

Annual Research & Review in Biology

37(5): 15-29, 2022; Article no.ARRB.86720
ISSN: 2347-565X, NLM ID: 101632869

Isolation and Characterization of Microbes of Different Kinds of Street Food and Determination of Antibiotic Susceptibility of the Isolates

Reddy Krishna Manasa ^{a*} and Anitha Thomas ^a

^a Department of Microbiology, St. Francis College for Women, Begumpet, Hyderabad, India.

Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ARRB/2022/v37i530505

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/86720>

Original Research Article

Received 15 February 2022

Accepted 27 April 2022

Published 03 May 2022

ABSTRACT

Aim: Improper personal hygiene can facilitate the transmission of the pathogenic microorganisms found in environment and on people's hands via food to humans. The present study was undertaken to investigate the microbiological quality of different street food and determining the antibiotic susceptibility of the isolated microorganisms.

Study Design: Collection of food samples for isolation of Pathogenic Microorganisms, to identify them by using Biochemical test, molecular test (16sr RNA typing), Antibiotic susceptibility was done by using different antibiotics against the isolates.

Place and Duration of Study: Food samples were collected from street vendors of Sainkपुरi area, work was done from December 2020 to April 2021 at Microbiology Department, St. Francis College for Women, Hyderabad.

Methodology: Five samples of street food were collected in sterilized bottles, tested for the presence of microorganisms by following standard microbiological method used for isolation of microorganisms. The organisms were identified by carrying out various biochemical test according to Bergey's Manual of Systematic Bacteriology. The Molecular characterization was done based on 16sr RNA typing. Determining the sensitivity of the isolates against different Antibiotics by employing Kirby Bauer technique.

Results: The organisms isolated from Manchuria and Ragada samples: *Lactobacillus delbrueckii*; *Lactobacillus casei*, *Klebsiella pneumonia*, *Enterococcus faecalis* and *Vibrio spp* respectively;

*Corresponding author: E-mail: reddykrishnamanasa@gmail.com;

Samosa and bonda: *Staphylococcus aureus*, *Lactobacillus delbrueckii* and *Pantoea dispersa*; Pani puri water: *Staphylococcus aureus* and *Providencia vermicola*. Antibiotic susceptibility tests showed that most of the isolated microorganisms were sensitive to Ciproflaxin.

Conclusion: Hence the quality of street food is found to be low due to following factor: lack hygiene conditions in the food preparation, ingredients may also affect quality of the food, and presence of air borne microorganisms in the surrounding areas of service points.

Keywords: Street food; quality status; pani-puri; bonda; samosa; manchuria; microorganism.

1. INTRODUCTION

Street food is ready-to-eat food in public place such as market, by a seller often from available food cart. There are a plenty of types of food sold on the street and different places bring different flavors of street foods. The street foods are the well appreciated by consumers, because of to its taste, low cost, and availability for immediate consumption [1]. The main reasons for spreading of diseases in street food is because sellers do not have sufficient information -and details about food safety. Rise of food originated diseases is related with wrong storage (50%), reheating-storing under inappropriate conditions (45%), and cross contamination (39%) [2].

These street foods are also subjected to cross-contamination from various sources like utensils, knives, raw foodstuffs, flies that sporadically landing on the foods, vendors use their bare hand for serving and occasional food handling by consumers. In most of the cases, tap water is not available for washing hands and utensils at vending sites; hand and utensil washing are usually done in one or more buckets-sometimes without soap [3].

Street food safety remains a major concern in many developing countries, including developed countries like China. Chinese food culture has a very long history of street food. Most cities provide street food for locals and tourists, and street food has become a part of the characteristic Chinese culture. Therefore, street food safety has become a matter of safety concern, and has been shown to be served in poor food handling and unsanitary conditions. The street food vendors are mostly uneducated and often uninformed, and have little effective regulatory or supervisory oversight. In some developing countries, street food has been associated with outbreaks of many foodborne diseases. High levels of coliform bacteria have

been found in street food in several countries around the globe, and street food has been identified as a common medium for transmission of antimicrobial-resistant pathogens [4].

Street vended foods are known to be contaminated with pathogens, which might pose a health hazard. Bacteria belonging to the genus *Bacillus*, *Staphylococcus*, *Clostridium*, *Vibrio*, *Campylobacter*, *Listeria*, *Salmonella* are reported mostly from street foods [5].

In this study we have chosen street food like chat, Chinese fast food and deep fry food. In Chat section we have chosen Pani (Pani puri water) of Pani-Puri and Ragada. In the deep fry section, we have chosen Bonda and Samosa. Finally, In Chinese fast food we have chosen Veg. Manchuria.

Different type of pathogenic organisms was identified from the street food, indicating that quality of street food is low and consumption of these kind if street food is not good for health.

2. MATERIALS AND METHODS

2.1 Collection of Samples

A total number of 5 food samples (Manchuria, Ragada, samosa, bonda and pani puri water samples) were included in this study. Sample was collected from different street vended foods in sainkpuri, samples were collected into sterilized bottles under sterile conditions. All the collected samples were kept on an icebox during transportation to the laboratory and stored at 4°C until testing. They were analyzed within 24h of sampling. The samples were labelled as:

- a. FS1- Manchuria
- b. FS2- Ragada
- c. FS3- Samosa
- d. FS4- Bonda
- e. FS5- Pani puri water

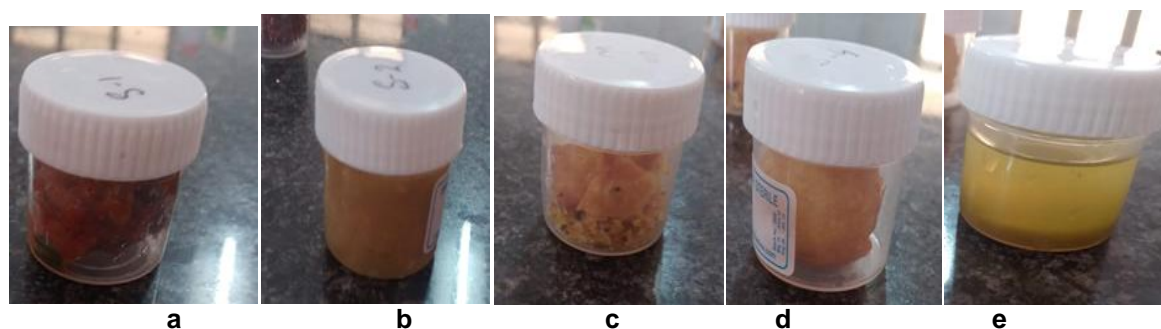


Fig. 1. Samples collected into sterile containers
Samples collected: a. Manchuria, b. Ragada, c. Samosa, d. Bonda, e. Pani puri water

2.2 Microbiological Analysis

One gram of each sample of Manchuria, Ragada, samosa, bonda and pani puri water samples separately inoculated into sterile water and homogenized using glass rod. After that 100µl of homogenized sample was inoculated to nutrient broth and incubated at 37°C for 24h [5]. The overnight culture was streaked into Nutrient agar, Mannitol Salt agar and Eosin Methylene Blue agar. The culture is inoculated into media by spread plate technique and incubated at 37°C for 24h. The colonies on primary cultures were sub cultured by streak plate method on to nutrient agar slants, and incubate at 37°C for 24h. These nutrient slants are preserved for further test.

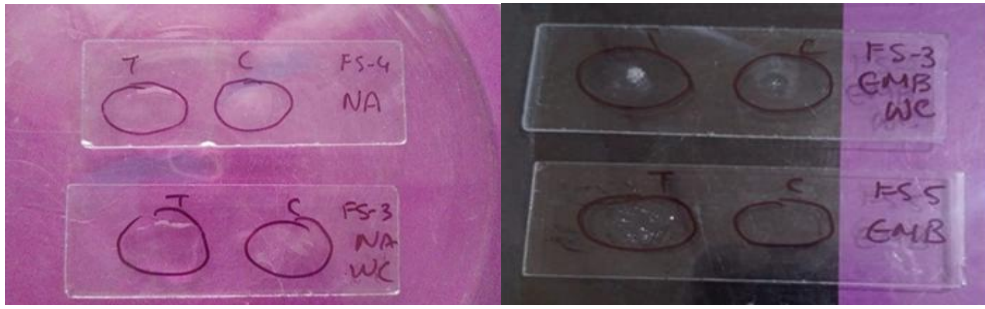
2.3 Auto Vitek Method

It is a fully automated system which performs antibiotic susceptibility testing and bacterial identification. User interface screen for immediate notification of system status to increase productivity. Unique vacuum filler provides both safety and the highest level of automation [6].

2.4 Identification and Biochemical Characterization of Isolates

The isolated colonies were identified by colony morphology and gram staining. The Biochemical test was done based on gram staining, and Bergeys manual was followed for biochemical test. In biochemical characterization endospore staining, acid fast staining, catalase test, oxidase test, carbohydrate fermentation (mannitol and glucose), sodium required for growth test, blood agar and luminescent technique were performed.

| S. No | Gram nature | Test performed |
|-------|---|--|
| 1. | Gram positive rod (FS1 NA, FS2 MSA, FS3 NA white colony and growth colony, FS4 NA) | Endospore staining, Acid fast staining, Catalase test, Glucose fermentation, Mannitol fermentation |
| 2. | Gram positive cocci (FS2 NA-small colony, FS3 MSA, FS3 EMB, FS5 NA, FS5 MSA (small and big colony), FS5 EMB) | Catalase test, blood agar and Mannitol fermentation. |
| 3. | Gram negative cocci (FS3 EMB and FS5 NA) | Molecular test |
| 4. | Gram negative rods (FS2 EMB, FS2 NA big colony) | Oxidase test, Glucose fermentation, Sodium required for growth test and Luminescent technique |



(a)

(b)

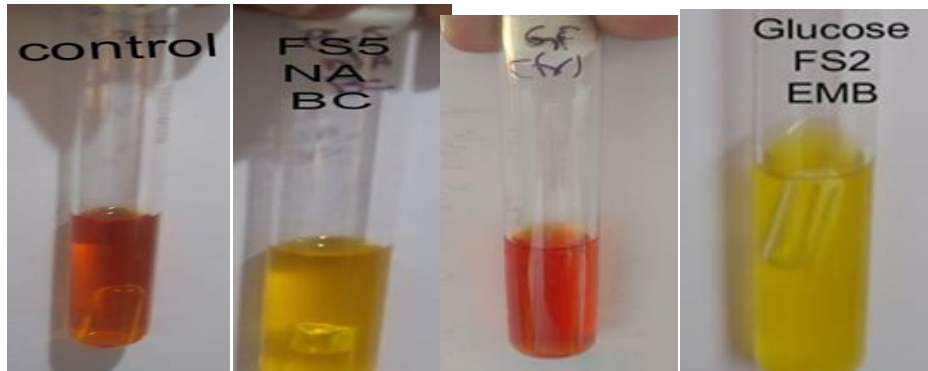


(b) i

ii

(d) i

ii

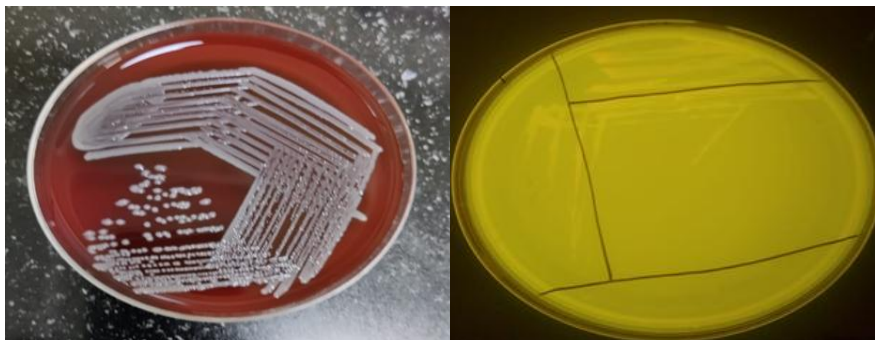


(e) i

ii

(f) i

ii



(g)

(h)



(i)

- Fig 2: represents the biochemical test performed to identify the organism:**
- a. Catalase test: Negative for sample FS4 NA; Negative for sample FS3 NA (White colony), b. Catalase test: positive for sample FS3 EMB and FS5 EMB,**
 - c. Glucose Fermentation Test: i) control; ii) Positive for the sample FS1,**
 - d. Mannitol Fermentation Test: i) control; ii) Positive for sample FS2 MSA,**
 - e. Mannitol Fermentation test: i) control; ii) Positive for sample FS5 NA,**
 - f. Glucose Fermentation test: i) control; ii) Positive for sample FS2 EMB,**
 - g. Blood agar: Gamma hemolysis of sample FS2 NA,**
 - h. Luminescent Technique: Sample FS2 EMB showing negative for luminescent technique,**
 - i. Oxidase test: Positive for sample FS2 EMB.**

2.5 Molecular Test (16sr RNA typing)

Traditional methods to identify and characterize microbes require growth of isolates on culture plates. Although these methods have allowed many essential discoveries, there are limitations. In particular, some microbial species do not grow readily in routine laboratory culture conditions or as monocultures, leading to underestimations of the diversity of microbial communities. (Jay-HyunJo1Elizabeth, A. Kenned et al, 2016)

Next-generation sequencing technologies have allowed direct and comprehensive sequence-based interrogation of microbial communities, complementing culture-based methods.

The 16S ribosomal RNA (rRNA) gene is conserved among prokaryotes with specific variable regions that can be used for taxonomic classification, making the 16S rRNA gene a molecular signature to identify members of bacterial communities.

Procedure that is followed

1. Pellet out 1.5ml of an overnight bacterial culture by centrifugation for 2min at 8000 rpm.
2. Remove the culture medium and discard.
3. Resuspend the pellet thoroughly in 180µl of lysis solution and add 20µl of RNase
4. solution, mix and incubate for 2min at RT (15-20 o C)
5. Add 20µl of proteinase k solution, mix and incubate for 30min at 55oC.
6. Add 200µl of lysis solution, vortex thoroughly (15 sec) and incubate at 55oC for 10min.
7. Add 200µl of Ethanol (96-100%) to the lysine and mix thoroughly by vertexing for 15seconds.
8. Load lysine in Hi-elute miniprep spin column, centrifuge at 6500xg for 1min. Discard the flow through liquid and place the spin column in a new 2ml collection tube.
9. Add 500µl of prewash solution to the Hi-elute miniprep spin column and centrifuge at 6500xg for 1min. Discard the flow through liquid and reuse the same collection tube with the column.
10. Add 500µl of wash solution to the column and centrifuge for 3min at maximum speed (14000rpm) to dry the column. The column must be free of ethanol before eluting the DNA.
11. Centrifuge the column for the additional 1min at maximum speed if residual ethanol is seen.
12. Pipette 200µl of the elution buffer directly in to the column without spilling to the slides. Incubate for 1min at RT. Centrifuge at 10000rpm for 1min to elute the DNA

2.6 Antibiotic Susceptibility Test

An antibiotic sensitivity test is used to help find the best treatment for a bacterial infection. Procedure that is followed:

1. Inoculate the culture into nutrient broth, incubate at 37 °C for 24h
2. This 24h active culture is used for antibiotic susceptibility test.
3. Prepare Muller Hinton media and autoclave at 121°C 15 minutes.
4. Pour into petri plates and allow it to solidify.
5. Inoculate 24h active culture by spread plate technique
6. The antibiotic disc should be inserted into petri plate by use of sterile forceps.
7. Incubate at 37°C for 24h.
8. After incubation, the growth of bacteria is observed.
9. Areas around the antibiotic disc where no bacterial growth can be seen are known as 'zones of inhibition.
10. These zones show that an antibiotic has been successful in stopping bacterial growth or killing the bacteria.
11. By measuring the diameter of these zones, we can compare the efficacy of antibiotics and monitor antimicrobial resistance.

2.7 Determination of Microbiological Quality of Street Food

The quality of street food depends upon organisms isolated from the street food, isolated organisms are identified by their morphology, gram staining, biochemical test. Depending on these tests we can identify that isolate organism is pathogenic or not, and it is either effecting the quality of street food or not.

3. RESULTS AND DISCUSSION

3.1 Collection of Samples

The food samples collected were homogenised and inoculated into nutrient broth, after 24h of incubation inoculated into 3 different media

- a. Nutrient agar (NA),
- b. Eosin methylene blue agar (EMB),
- c. Mannitol salt agar (MSA).

3.2 Microbiological Analysis

After 24h, the culture is inoculated into 3 different media (NA EMB, MSA), incubated at 37°C for 24h, Fig.3 represents the colonies observed on respective media, the colonies were sub cultured into nutrient agar and incubated at 37°C for 24h, the growth on nutrient agar slants was observed, these nutrient slants are preserved for further test.

Eight genera of bacteria were isolated, *Lactobacillus delbrueckii*, *Enterococcus faecalis*, *Klebsiella pneumonia*, *Lactobacillus casei*, *Vibrio* species, *Staphylococcus aureus*, *Pantoea dispersa*, *Providencia vermicola* were isolated from respective street food samples.

3.3 Auto Vitek Method

The isolated organism from the Ragada sample is determined as *Klebsiella pneumonia* by auto Vitek method.

Hence the organisms identified are *Klebsiella pneumonia* is a bacterium that normally lives inside human intestines, where it doesn't cause disease. But if *K. pneumoniae* enters into other areas of the body, it can lead to a range of illnesses, including pneumonia, bloodstream infections, meningitis, and urinary tract infections [7].

The *Vibrio* spp is a Gram-negative bacterium, several species of which can cause foodborne infection. These bacteria can cause two types of illness: vibriosis and cholera.

3.4 Identification and Biochemical Characterization of Isolates and Gram Staining

In this study, different microbiological techniques were used to identify bacteria isolated from street food samples. The results of cultural characteristics, Gram's staining was used to identify the organisms isolated from Manchuria, Ragada, Samosa, Bonda and Pani-Puri water samples. Gram character and Cultural characteristics of isolated bacteria exhibited on the selective media are represented in table 1.

3.4.1 Biochemical characteristics

The organisms isolated from the food samples (FS1, FS2, FS3, FS4, FS5) were characterized

by gram staining and further identified based on Bergey's Manual of Systematic Bacteriology.

infection. [8] The *Lactobacillus delbrueckii* is a pathogenic organism.

Below tables represents the biochemical test done based on gram nature.

3.4.1.1 Gram-positive rods

In current study the organisms isolated from the above samples are *Lactobacillus casei* and *Lactobacillus delbrueckii* is determined to be the causative microorganism for urinary tract

3.4.1.2 Gram-positive cocci

In current study organisms isolated from Ragada were *Enterococcus faecalis* which causes nausea, diarrhea, UTI [9] and other isolated organism is *Staphylococcus aureus* causes diseases like Staph food poisoning- it is a gastrointestinal illness, pneumonia and blood stream infections [10].

Table 1. Colony characteristics, morphology and gram staining of the isolated organisms

| Sample | Size | Shape | Colour | Opacity | Margin | Elevation | Consistency |
|-------------------------------|--------|-----------|------------------------|-------------|----------------------------|-----------|-------------|
| FS-1; NA | Medium | Round | Crème | Opaque | Irregular | Flat | Smooth |
| FS2; NA (Small colony) | Small | Round | Crème | Opaque | Irregular | Flat | Mucoid |
| FS2; NA (Big colony) | Medium | Round | Crème | Opaque | Irregular | Flat | Mucoid |
| FS2; MSA FS2; EMB | Small | Pinpoint | Yellow Light purple | Opaque | Regular Diffused growth | Flat | Mucoid |
| FS-3; NA (white colonies) | Medium | Round | White | Opaque | Irregular | Flat | Rough |
| FS-3; NA (growth colony) | | | White | | Diffused growth | | |
| FS-3; MSA | Small | Round | Yellow | Opaque | Regular | Flat | Moist |
| FS3; EMB (metallic sheen) | Small | Irregular | Metallic sheen | Opaque | Regular | Raised | Smooth |
| FS-3: EMB (white colonies) | Small | Pinpoint | White | Opaque | Regular | Raised | Smooth |
| FS4; NA | | | White | | Diffused growth | | |
| FS5; NA (white colony) | Big | Round | White | Translucent | Entire | Flat | Smooth |
| FS5; NA (small colonies) | Small | Round | White | Translucent | Entire | Flat | Rough |
| FS5; MSA (small colony) | Small | Round | White | Translucent | Entire | Raised | Moist |
| FS5; MSA (medium colony) | Medium | Round | White | Translucent | Entire | Raised | Moist |
| FS5; EMB | Small | Oval | Light violet | Translucent | Entire | Raised | Moist |

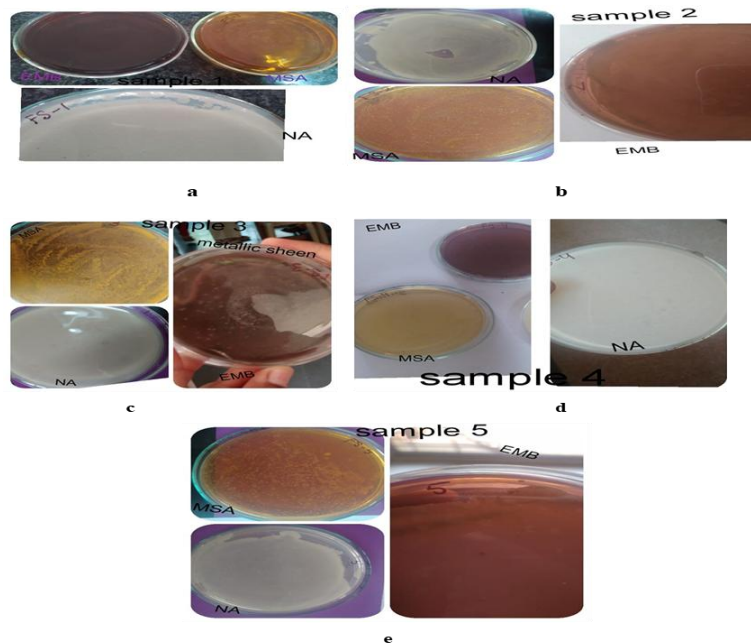


Fig. 3. Represents colony characteristics of isolated organisms from street food
a. Sample 1- Manchuria, b. Sample 2- Ragada, c. sample 3- samosa, d. sample 4- Bonda, e. Sample 5- pani puri water

3.4.1.3 Gram-negative rods

Hence the organisms identified are *Klebsiella pneumoniae* is a bacterium that normally lives inside human intestines, where it doesn't cause disease. But if *K. pneumoniae* enters into other areas of the body, it can lead to a range of illnesses, including pneumonia, bloodstream infections, meningitis, and urinary tract infections [11].

The *Vibrio spp* is a Gram-negative bacterium, several species of which can cause foodborne infection. These bacteria can cause two types of illness: vibriosis and cholera [12].

3.4.1.4 Gram-negative cocci

The isolated organisms from the sample's samosa and pani puri water are determined as *Pantoea dispersa* and *Providencia vermicola* respectively by 16s r RNA typing.

3.5 Molecular detection (16s r RNA typing)

3.5.1 Characterization of aerobic and anaerobic isolate of FS3 EMB and FS5 NA by 16s rRNA typing

Organism identified as *Pantoea dispersa* and *Providencia vermicola*

3.5.2 Blast report

Hence the organisms identified by 16s r RNA typing *Providencia vermicola* are a gram-negative organism that produce bacterial urease, an important virulence factor associated with the formation of urinary tract stones, the obstruction of long-term urinary catheters, and the development of acute pyel the obstruction of long-term urinary catheters, and the development of acute pyelonephritis [13], *Pantoea dispersa* natural habitat are plant or insect pathogens [14]; although they may also play some beneficial roles, but in some special circumstances such as puncture wounds because of plant matter; contaminated cotton bolls/dressing material, or in the hospital settings, in intravenous fluids, as virulent opportunists; in immunosuppressed and elderly or new-born, this organism grown in the tracheal secretions turned out to be an uncommon, unusual Gram-negative Coccobacillus by the name of *Pantoea dispersa* [15].

3.6 Antibiotic Susceptibility Test

This test will determine the isolated organism is resistant or sensitive to particular drug, hence we determine the drug of choice for the particular organism depending upon on the zone of inhibition. Zone of inhibition is measured in mm.

Table 2. Biochemical characterization of Gram-positive rods

| Sample | Endospore staining | Acid fast staining | Catalase | Glucose fermentation | Mannitolfermentation | Organism |
|-----------------------|--------------------|--------------------|----------|----------------------|----------------------|----------------------------------|
| FS1 NA | - | - | - | + | - | <i>Lactobacillus delbrueckii</i> |
| FS2 MSA | - | - | - | + | + | <i>Lactobacillus casei</i> |
| FS3 NA(white colony) | - | - | - | + | - | <i>Lactobacillus delbrueckii</i> |
| FS3 NA(growth colony) | - | - | - | + | - | <i>Lactobacillus delbrueckii</i> |
| FS4 NA | - | - | - | + | - | <i>Lactobacillus delbrueckii</i> |

Table 3. Biochemical characterization of Gram-positive cocci

| Sample | Catalase | Mannitol | Haemolytic pattern on blood agar | Automated Vitek method | Organism |
|--------------------------|----------|----------|----------------------------------|------------------------|------------------------------|
| FS2 NA (small colonies) | - | - | Gamma haemolysis | + | <i>Enterococcus faecalis</i> |
| FS3 MSA | + | + | Not required | NR | <i>Staphylococcus aureus</i> |
| FS3 EMB | + | + | Not required | NR | <i>Staphylococcus aureus</i> |
| FS5 NA | + | + | Not required | NR | <i>Staphylococcus aureus</i> |
| FS5 MSA (small colonies) | + | + | Not required | NR | <i>Staphylococcus aureus</i> |
| FS5 MSA (large colony) | + | + | Not required | NR | <i>Staphylococcus aureus</i> |

Table 4. Biochemical characterization of gram-negative rods

| Sample | Oxidase Test | Glucose Fermentation | Sodium required for growth | Luminescent technique | Organism |
|---------|--------------|----------------------|----------------------------|-----------------------|-------------------|
| FS2 EMB | + | + | + | - | <i>Vibrio spp</i> |

| Sample | Haemolytic pattern on blood agar | Automated method | Vitek | Organism |
|-----------------------|----------------------------------|------------------|-------|------------------------------|
| FS2 NA (large colony) | Gamma Haemolysis | + | | <i>Klebsiella pneumoniae</i> |

Table 5. Represents samples which were characterized by 16s r RNA typing

| Sample | 16s r RNA typing | Organism |
|-------------------------|------------------|------------------------------|
| FS3 EMB(metallic sheen) | + | <i>Pantoea dispersa</i> |
| FS5 NA (small colony) | + | <i>Providencia vermicola</i> |

BLAST REPORT-FS3

| Description | Scientific name | Max Score | Total Score | Query Cover | E Value | Per ident |
|--|------------------|-----------|-------------|-------------|---------|-----------|
| Pantoea dispersa stram BJQ000/chromosome complete genome | Pantoea Dispersa | 1602 | 20022 | 99% | 0.0 | 99.32% |
| Pantoea dispersa stram AS1816S ribosomal RNA gene partial sequence | Pantoea dispersa | 1602 | 2862 | 99% | 0.0 | 99.32% |
| Pantoea dispersa stram S38 ITI 16S ribosomal RNA gene partial sequence | Pantoea dispersa | 1600 | 2846 | 98% | 0.0 | 99.32% |
| Pantoea dispersa stram YBB19B 16S ribosomal RNA gene partial sequence | Pantoea dispersa | 1589 | 2814 | 96% | 0.0 | 99.54% |

BLAST REPORT-FS5

| Description | Scientific name | Max Score | Total Score | Query Cover | E Value | Per ident |
|--|-------------------------------|-----------|-------------|-------------|---------|-----------|
| <i>Providencia sp</i> RS-2A16S ribosomal RNA gene, partial sequence | <i>Providencia sp</i> R4-2A | 1219 | 2367 | 97% | 0.0 | 91.93% |
| <i>Providencia sp</i> 1701091 chromosome, complete genome | <i>Providencia sp</i> 1701091 | 1216 | 16615 | 97% | 0.0 | 92.00% |
| <i>Providencia sp</i> 1701011 chromosome, complete genome | <i>Providencia sp</i> 1701011 | 1216 | 16615 | 97% | 0.0 | 92.00% |
| <i>Providencia vermicola</i> strain AAUBC-Pv2 16S ribosomal RNA gene, partial sequence | <i>Providencia vermicola</i> | 1219 | 1219 | 54% | 0.0 | 92.19% |

3.6.1 Antibiotic susceptibility of gram-positive rod

The above Fig 4 represents the graphical presentation of antibiotic susceptibility of gram positive rods which determines that zone of inhibition for sample FS3 NA (white colony) is high for antibiotic Azithromycin and zone of inhibition for sample FS1 NA is high for antibiotic Ciproflaxin, zone of inhibition for sample FS2 MSA is high for antibiotic is amikacin, FS3 NA (growth colony) is having zone of inhibition is high for antibiotic Ciproflaxin, FS4 NA having zone of inhibition high for antibiotic Clarithromycin. The isolate *Lactobacillus delbrueckii* is resistant to Ampiclox, cefapoperazone, clarithromycin, Cefuroxime. The isolate *Lactobacillus casei* is resistant to Ampiclox and Cefotaxime.

In current study *Lactobacillus delbrueckii* is resistant to Ampiclox, Cefapoperazone, Clarithromycin and Cefuroxime are drug of choice for the diseases caused by this organism is Ciproflaxin, the isolated organism *Lactobacillus casei* is resistant to both Ampiclox and Cefotaxime, and drug of choice for the diseases that caused by this organism is Ciprofloxacin and Amikacin.

3.6.2 Antibiotic susceptibility of gram positive cocci

The Fig 5 determines the zone of inhibition of the isolates, the sample FS2 NA is having zone of inhibition high for antibiotic Ciproflaxin, the FS3 MSA is having zone of inhibition high for antibiotic Ciproflaxin, for FS3 EMB having zone of inhibition high for antibiotic levofloxacin, for FS5 NA having zone of inhibition high for antibiotic Ciproflaxin, for FS 5 MSA (small colony) having zone of inhibition high for antibiotic Ciproflaxin, for FS5 MSA (large colony) having zone of inhibition high for antibiotic cefotaxime. The isolate staphylococcus aureus is resistant to Ampiclox and roxithromycin.

In current study *Enterococcus faecalis* is resistant Ampiclox, Cefuroxime and Roxithromycin and drug of choice for the diseases caused by this organism is Ampiclox, Cefuroxime and Roxithromycin, and the isolate *Staphylococcus aureus* is resistant to Ampiclox and roxithromycin and drug of choice for the diseases that caused by this organism is Ciproflaxin.

Table 6. Antibiotic susceptibility test

| Sample | AN | ACX | CIP | CLR | CF | LE | CR | CFP | BA | AZ | G | RX |
|-----------|----|-----|-----|-----|----|----|----|-----|----|----|----|----|
| FS1 NA | 20 | - | 27 | - | - | 20 | - | - | 17 | 7 | 18 | 12 |
| FS2 MSA | 17 | - | 17 | 13 | - | 13 | 6 | 13 | 9 | 10 | 15 | 10 |
| FS3 NA WC | 10 | - | 22 | 20 | - | 3 | 6 | 13 | 21 | 28 | 15 | 10 |
| FS3 NA GC | 15 | - | 20 | 19 | - | 19 | - | 13 | 14 | 13 | 14 | 16 |
| FS4 NA | 15 | - | 20 | 19 | - | 19 | - | 13 | 7 | 13 | 15 | 15 |

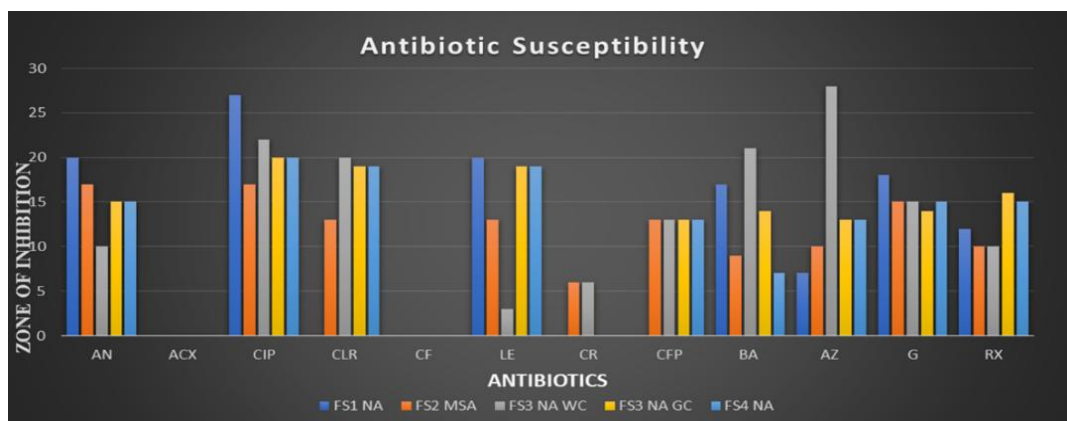


Fig. 4 .Graphical presentation of antibiotic susceptibility of gram-positive rod

Table 7. Antibiotic susceptibility test

| Sample | AN | ACX | CIP | CLR | CF | LE | CR | CFP | BA | AZ | G | RX |
|---------------------------|----|-----|-----|-----|----|----|----|-----|----|----|----|----|
| FS2 NA (Small colony) | 16 | - | 21 | 1 | 17 | 21 | - | 17 | 15 | 16 | 14 | - |
| FS3 MSA | 20 | 24 | 25 | 19 | 24 | 22 | 24 | 23 | 14 | 15 | 12 | 15 |
| FS3 EMB | 15 | 2 | 1 | 1 | 14 | 18 | 15 | 16 | 15 | 12 | 12 | - |
| FS5 NA | 17 | - | 22 | 2 | 18 | 20 | 13 | 19 | 11 | 13 | 15 | - |
| FS5 MSA (small colony) | 15 | 17 | 23 | 1 | 18 | 15 | 18 | 17 | 15 | 12 | 13 | - |
| FS5 MSA (Large colony) | 17 | 15 | 19 | 7 | 21 | 18 | 16 | 17 | 15 | 12 | 13 | 5 |

Table 8. Antibiotic susceptibility

| Sample | SLB | CTX | CIP | NET | CF | CPZ | AN | CFP | PIT | BA | MF | G |
|----------------------|-----|-----|-----|-----|----|-----|----|-----|-----|----|----|----|
| FS2; EMB | - | 14 | 19 | 15 | 14 | 13 | 16 | 13 | 23 | 10 | 18 | 16 |
| FS2 NA big colony | 22 | 25 | 25 | 20 | 21 | 20 | 20 | 29 | 20 | 20 | 18 | 20 |

Table 9 antibiotic susceptibility

| Sample | SLB | CTX | CIP | NET | CF | CPZ | AN | CFP | PIT | BA | MF | G |
|------------|-----|-----|-----|-----|----|-----|----|-----|-----|----|----|----|
| FS3 EMB | 20 | 26 | 35 | 25 | 26 | 25 | 30 | 25 | 23 | 15 | 25 | 20 |
| F5NASC | 17 | - | 20 | 1 | 20 | 16 | 15 | 20 | 17 | 18 | 15 | 1 |

3.6.3 Antibiotic susceptibility of gram-negative rods

The above Fig 6 represents Graphical presentation of antibiotic susceptibility of gram-negative rods, for sample FS2 EMB zone of inhibition is high for antibiotic Piperacillin and for sample FS2 NA zone of inhibition is high for antibiotic Cefoperazone. The isolate vibrio spp is resistant to Sulbactam and isolate Klebsiella pneumonia is sensitive to all antibiotics.

3.6.4 Antibiotic susceptibility of gram negative cocci

The above Fig 7 determines the Graphical presentation of antibiotic susceptibility of gram-negative cocci, FS3 EMB having zone of inhibition high for antibiotic Ciproflaxin, for FS5 NA having zone of inhibition high for antibiotic Ciproflaxin, cefotaxime and cefoperazone. These isolate *Providencia vermicola* is resistant to antibiotic ceftriaxone, whereas *Pantoea dispersa* is sensitive to all the antibiotics that are used in current study.

In our current study *Pantoea dispersa* has shown sensitive to all antibiotics, research study by [16] states that *Pantoea dispersa* is a multidrug resistant organism.

In current study *Providencia vermicola* is resistant to Cefotaxime and drug of choice for the diseases that caused by this organism is Ciprofloxacin and Cefoperazone.

Hence the drug of choice will depend on the zone of inhibition, as the zone of inhibition increases the sensitivity of the organism towards particular antibiotic also.

3.7 Determination of the Microbiological Quality of Different Type of Street Food

The FS 1 and 4 which is Manchuria and bonda sample consists *Lactobacillus delbrueckii*. *Lactobacillus delbrueckii* is determined to be the causative microorganism for urinary tract infection [17].

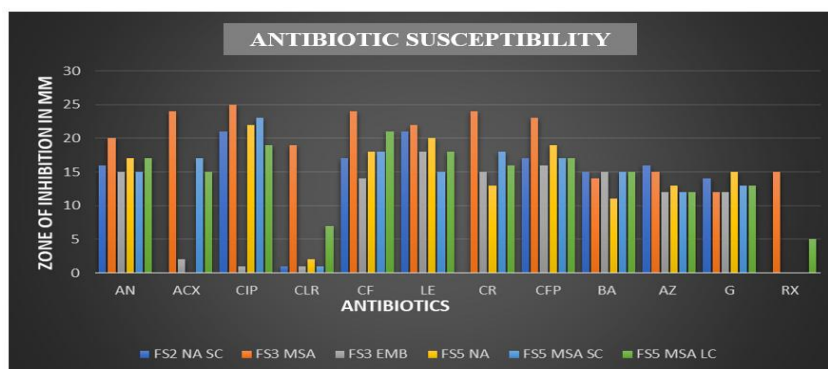


Fig. 5. Graphical presentation of antibiotic susceptibility of gram-positive cocci

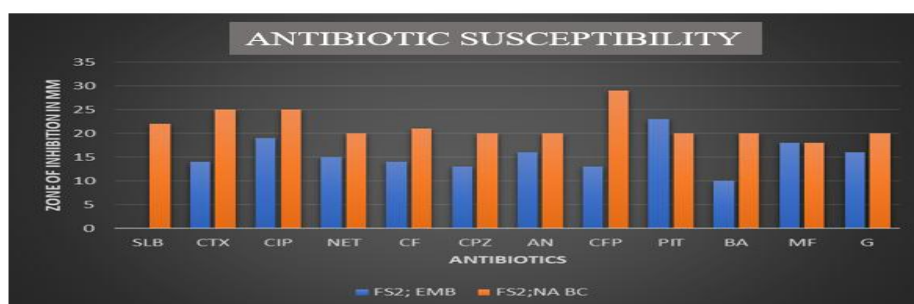


Fig. 6. Graphical presentation of antibiotic susceptibility of gram-negative rods

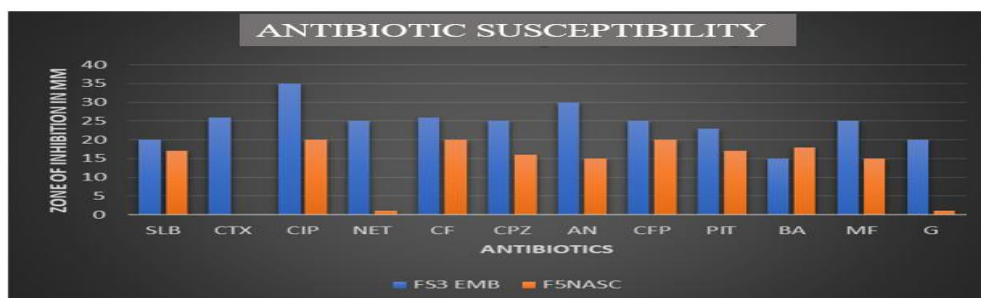


Fig. 7. Graphical presentation of antibiotic susceptibility of gram-negative cocci

In FS 2 (Ragada) we have observed organism *Lactobacillus delbrueckii*, pathogenic species which are Vibrio species which causes diarrhea, vibriosis and septicemia, *Enterococcus faecalis* causes diarrhea, nausea and Urinary tract infections, *Klebsiella pneumonia* it causes bloodstream infections, meningitis, and urinary tract infections. These pathogens are harmful for our system. Hence, this street food is harmful for consumption.

In FS 3 (samosa) we observed *Lactobacillus delbrueckii*, pathogenic species which are *Staphylococcus aureus* it causes diseases like staph food poisoning- it is a gastrointestinal illness, pneumonia and blood stream infections

and *Pantoea dispersa* [18] it causes respiratory infections, neonatal sepsis, and bloodstream infections; hence this street food is harmful for consumption.

In FS5 we observed only *Staphylococcus aureus* which causes food poisoning, pneumonia and blood stream infection, *Providencia vermicola* produce bacterial urease, an important virulence factor associated for formation of urinary tract stones. Hence the street food Pani-Puri is harmful for consumption.

These organisms are found and observed by basic testing methods present in our Laboratory.

Hence these 5 street foods are analyzed to be not good due to presence of pathogenic organisms as they cause diseases like food poisoning which in severe cases may lead to death, and various illness, hence these street foods are harmful to health.

4. CONCLUSION

The organisms isolated from the different street food samples comprised of Gram-negative rods (*Klebsiella pneumonia* and *Vibrio* spp), Gram positive cocci (*Staphylococcus aureus*, *Enterococcus faecalis*) and Gram-positive rods (*Lactobacillus* spp). The organisms are identified by 16s r RNA typing *Pantoea dispersa*, *Providencia vermicola* (gram negative cocci). These isolates of street food were identified by biochemical test and molecular test. Antibiotic susceptibility test is done by Kirby-Bauer method, where most of the isolates were resistant to Ampiclox, Cefotaxime, Cefuroxime, Sulbactam. These multidrug resistant foods borne bacteria cause a public health hazard. The microbiological quality of these five-street food is found to be not good due to presence of pathogenic species. The quality of these street food is observed to be poor, due to lack of hygiene conditions while preparing the food, ingredients and the water that are used for preparation may affect quality of the food, due to lack of maintenance of the utensils, surrounding area near food preparation, and due to air borne microorganisms [19].

Hence the microbiological quality of street food can be increased by maintaining hygienic conditions, use of clean water for preparation, preventing the contamination of food by pathogenic organisms.

Acknowledgements

I would first like to thank the college management for providing facility to do project in these tough times. I wish to express my sincere gratitude to Dr. Shailaja Raj, HOD of the Department of Microbiology at St. Francis College for Women, Begumpet, for providing the opportunity to carry out my project work.

My heartfelt thanks to all the faculty members for their help and support.

I would also like to thank the lab assistants of the Microbiology Department for their cooperation

and for providing with all the equipment required for the project.

I would sincerely thank Pathcare Lab Pvt.Ltd and Synteny Lifesciences Pvt.Ltd for their logistic support in doing my experiments of Project.

Many thanks to my family and friends for their constant encouragement and support throughout.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Happy, Hedayetul & Alam, Md & Mahmud, Sumodro, Imran, Sayeed, Rony, Mehadi & Azim, Mohd et al. Isolation, Identification and Characterization of Gram-Negative Bacteria from Popular Street Food (Chotpoti) at Savar Area, Dhaka, Bangladesh. Open Access Library Journal. 2018;05:1-11. DOI:10.4236/oalib.1104986.
2. Sezgin, Aybuke. Street food consumption in terms of the food safety and health. Journal of Human Sciences. 2016;13: 4072. DOI:10.14687/jhsv13i3.3925.
3. Eromo, Temesgen, Tassew, Haimanot, Daka, Deresse, Kibru, Gebre. Bacteriological Quality of Street Foods and Antimicrobial Resistance of Isolates in Hawassa, Ethiopia. Ethiopian Journal of Health Sciences. 2016;26:533. DOI:10.4314/ejhsv26i6.5.
4. Ma, Lihua & Chen, Hong, Yan, Huizhe & Wu, Lifeng & Zhang, Wenbin. Food safety knowledge, attitudes, and behavior of street food vendors and consumers in Handan, a third-tier city in China. BMC Public Health. 2019;19. DOI:10.1186/s12889-019-7475-9.
5. Khandoker A, Islam, Md, Rahman, Md. Mahafujur & Husna, A, Das S, Khatun, Mst. Bacterial contamination of street-vended spicy puffed-rice sold at Bangladesh Agricultural University campus. Bangladesh Veterinarian. 2015;31. DOI:10.3329/bvet.v31i1.22839. Available:https://www.biomerieux-usa.com
6. Vading M, Nauc ler P, Kalin M, Giske CG. Invasive infection caused by *Klebsiella*

- pneumoniae is a disease affecting patients with high comorbidity and associated with high long-term mortality; 2018.
7. Bayot, Marlon L, Bradley N. Bragg. "Antimicrobial susceptibility testing." StatPearls [Internet]; 2020.
 8. Chanda, Warren & Manyepa, Mespa & Chikwanda, Ephraim & Daka, Victor & Chileshe, Justin & Tembo, Mathias & Kasongo, et al. Evaluation of antibiotic susceptibility patterns of pathogens isolated from routine laboratory specimens at Ndola Teaching Hospital: A retrospective study. PLOS ONE. 2019;14. e0226676. DOI:10.1371/journal.pone.0226676.
 9. Poonia, Shubra, Singh T, Tsering, Dechen. Antibiotic Susceptibility Profile of Bacteria Isolated from Natural Sources of Water from Rural Areas of East Sikkim. Indian Journal of Community Medicine: Official publication of Indian Association of Preventive & Social Medicine. 2014;39: 156-60. DOI:10.4103/0970-0218.137152.
 10. Jo, Jay-Hyun, Kennedy, Elizabeth & Kong, Heidi. Research Techniques Made Simple: Bacterial 16S Ribosomal RNA Gene Sequencing in Cutaneous Research. Journal of Investigative Dermatology. 2016;136:e23-e27. DOI:10.1016/j.jid.2016.01.005.
 11. Darbro, Benjamin & Petroelje, Brian & Doern, Gary. Lactobacillus delbrueckii as the Cause of Urinary Tract Infection. Journal of clinical microbiology. 2008;47: 275-7. DOI:10.1128/JCM.01630-08.
 12. Kau, Andrew, Martin, Steven, Lyon, William, Hayes, Ericka, Caparon, Michael, Hultgren, Scott. Enterococcus faecalis Tropism for the Kidneys in the Urinary Tract of C57BL/6J Mice. Infection and immunity. 2005;73: 2461-8. DOI:10.1128/IAI.73.4.2461-2468.2005.
 13. Bush, Larry, Abrams, Barry, Beall, Anne & Johnson, Caroline. Index Case of Fatal Inhalation Anthrax Due to Bioterrorism in the United States. The New England journal of medicine. 2001;345: 1607. DOI:10.1056/NEJMoa012948.
 14. Vading M, Naucler, Pontus Kalin, M, Giske, C. Invasive infection caused by Klebsiella pneumoniae is a disease affecting patients with high comorbidity and associated with high long-term mortality. PLOS ONE. 2018;13:e0195258. DOI:10.1371/journal.pone.0195258.
 15. Baker-Austin, Craig, Oliver, James, Alam, Munirul, Ali, Afsar, Waldor, Matthew & Qadri, Firdausi et al. Vibrio spp. infections. Nature Reviews Disease Primers. 2018;4. DOI:10.1038/s41572-018-0005-8.
 16. Ramkumar RM Mani, Ravi, Chidambaram, Jayaseelan, Rahuman, Abdul & Anandhi, M, Rajthilak, Chandrasekaran et al. Description of Providencia vermicola isolated from diseased Indian major carp, Labeo rohita. Aquaculture. 1822;420–421. 193–197. DOI:10.1016/j.aquaculture.2013.11.010.
 17. Mehar, Veerendra, Yadav, Dinesh, Sanghvi, Jyoti, Gupta, Nidhi, Singh, Kuldeep. Pantoea dispersa: An unusual cause of neonatal sepsis. The Brazilian journal of infectious diseases: an official publication of the Brazilian Society of Infectious Diseases. 2013; 17. DOI:10.1016/j.bjid.2013.05.013.
 18. Panditrao Mridul, Panditrao, Minnu.. Pantoea dispersa: Is it the Next Emerging "Monster" in our Intensive Care Units? A Case Report and Review of Literature. Anesth Essays Res 2018;12:963-6. Anesthesia, Essays and Researches. 12. 963-6. DOI:10.4103/aer.AER_147_18.
 19. Darbro, Benjamin, Petroelje, Brian, Doern, Gary. Lactobacillus delbrueckii as the Cause of Urinary Tract Infection. Journal of clinical microbiology. 2008;47:275-7. DOI:10.1128/JCM.01630-08.

© 2022 Manasa and Thomas; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/86720>