The mutagenized plants are subjected to disease inoculation, and individuals displaying improved resistance are identified and selected for further breeding [48]. The selection process focuses on identifying individuals with reduced disease symptoms, delayed disease progression, or lower pathogen loads compared to the non-mutagenized population. The selected resistant mutants can be used as parents in subsequent breeding programs to develop castor varieties with enhanced resistance [49].

Polyploidy induction involves manipulating the chromosome number in plants. Polyploids are organisms with more than two sets of chromosomes. Polyploidy can occur naturally through mechanisms such as unreduced gamete formation or can be induced through chemical treatments, such as colchicine. Polyploid castor plants possess more genetic material, which can result in altered traits, including enhanced disease resistance [50]. Polyploid populations are evaluated for disease resistance against wilt and root rot pathogens. The resistance levels of the polyploid individuals are compared to the diploid (normal chromosome number) individuals to assess any improvements in resistance. The polyploid individuals displaying enhanced resistance are selected for further breeding to develop new castor varieties with improved disease resistance [51].

## 4. BIOTECHNOLOGICAL APPROACHES

Biotechnological approaches have revolutionized plant breeding by providing powerful tools for the manipulation and enhancement of specific traits, including disease resistance [52,53]. In the context of castor breeding for wilt and root rot resistance, several biotechnological techniques have been employed with promising results explained in subheadings below.

### 4.1 Marker-Assisted Selection (MAS) for Disease Resistance Genes

Marker-assisted selection (MAS) has revolutionized plant breeding by providing a powerful tool for the targeted selection of individuals carrying specific genes associated with disease resistance. In the case of castor breeding for wilt and root rot resistance, MAS has been successfully employed to identify and select resistance genes against these pathogens [54].

The first step in MAS is the identification and validation of molecular markers closely linked to the target resistance genes. These markers can be DNA-based, such as simple sequence repeats (SSRs), single nucleotide polymorphisms (SNPs), or insertion-deletion (InDel) markers, or they can be derived from other sources, such as expressed sequence tags (ESTs) or amplified fragment length polymorphisms (AFLPs). The markers are selected based on their genetic linkage to the resistance genes, ensuring that they co-segregate with the target trait in the population [55]. Once the markers are validated and linked to the resistance genes, genotyping is performed on a large population or germplasm collection. This allows for the identification of individuals that carry the markers associated with resistance. The genotyping can be done using various techniques, including polymerase chain

reaction (PCR), high-throughput genotyping platforms, or sequencing-based approaches. The genotyping data are then analyzed to identify individuals with the desired resistance gene(s) [56]. The individuals identified as carrying the resistance genes can be selected for further breeding programs. These individuals serve as valuable parents for developing new castor varieties with enhanced resistance to wilt and root rot diseases. By directly selecting plants based on their genotype, MAS expedites the breeding process by eliminating the need for time-consuming and labor-intensive phenotypic evaluations. It also enables the selection of individuals at an early stage, reducing the cost and time required for field evaluations. MAS offers several advantages in castor breeding for disease resistance [57]. It allows for the precise and efficient selection of individuals carrying the target resistance genes, resulting in a higher success rate in breeding for resistance. MAS also enables the combination of multiple resistance genes through the simultaneous genotyping of multiple markers. This facilitates the pyramiding of resistance genes, enhancing the durability and effectiveness of disease resistance in castor varieties. MAS can be combined with other breeding approaches, such as hybridization or recurrent selection, to further enhance disease resistance in castor populations [58]. By incorporating MAS into the breeding process, breeders can accelerate the development of resistant castor varieties and contribute to sustainable and productive cultivation.

## 4.2 Genetic Transformation and Transgenic Approaches

Genetic transformation and transgenic approaches have played a significant role in enhancing disease resistance in castor, including resistance to wilt and root rot pathogens. These techniques involve the introduction of foreign genes into the castor genome, providing a targeted and efficient method to enhance resistance traits. Agrobacterium-mediated transformation is one of the commonly used methods for genetic transformation in castor. In this approach, the desired resistance gene(s) are inserted into a transfer DNA (T-DNA) vector, which is then introduced into Agrobacterium tumefaciens [59]. The transformed Agrobacterium is then used to infect castor explants, such as leaf or cotyledon segments. The T-DNA containing the resistance gene(s) is transferred from the Agrobacterium into the plant cells, integrating into the castor genome. Biolistics, or particle bombardment, is another technique employed for genetic transformation in castor. In this approach, the resistance gene(s) are coated onto tiny gold or tungsten particles, which are then propelled into the target plant cells using a particle gun or gene gun [60]. The foreign gene(s) are delivered into the plant cells, where they can integrate into the castor genome and confer the desired resistance trait. The introduced resistance genes in transgenic castor plants can originate from various sources, including other plant species that exhibit resistance to wilt and root rot pathogens. These genes can encode antifungal proteins, such as chitinases, glucanases, or defensins, which directly inhibit the growth or infectivity of the pathogens [61]. Other genes may encode pathogen recognition receptors, such as receptor-like kinases (RLKs) or receptor-like proteins (RLPs), which activate defense responses upon pathogen detection. Enzymes involved in defense pathways, such as those involved in the production of phytoalexins or antimicrobial compounds, have also been introduced into castor to enhance resistance. After successful genetic transformation, transgenic castor plants carrying the resistance genes are subjected to rigorous evaluation and screening for their response to wilt and root rot pathogens [62]. It is important to consider the potential challenges and regulatory requirements associated with genetically modified organisms (GMOs) when employing transgenic approaches in castor breeding. Compliance with biosafety regulations and public acceptance of genetically modified castor varieties are critical considerations in the application of transgenic approaches [63].

## 4.3 RNA Interference (RNAi) Technology

RNA interference (RNAi) technology has emerged as a powerful approach for enhancing disease resistance in castor, including resistance to wilt and root rot pathogens. RNAi enables the silencing or downregulation of specific genes in the pathogens, effectively inhibiting their growth, reproduction, and virulence. The RNAi process involves the introduction of double-stranded RNA (dsRNA) molecules that are complementary to the target genes in the pathogens. These dsRNA molecules can be introduced into the castor plants through genetic transformation or other delivery methods [64]. Once inside the plant cells, the dsRNA is processed into small interfering RNAs (siRNAs) by cellular machinery.

The siRNAs act as guides and specifically bind to the complementary RNA sequences of the target genes in the pathogens. This triggers a cascade of molecular events that leads to the degradation or inhibition of the target gene transcripts. As a result, the expression of essential genes in the pathogens is suppressed, hindering their ability to cause disease in the castor plants. RNAi-mediated resistance has shown promising results in protecting castor plants against wilt and root rot diseases [65]. Specifically targeting genes involved in pathogen virulence, reproduction, or infection, RNAi can significantly reduce the severity and incidence of disease symptoms. For example, genes encoding enzymes essential for pathogen cell wall degradation, toxin production, or virulence factor synthesis can be targeted using RNAi, impairing the pathogen's ability to cause disease. RNAi-mediated resistance in castor can be achieved by introducing the dsRNA molecules targeting the pathogen genes through genetic transformation or other delivery methods [66]. Transgenic castor plants expressing the dsRNA can pass on the resistance trait to subsequent generations. Alternatively, RNAi-inducing molecules, such as synthetic dsRNA or small RNA molecules, can be applied to the plants through foliar sprays or root treatments to trigger the RNAi response against the pathogens. The integration of RNAi technology with other breeding strategies, such as marker-assisted selection and genetic transformation, can further enhance the development of wilt and root rot-resistant castor varieties [67]. Combining multiple approaches, breeders can harness the power of RNAi to silence crucial genes in the pathogens while utilizing other methods to introduce and accumulate resistance genes and traits in castor germplasm. It is worth noting that the successful application of RNAi-mediated resistance requires a deep understanding of the target pathogen's genetic makeup and the specific genes involved in disease development [68]. It also necessitates the identification and selection of effective target genes and the optimization of delivery methods to ensure efficient and consistent gene silencing.

## 5. GENOMIC APPROACHES

Genomic approaches have emerged as powerful tools in plant breeding, including the enhancement of wilt and root rot resistance in castor. These approaches involve the utilization of genomic information to understand the genetic basis of disease resistance, identify candidate genes, and accelerate the breeding process [53,69].

## 5.1 Genome Sequencing and Assembly

Advancements in next-generation sequencing technologies have enabled the sequencing and assembly of the castor genome [70]. The availability of a high-quality reference genome provides a foundation for studying the genetic makeup of castor and identifying genes associated with disease resistance [71]. The genome sequence can facilitate the identification of candidate genes involved in defense responses and contribute to the development of molecular markers for marker-assisted selection [72].

## 5.2 Quantitative Trait Loci (QTL) Mapping

Quantitative trait loci (QTL) mapping is a powerful genetic approach used to identify genomic regions associated with the variation of a quantitative trait, such as disease resistance in castor. It involves the analysis of genetic segregation patterns in controlled crosses and the identification of genetic markers linked to the target trait. In the context of enhancing wilt and root rot resistance in castor, QTL mapping is conducted by crossing two castor lines or genotypes that exhibit contrasting levels of resistance [68]. The resulting mapping population, usually comprised of multiple generations, is phenotypically evaluated for disease resistance. Phenotypic data, such as disease severity scores or pathogen load measurements, are collected for each individual in the population. The mapping population is genotyped using molecular markers distributed throughout the genome. These markers can include simple sequence repeats (SSRs), single nucleotide polymorphisms (SNPs), or other DNA-based markers. The genotyping data and phenotypic data are then analyzed together to identify associations between specific genomic regions and the resistance phenotype. Statistical methods, such as linkage analysis or association mapping, are employed to identify significant marker-trait associations [73]. These associations indicate the presence of QTLs linked to the resistance trait. QTLs are specific regions in the genome that contribute to the variation of the trait of interest. By analyzing the co-segregation patterns of genetic markers and the resistance phenotype, breeders can identify and map QTLs associated with wilt and root rot resistance [74].

The identified QTLs and associated markers can then be used for marker-assisted selection (MAS) in breeding programs. The markers linked to the resistance QTLs can be used to screen larger populations or germplasm collections to identify individuals carrying the resistance alleles. By selecting individuals with the favorable markers, breeders can increase the efficiency of selecting for wilt and root rot resistance during the breeding process [75]. QTL mapping can facilitate the introgression of resistance alleles into elite castor varieties. Crossing resistant individuals from the mapping population with elite breeding lines, breeders can transfer the resistance QTLs into improved genetic backgrounds. This process involves multiple rounds of backcrossing and selection to introgress the resistance alleles while retaining desirable agronomic traits from the elite parent [76].

## 6. PHYSIOLOGICAL AND BIOCHEMICAL APPROACHES

Physiological and biochemical approaches play a crucial role in enhancing disease resistance in castor plants by uncovering the underlying mechanisms of host-pathogen interactions and identifying the defense responses of castor against wilt and root rot diseases. These approaches provide insights into the physiological and biochemical processes involved in disease resistance, enabling the manipulation of these processes to enhance resistance.

Physiological approaches involve the study of various physiological parameters and responses of castor plants during pathogen infection. Researchers analyze traits such as water relations, photosynthetic efficiency, stomatal conductance, hormone levels, and nutrient uptake to understand how pathogens affect the physiological status of the plant [77]. By elucidating the impact of pathogens on these physiological processes, strategies can be developed to enhance the plant's resilience against wilt and root rot diseases. For instance, optimizing irrigation practices to maintain optimal water relations and nutrient management to improve plant nutrition can help strengthen the plant's defense mechanisms [78].

Biochemical approaches focus on the study of the biochemical pathways and defense compounds involved in disease resistance. Researchers investigate the production of phytoalexins, antimicrobial compounds, and signaling molecules involved in defense signaling pathways [79]. They also analyze the activity of enzymes such as chitinases, glucanases, peroxidases, and phenylalanine ammonia-lyase (PAL) involved in the synthesis of defense compounds. Understanding the biochemical processes and the regulation of defense compounds allows researchers to manipulate these pathways to enhance resistance. For example, the application of exogenous defense compounds or the overexpression of genes involved in defense pathways can bolster the plant's ability to combat pathogens [80].

## 6.1 Understanding Host-pathogen Interactions

Studying the interactions between castor plants and wilt and root rot pathogens is crucial for developing effective strategies for disease management. Elucidating the molecular and biochemical mechanisms involved in the infection process, researchers can identify potential targets for enhancing resistance [11,81]. This understanding allows for the development of targeted interventions to disrupt the pathogen's infection strategies or strengthen the plant's defense mechanisms.

## 6.2 Induction of Resistance Mechanisms through Elicitors

Induction of resistance mechanisms through elicitors is a promising approach for enhancing disease resistance in castor plants. Elicitors are molecules that can activate the plant's defense responses, thereby priming the plant to effectively combat pathogen attacks. By applying specific elicitors or elicitor-inducing compounds, researchers can trigger a cascade of biochemical and molecular events that lead to the production of defense-related compounds [82]. Various types of elicitors have been studied for their ability to induce resistance in castor against wilt and root rot pathogens. These elicitors can include natural compounds derived from pathogens, such as fungal cell wall components (e.g., chitin, β-glucans) or pathogen-derived molecules (e.g., elicitor proteins), as well as synthetic compounds that mimic pathogen signals (e.g., synthetic elicitors or analogs). When elicitors are applied to castor plants, they are recognized by specific receptors or sensors on the plant's cell surface or within the cell [83]. This recognition triggers a signaling cascade that leads to the activation of defense-related genes

and the subsequent synthesis and accumulation of defense compounds. One of the key defense compounds induced by elicitors is phytoalexins, which are antimicrobial metabolites produced by plants in response to pathogen attack. Phytoalexins have direct antimicrobial activity against pathogens and contribute to limiting pathogen growth and spread [84]. By stimulating the production of phytoalexins, elicitors can enhance the plant's ability to resist infection by wilt and root rot pathogens. Elicitors can also induce the synthesis of antimicrobial peptides (AMPs) and pathogenesis-related (PR) proteins. AMPs are small peptides with antimicrobial activity, and PR proteins are a diverse group of proteins involved in plant defense responses. These defense-related proteins act as molecular weapons against invading pathogens, inhibiting their growth and enhancing the plant's resistance [85]. Elicitor treatments can be applied directly to castor plants through foliar sprays, stem injections, or root drenches. Seed treatments with elicitors have also been investigated as a proactive approach to prime the plant's defense mechanisms from the early stages of growth. The timing and concentration of elicitor application are critical factors for achieving effective resistance induction [86]. The optimal timing depends on the specific pathogen and the plant's growth stage, as well as the mode of action of the elicitor. Careful consideration should be given to the dosage and application frequency to ensure optimal elicitor concentrations that activate the plant's defenses without causing adverse effects. The induction of resistance mechanisms through elicitors provides a proactive and sustainable approach to disease management in castor cultivation [87]. Priming the plant's defense responses, elicitors can enhance resistance to wilt and root rot pathogens, reducing disease severity and improving overall crop health. Moreover, elicitor treatments can be integrated with other management strategies, such as cultural practices and resistant cultivars, to develop integrated disease management programs that promote sustainable and efficient disease control in castor crops [88].

## 6.3 Metabolic Engineering for Disease Resistance

Metabolic engineering involves modifying the metabolic pathways in plants to enhance specific traits, including disease resistance. By manipulating the genes responsible for the synthesis of defense-related compounds or enzymes involved in defense pathways, researchers can increase the production of metabolites that confer resistance against wilt and root rot pathogens [89]. This approach allows for the development of castor plants with enhanced production of antimicrobial compounds or strengthened defense responses, resulting in improved resistance. Physiological and biochemical approaches provide insights into the intricate mechanisms of host-pathogen interactions and the plant's defense responses. By understanding these processes, researchers can identify potential targets for intervention and develop strategies to induce or enhance resistance mechanisms in castor plants [11,90]. The use of elicitors and metabolic engineering techniques offers promising avenues for improving the resistance of castor to wilt and root rot diseases, thereby reducing the yield losses associated with these pathogens.

## 7. FUTURE PERSPECTIVES

### 7.1 Integration of Multiple Approaches for Durable Resistance

The future of breeding strategies for enhancing wilt and root rot resistance in castor lies in the integration of multiple approaches. Combining traditional breeding methods with biotechnological tools, genomic approaches, and physiological and biochemical insights can lead to the development of castor varieties with durable resistance [68]. By utilizing the strengths of each approach, breeders can enhance the efficiency and precision of selecting resistant plants and introduce novel resistance genes or traits.

### 7.2 Gene Editing Technologies and CRISPR/Cas9

Gene editing technologies, particularly the CRISPR/Cas9 system, hold immense potential for precise modification of target genes in castor. With the ability to edit specific genes, including those involved in disease resistance, breeders can introduce precise changes or modifications to enhance resistance to wilt and root rot pathogens [91]. Gene editing technologies offer a faster and more targeted approach compared to traditional breeding or genetic transformation, and they are expected to play a significant role in future castor breeding programs [92].
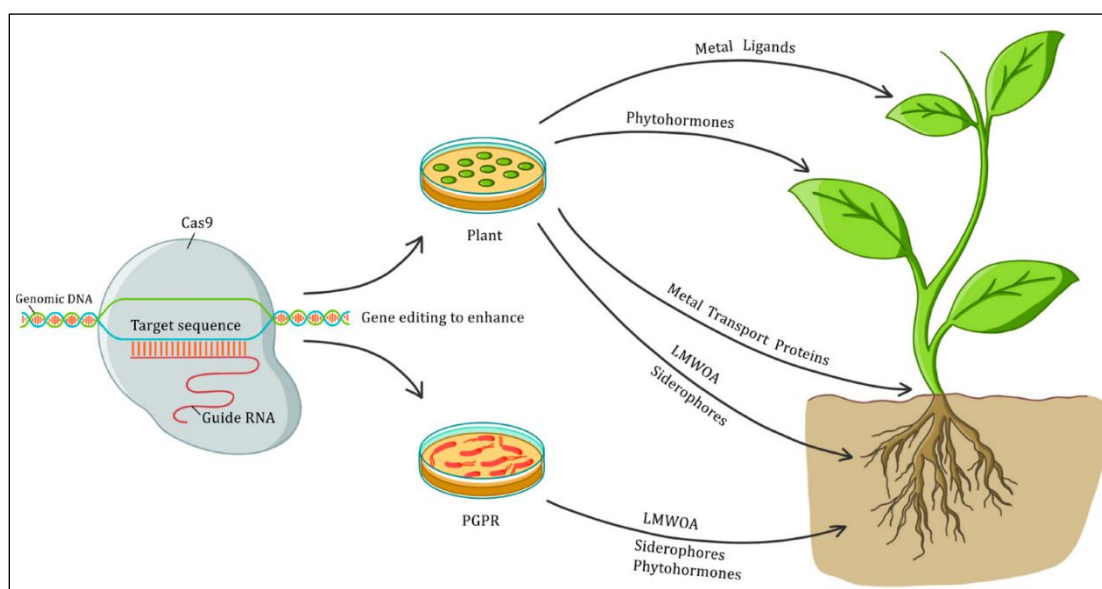
**Fig. 2. Gene editing technology**
*Source: Basharat et al. [93]*

## 7.3 Omics Technologies and Systems Biology Approaches

Omics technologies, such as genomics, transcriptomics, proteomics, and metabolomics, provide comprehensive insights into the complex biological processes underlying disease resistance in castor. Integration of these omics data with systems biology approaches can unravel the intricate molecular networks and pathways associated with resistance mechanisms [94]. This knowledge can be utilized to identify key genes, regulatory elements, and metabolic pathways that can be targeted to enhance resistance. Omics technologies and systems biology approaches will continue to advance our understanding of castor's defense mechanisms and facilitate the development of improved breeding strategies [95].

The future of breeding strategies for enhancing wilt and root rot resistance in castor is promising, with the integration of multiple approaches, the application of gene editing technologies like CRISPR/Cas9, and the utilization of omics technologies and systems biology approaches. These advancements will contribute to the development of castor varieties with enhanced resistance, improved productivity, and sustainable cultivation practices. Continued research and collaboration between breeders, biotechnologists, and scientists across disciplines will drive innovation and progress in castor breeding for disease resistance.

## 8. CONCLUSION

Improving wilt and root rot resistance in castor is vital for sustainable and productive cultivation of this important oilseed crop. A comprehensive review has highlighted advancements in breeding strategies for this purpose. Traditional breeding methods, such as phenotypic selection, recurrent selection, hybridization, mutation breeding, and polyploidy induction, have effectively introduced and accumulated resistance genes and traits in castor germplasm. Biotechnological approaches, such as marker-assisted selection (MAS), genetic transformation, and RNA interference (RNAi), have provided powerful tools to expedite the development of resistant castor varieties. These approaches enable targeted selection and manipulation of specific genes associated with disease resistance, resulting in improved resilience against wilt and root rot pathogens. Moreover, physiological and biochemical approaches focused on understanding host-pathogen interactions, elicitor-induced defense responses, and metabolic engineering show promise in enhancing resistance mechanisms in castor. By gaining insights into the molecular and biochemical processes underlying disease resistance, these approaches open up new avenues for developing innovative strategies to bolster castor's defense against wilt and root rot diseases. The future of breeding strategies for wilt and root rot resistance in castor lies in the integration of multiple approaches, such as

combining traditional breeding methods with biotechnological tools, genomic approaches, and physiological and biochemical insights. Additionally, the advent of gene editing technologies like CRISPR/Cas9 and the utilization of omics technologies and systems biology approaches hold immense potential for precise modification of target genes and unraveling the intricate molecular networks associated with disease resistance.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Román-Figueroa C, Cea M, Paneque M. Industrial oilseed crops in Chile: Current situation and future potential. Biofuels, Bioproducts and Biorefining. 2023;17(1): 273-90.
2. Bhowmick S, Verma A, Kar S, Rai G, Chauhan PS. Economic aspects for eco-friendly strategies to minimize disease incidences and yield loss due to *Macrophomina phaseolina*. In *Macrophomina phaseolina*. Academic Press. 2023;345-355.
3. Pradhan DA, Bagagoni P, Makandar R. Assessing rhizosphere Trichoderma asperellum strains for root colonizing and antagonistic competencies against Fusarium wilt through molecular and biochemical responses in castor. Biological Control. 2023;105280.
4. Tiwari A, Tikoo SK, Angadi SP, Kadaru SB, Ajanahalli SR, Vasudeva Rao MJ. Plant Breeding: Its Evolution and Recent Trends. In Market-Driven Plant Breeding for Practicing Breeders. Singapore: Springer Nature Singapore. 2023; 1-32.
5. Lavanya C, Vishnuvardhan Reddy A, Dutta B, Bandopadhyay R. Classical genetics, cytogenetics, and traditional breeding in castor bean. The castor bean genome. 2018:33-65.
6. Bigini V, Camerlengo F, Botticella E, Sestili F, Savatin DV. Biotechnological resources to increase disease-resistance by improving plant immunity: A sustainable approach to save cereal crop production. Plants. 2021;10(6):1146.
7. Boopathi NM, Boopathi NM. Marker-assisted selection (MAS). Genetic Mapping and Marker Assisted Selection: Basics, Practice and Benefits. 2020;343-88.
8. Gadag RN, Bhat JS, Mukri G, Gogoi R, Suby SB, Das AK, Yadav S, Yadava P, Nithyashree ML, Naidu GK, Yadav SK. Resistance to biotic stress: theory and applications in maize breeding. Genomic Designing for Biotic Stress Resistant Cereal Crops. 2021;129-75.
9. Bocos-Asenjo IT, Niño-Sánchez J, Ginésy M, Diez JJ. New insights on the integrated management of plant diseases by RNA strategies: Mycoviruses and RNA interference. International journal of molecular sciences. 2022;23(16):9236.
10. Martins LA, Kotál J, Bensaoud C, Chmelař J, Kotsyfakis M. Small protease inhibitors in tick saliva and salivary glands and their role in tick-host-pathogen interactions. Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics. 2020;1868(2): 140336.
11. Kaur S, Samota MK, Choudhary M, Choudhary M, Pandey AK, Sharma A, Thakur J. How do plants defend themselves against pathogens-Biochemical mechanisms and genetic interventions. Physiology and Molecular Biology of Plants. 2022;28(2):485-504.
12. Mir AS, Maria M, Muhammad S, Ali SM. Potential of mutation breeding to sustain food security. Genetic Variation. 2020;1-5.
13. Raina A, Khan S, Wani MR, Laskar RA, Mushtaq W. Chickpea (*Cicer arietinum* L.) cytogenetics, genetic diversity and breeding. Advances in Plant Breeding Strategies: Legumes: Volume 7. 2019:53-112.
14. Allan G, Williams A, Rabinowicz PD, Chan AP, Ravel J, Keim P. Worldwide genotyping of castor bean germplasm (*Ricinus communis* L.) using AFLPs and SSRs. Genetic Resources and Crop Evolution. 2008;55:365-78.
15. Saxena RK, Von Wettberg E, Upadhyaya HD, Sanchez V, Songok S, Saxena K, Kimurto P, Varshney RK. Genetic diversity and demographic history of Cajanus spp. illustrated from genome-wide SNPs. Plos One. 2014;9(2):e88568.
16. Wilkes G, Williams JT. Current status of crop plant germplasm. Critical Reviews in Plant Sciences. 1983;1(2):133-81.
17. Dwiningsih Y, Rahmaningsih M, Alkahtani J. Development of single nucleotide polymorphism (SNP) markers in tropical crops. Advance Sustainable Science,

Engineering and Technology. 2020;2(2): 343558.

18. Xu W, Yang T, Qiu L, Chapman MA, Li DZ, Liu A. Genomic analysis reveals rich genetic variation and potential targets of selection during domestication of castor bean from perennial woody tree to annual semi-woody crop. Plant Direct. 2019; 3(10):e00173.

19. Thabuis A, Palloix A, Pflieger S, Daubèze AM, Caranta C, Lefebvre V. Comparative mapping of Phytophthora resistance loci in pepper germplasm: evidence for conserved resistance loci across Solanaceae and for a large genetic diversity. Theoretical and Applied Genetics. 2003;106:1473-85.

20. Anjani K. Castor genetic resources: A primary gene pool for exploitation. Industrial Crops and Products. 2012; 35(1):1-4.

21. Anglin NL, Amri A, Kehel Z, Ellis D. A case of need: Linking traits to genebank accessions. Biopreservation and Biobanking. 2018;16(5):337-49.

22. Anjani K, Raoof MA, Prasad MS, Duraimurugan P, Lucose C, Yadav P, Prasad RD, Lal JJ, Sarada C. Trait-specific accessions in global castor (*Ricinus communis* L.) germplasm core set for utilization in castor improvement. Industrial Crops and Products. 2018;112:766-74.

23. Sharma OP, Bambawale OM. Integrated management of key diseases of cotton and rice. InIntegrated management of diseases caused by fungi, phytoplasma and bacteria 2008 (pp. 271-302). Dordrecht: Springer Netherlands.

24. Padma S, Bongale U, Sukumar J, Mallikarjunappa R, Pushpa R. Root-Knot Nematode Meloidogyne Incognita (Kofoid And White) Chitwood In Mulberry: A Review. Séricologia. 2008;48(1):1-21.

25. Petrasch S, Knapp SJ, Van Kan JA, Blanco-Ulate B. Grey mould of strawberry, a devastating disease caused by the ubiquitous necrotrophic fungal pathogen Botrytis cinerea. Molecular plant pathology. 2019;20(6):877-92.

26. Buerstmayr M, Steiner B, Buerstmayr H. Breeding for Fusarium head blight resistance in wheat—Progress and challenges. Plant breeding. 2020;139(3): 429-54.

27. Zhang J, Abdelraheem A, Zhu Y, Wheeler TA, Dever JK, Elkins-Arce H, Nichols R, Wedegaertner T. Pedigree selection under field conditions within Acala 1517-08 and its glandless derivatives for development of cotton resistant to Fusarium wilt caused by *Fusarium oxysporum* f. sp. *vasinfectum* race 4. Euphytica. 2020;216:1-21.

28. Irulappan V, Mali KV, Patil BS, Manjunatha H, Muhammad S, Senthil-Kumar M. A sick plot–based protocol for dry root rot disease assessment in field-grown chickpea plants. Applications in Plant Sciences. 2021;9(8): e11445.

29. Goggin FL, Lorence A, Topp CN. Applying high-throughput phenotyping to plant–insect interactions: picturing more resistant crops. Current Opinion in Insect Science. 2015;9:69-76.

30. Li W, Ma X, Yong YC, Liu G, Yang Z. Review of paper-based microfluidic analytical devices for in-field testing of pathogens. Analytica Chimica Acta. 2023:341614.

31. Jiang GL. Molecular markers and marker-assisted breeding in plants. Plant breeding from laboratories to fields. 2013;3: 45-83.

32. Singh AS, Kumari S, Modi AR, Gajera BB, Narayanan S, Kumar N. Role of conventional and biotechnological approaches in genetic improvement of castor (*Ricinus communis* L.). Industrial Crops and Products. 2015;74:55-62.

33. Goswami M, Attri K, Goswami I. Applications of molecular markers in fruit crops: A review. International Journal of Economic Plants. 2022;9(2):121-6.

34. Guler BA, Imamoglu E. Molecular marker technologies in food plant genetic diversity studies: An overview. Foods and Raw Materials. 2023;11(2):282-292.

35. Pandurangan S, Workman C, Nilsen K, Kumar S. Introduction to marker-assisted selection in wheat breeding. InAccelerated breeding of cereal crops Oct 21. New York, NY: Springer US. 2021;77-117.

36. Tryphone GM, Chilagane LA, Protas D, Kusolwa PM, Nchimbi-Msolla S. Marker assisted selection for common bean diseases improvements in Tanzania: prospects and future needs. Plant breeding from laboratories to fields. Intech. 2013;22:121-47.

37. Lamichhane S, Thapa S. Advances from conventional to modern plant breeding methodologies. Plant breeding and biotechnology. 2022;10(1):1-4.

38. Lamichhane S, Thapa S. Advances from conventional to modern plant breeding

methodologies. Plant breeding and biotechnology. 2022;10(1):1-4.

39. Maphosa L, Richards MF, Norton SL, Nguyen GN. Breeding for abiotic stress adaptation in chickpea (*Cicer arietinum* L.): A comprehensive review. Crop Breeding, Genetics and Genomics. 2020;4(3).

40. Pathirana R. Plant mutation breeding in agriculture. CABI Reviews. 2011;(2011): 1-20.

41. Rauf S, Al-Khayri JM, Zaharieva M, Monneveux P, Khalil F. Breeding strategies to enhance drought tolerance in crops. Advances in plant breeding strategies: agronomic, abiotic and biotic stress traits. 2016:397-445.

42. Fernández-Martínez JM, Velasco L. Castor. Technological Innovations in Major World Oil Crops, Volume 1: Breeding. 2012:237-65.

43. Arrones A, Vilanova S, Plazas M, Mangino G, Pascual L, Díez MJ, Prohens J, Gramazio P. The dawn of the age of multi-parent MAGIC populations in plant breeding: Novel powerful next-generation resources for genetic analysis and selection of recombinant elite material. Biology. 2020;9(8):229.

44. Facon B, Jarne P, Pointier JP, David P. Hybridization and invasiveness in the freshwater snail Melanoides tuberculata: hybrid vigour is more important than increase in genetic variance. Journal of evolutionary biology. 2005;18(3):524-35.

45. Czajkowski R, Perombelon MC, van Veen JA, van der Wolf JM. Control of blackleg and tuber soft rot of potato caused by *Pectobacterium* and Dickeya species: a review. Plant pathology. 2011;60(6):999-1013.

46. Chattopadhyay A, Seth T, Dutta S, Ghosh PP, Chattopadhyay SB, Majumder D, Hazra P. Breeding eggplant for higher productivity and bacterial wilt tolerance. International journal of vegetable science. 2012;18(4):376-92.

47. Khursheed S, Wani MR, Khan S, Bhat TA, Mutagens. Their types and mechanism of action with an emphasis on sodium azide and gamma radiations aamir raina, shazia bi ansari. mutagenesis, cytotoxicity and crop improvement: Revolutionizing food science; 2021:1.

48. Sharma A, Rathour R, Plaha P, Katoch V, Khalsa GS, Patial V, Singh Y, Pathania NK. Induction of Fusarium wilt (*Fusarium oxysporum* f. sp. pisi) resistance in garden pea using induced mutagenesis and in vitro selection techniques. Euphytica. 2010;173:345-56.

49. Biswas K, Philip S, Yadav A, Martin BK, Burkett S, Singh V, Babbar A, North SL, Chang S, Sharan SK. BRE/BRCC45 regulates CDC25A stability by recruiting USP7 in response to DNA damage. Nature Communications. 2018;9(1):537.

50. Manzoor A, Ahmad T, Bashir MA, Hafiz IA, Silvestri C. Studies on colchicine induced chromosome doubling for enhancement of quality traits in ornamental plants. Plants. 2019;8(7):194.

51. Mason AS, editor. Polyploidy and hybridization for crop improvement. CRC Press; 2017 Nov 22.

52. Dhulgande GS, Dhale DA. Basics of Plant breeding and Seed Technology. Book Saga Publications; 2023.

53. Jauhar PP. Modern biotechnology as an integral supplement to conventional plant breeding: the prospects and challenges. Crop Science. 2006;46(5):1841-59.

54. Yadava DK, Vasudev S, Singh N, Mohapatra T, Prabhu KV. Breeding major oil crops: Present status and future research needs. Technological innovations in major world oil crops. Breeding. 2012;1:17-51.

55. Kaur S, Kimber RB, Cogan NO, Materne M, Forster JW, Paull JG. SNP discovery and high-density genetic mapping in faba bean (*Vicia faba* L.) permits identification of QTLs for ascochyta blight resistance. Plant Science. 2014;217:47-55.

56. Shi Z, Liu S, Noe J, Arelli P, Meksem K, Li Z. SNP identification and marker assay development for high-throughput selection of soybean cyst nematode resistance. BMC genomics. 2015;16:1-2.

57. Dong Z, Hegarty JM, Zhang J, Zhang W, Chao S, Chen X, Zhou Y, Dubcovsky J. Validation and characterization of a QTL for adult plant resistance to stripe rust on wheat chromosome arm 6BS (Yr78). Theoretical and Applied Genetics. 2017; 130:2127-37.

58. Asea G. Genetic characterization of partial resistance and comparative strategies for improvement of host-resistance to multiple foliar pathogens of maize. The Ohio State University; 2005.

59. Veluthambi K, Gupta AK, Sharma A. The current status of plant transformation technologies. Current Science. 2003;84(3): 368-80.

60.  Zhimin Yin GB, Szwacka M, Malepszy S. Cucumber transformation methods-the review. Biotechnologia. 2005;1:95-113.

61.  Shekhawat UK, Ganapathi TR, Hadapad AB. Transgenic banana plants expressing small interfering RNAs targeted against viral replication initiation gene display high-level resistance to banana bunchy top virus infection. Journal of general virology. 2012;93(8):1804-13.

62.  Sanabria N, Goring D, Nürnberger T, Dubery I. Self/nonself perception and recognition mechanisms in plants: a comparison of self-incompatibility and innate immunity. New Phytologist. 2008; 178(3):503-14.

63.  Goodman RE. Biosafety: Evaluation and regulation of genetically modified (GM) crops in the United States.

64.  Zotti M, Dos Santos EA, Cagliari D, Christiaens O, Taning CN, Smagghe G. RNA interference technology in crop protection against arthropod pests, pathogens and nematodes. Pest management science. 2018;74(6): 1239-50.

65.  Guo Z, Li Y, Ding SW. Small RNA-based antimicrobial immunity. Nature Reviews Immunology. 2019;19(1):31-44.

66.  Ghag SB. Host induced gene silencing, an emerging science to engineer crop resistance against harmful plant pathogens. Physiological and Molecular Plant Pathology. 2017;100:242-54.

67.  Vetukuri RR, Dubey M, Kalyandurg PB, Carlsson AS, Whisson SC, Ortiz R. Spray-induced gene silencing: an innovative strategy for plant trait improvement and disease control. Crop breeding and applied biotechnology. 2021;21.

68.  Kumaraswamy HH, Kumar VD, Lavanya C, Ushakiran B, Senthilvel S, Sujatha M, Durga Bhavani KB, Konda AK, Lakshmidevi G, Ashfaq AM, Kumar BS. Biotechnological approaches for genetic improvement of castor bean (*Ricinus communis* L.). Accelerated Plant Breeding, Volume 4: Oil Crops. 2022:359-418.

69.  Amin S, Islam T. Genetics and Genomics of Biotic Stress Resistance of Jute. InThe Jute Genome 2022;(pp. 269-289). Cham: Springer International Publishing.

70.  Jaganathan D, Bohra A, Thudi M, Varshney RK. Fine mapping and gene cloning in the post-NGS era: Advances and prospects. Theoretical and Applied Genetics. 2020;133(5):1791-810.

71.  Liu Y, Shao L, Zhou J, Li R, Pandey MK, Han Y, Cui F, Zhang J, Guo F, Chen J, Shan S. Genomic insights into the genetic signatures of selection and seed trait loci in cultivated peanut. Journal of Advanced Research. 2022;42:237-48.

72.  Abebe AM, Choi J, Kim Y, Oh CS, Yeam I, Nou IS, Lee JM. Development of diagnostic molecular markers for marker-assisted breeding against bacterial wilt in tomato. Breeding science. 2020;70(4):462-73.

73.  Polanco C, Sáenz de Miera LE, González AI, García P, Fratini R, Vaquero F, Vences FJ, Pérez de la Vega M. Construction of a high-density interspecific (*Lens culinaris* x *L. odemensis*) genetic map based on functional markers for mapping morphological and agronomical traits, and QTLs affecting resistance to Ascochyta in lentil. PloS one. 2019;14(3):e0214409.

74.  Spielmeyer W, Green AG, Bittisnich D, Mendham N, Lagudah ES. Identification of quantitative trait loci contributing to Fusarium wilt resistance on an AFLP linkage map of flax (*Linum usitatissimum*). Theoretical and applied genetics. 1998; 97:633-41.

75.  Xu Y, Crouch JH. Marker-assisted selection in plant breeding: From publications to practice. Crop science. 2008;48(2):391-407.

76.  Shaw RK, Shaik M, Prasad MS, Prasad RD, Mohanrao MD, Senthilvel S. Genomic regions associated with resistance to Fusarium wilt in castor identified through linkage and association mapping approaches. Genome. 2022;65(3):123-36.

77.  Aslam S, Chouhan S, Harish V, Lalotra S. Seed Priming: A Key to Sustainability in Drought Stress. International Journal of Environment and Climate Change. 2023;13(8):1871-82.

78.  Schlatter D, Kinkel L, Thomashow L, Weller D, Paulitz T. Disease suppressive soils: new insights from the soil microbiome. Phytopathology. 2017; 107(11):1284-97.

79.  Heil M, Bostock RM. Induced systemic resistance (ISR) against pathogens in the context of induced plant defences. Annals of botany. 2002;89(5):503-12.

80.  Yu J, Yu X, Li C, Ayaz M, Abdulsalam S, Peng D, Qi R, Peng H, Kong L, Jia J, Huang W. Silicon mediated plant immunity against nematodes: Summarizing the underline defence mechanisms in plant

nematodes interaction. International Journal of Molecular Sciences. 2022;23(22):14026.

81. Fernandes LB, Ghag SB. Molecular insights into the jasmonate signaling and associated defense responses against wilt caused by Fusarium oxysporum. Plant Physiology and Biochemistry. 2022;174:22-34.

82. Chandra S, Chakraborty N, Dasgupta A, Sarkar J, Panda K, Acharya K. Chitosan nanoparticles: A positive modulator of innate immune responses in plants. Scientific reports. 2015;5(1):15195.

83. Doughari J. An overview of plant immunity. J. Plant Pathol. Microbiol. 2015;6(11):10-4172.

84. Hammond-Kosack KE, Jones JD. Resistance gene-dependent plant defense responses. The Plant Cell. 1996;8(10):1773.

85. Odintsova TI, Slezina MP, Istomina EA, Korostyleva TV, Kasianov AS, Kovtun AS, Makeev VJ, Shcherbakova LA, Kudryavtsev AM. Defensin-like peptides in wheat analyzed by whole-transcriptome sequencing: A focus on structural diversity and role in induced resistance. PeerJ. 2019;7:e6125.

86. Pontarin N. Metabolic response of flax (*Linum usitatissimum*) plant and seed to the elicitation with aminobutyric acids (Doctoral dissertation, Amiens).

87. Stout MJ, Zehnder GW, Baur ME. Potential for the use of elicitors of plant resistance in arthropod management programs. Archives of Insect Biochemistry and Physiology: Published in Collaboration with the Entomological Society of America. 2002;51(4):222-35.

88. Khan SM, Ali S, Nawaz A, Bukhari SA, Ejaz S, Ahmad S. Integrated pest and disease management for better agronomic crop production. Agronomic Crops: Volume 2: Management Practices. 2019;385-428.

89. Abbas F, Zhou Y, O'Neill Rothenberg D, Alam I, Ke Y, Wang HC. Aroma components in horticultural crops: Chemical diversity and usage of metabolic engineering for industrial applications. Plants. 2023;12(9):1748.

90. Wenneker M, Thomma BP. Latent postharvest pathogens of pome fruit and their management: From single measures to a systems intervention approach. European Journal of Plant Pathology. 2020;156:663-81.

91. Montecillo JA, Chu LL, Bae H. CRISPR-Cas9 system for plant genome editing: Current approaches and emerging developments. Agronomy. 2020;10 (7):1033.

92. Miladinovic D, Antunes D, Yildirim K, Bakhsh A, Cvejić S, Kondić-Špika A, Marjanovic Jeromela A, Opsahl-Sorteberg HG, Zambounis A, Hilioti Z. Targeted plant improvement through genome editing: from laboratory to field. Plant cell reports. 2021;40:935-51.

93. Basharat Z, Novo LA, Yasmin A. Genome editing weds CRISPR: What is in it for phytoremediation?. Plants. 2018;7 (3):51.

94. Raza A, Tabassum J, Fakhar AZ, Sharif R, Chen H, Zhang C, Ju L, Fotopoulos V, Siddique KH, Singh RK, Zhuang W. Smart reprograming of plants against salinity stress using modern biotechnological tools. Critical Reviews in Biotechnology. 2022; 12:1-28.

95. Mahmood U, Li X, Fan Y, Chang W, Niu Y, Li J, Qu C, Lu K. Multi-omics revolution to promote plant breeding efficiency. Frontiers in Plant Science. 2022;13: 1062952.

96. Anonymous. Department of Agriculture and Co-operation, Government of India; 2017. Available: http: // Agricop.nic.in. accessed on 26th Aug.

97. Priya PB. Identification of potential sources of resistance for wilt caused by *Fusarium oxysporumf. sp. riciniin* castor (*Ricinus communis* L.). Asian Journal of Bio Science. 2016;11(2):284-8.