

International Journal of Plant & Soil Science

Volume 34, Issue 24, Page 930-936, 2022; Article no.IJPSS.95860 ISSN: 2320-7035

Efficacity of Arbuscular Mycorrhizal Inoculations on Tree Species Germination and Growth

Mamta Khaiper^{a*}, Pawan Kumar Poonia^a, Sunil Kumar Dhanda^a, K. S. Ahlawat^a, Rakesh Chugh^b, Anil Kumar^c, Preety Verma^b, Monika Jangra^a, Arun Kumar Kagra^a and Karishma Nanda^a

 ^a Department of Forestry, College of Agriculture, Chaudhary Charan Singh Haryana Agricultural University, Hisar-125004, India.
^b Department of Plant Pathology, College of Agriculture, Chaudhary Charan Singh Haryana Agricultural University, Hisar-125004, India.
^c Krishi Vigyan Kendra, Yamunanagar-135001, Haryana, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJPSS/2022/v34i242720

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/95860

Review Article

Received: 27/10/2022 Accepted: 30/12/2022 Published: 31/12/2022

ABSTRACT

Researchers are very concerned about addressing or managing the growing problem of global warming or climate change. Global warming is caused by greenhouse gases, of which carbon dioxide (CO_2) alone contributes 60%. By absorbing carbon dioxide (CO_2) during photosynthesis and naturally storing it as biomass, trees serve as a sink for CO_2 because trees have the ability to store carbon and can lessen the effects of climate change and global warming, planting trees or restoring forests may be a solution to this growing problem. This review paper can be helpful since it

Int. J. Plant Soil Sci., vol. 34, no. 24, pp. 930-936, 2022

^{*}Corresponding author: E-mail: mamtakhaiper247@gmail.com;

discusses the interaction between arbuscular mycorrhizal fungi (AMF) and the roots of trees, which benefits the plant for better germination and growth. dormancy of seeds or a delay in growth can be identified as a disadvantage in a plantation program. With the hope that this information would advance the field of research, we have concentrated on mycorrhizal association and its impact on different species of trees in this work. It can be useful for tree species that have lower biomass or slower growth.

Keywords: Global warming; photosynthesis; arbuscular mycorrhizal fungi (AMF); dormancy and biomass.

1. INTRODUCTION

Seed dormancy, a physiological phenomenon in plants that can be brought on by internal or external stimuli, prevents seeds from sprouting even under ideal circumstances. Hard seed primitive coats. underdeveloped embryos, embryos, and inhibitory compounds can all contribute to seed dormancy [1]. Some species' seeds cannot fully germinate because the embryo is restricted by the structures in its environment. Embryos removed from these seeds are not dormant; this phenomenon is known as seed coat augmented dormancy or physical dormancy. In seeds of other species with underdeveloped embryos, a second kind of dormancy known as morphophysiological dormancy is also present, but it also has a physiological component. As a result, these seeds need a treatment to break their dormancy. such as a specific mix of warm and/or cold Large-scale cultivation stratification [2]. is hampered in forestry and home garden plantation initiatives by delayed nursery development and deprived seed germination [3-5].

Bonner [6] stated that germination is defined as "the resumption of active growth in an embryo which results in its emergence from the seed and development of those structures essential for plant development". It is the culminating event of seed maturation, the establishment of the seedling [7]. Pre-sowing treatments of seed are crucial for constructing a nursery of a specific species in order to forecast the largest number of high-quality seedlings with the least amount of money, time, and labor [8]. To overcome this dormancy in forest seeds, various pre-sowing seed treatments are used.

One of the soil microorganisms that makes up the fundamental components of a good soil-plant connection is the arbuscular mycorrhizal fungus (AMF). By reducing stresses through symbiosis, the mycorrhizal fungus is an essential part of the soil microbial community that promotes plant growth and survival [9]. The advantages that these fungi provide their host include improved phosphorus uptake [10], increased nitrogen absorption [11], the production of plant growth hormones [12], root defense against soil-borne diseases [13], and improved plant growth and productivity [14].

The ability of the AM fungus to colonize roots and act as a biofertilizer and bioprotectant to protect plants against parasitic nematodes and fungi as well as to boost plant development and yield has been demonstrated [15]. (Wei et al., 2016). AMF is a form of symbiotic soil fungi that colonizes the roots of about 80% of vascular plants, according to Vierheirlig 16]. It is one of the essential soil microbes that contribute to a balanced soil-plant system [17]. They improve plant fitness and soil quality [18]. Since it has been demonstrated that the AMF colonizes its roots in prodigious quantities, the neem tree species is extremely dependent on mycorrhizal fungus [19]. In order to lessen the negative effects of slow growth and low germination rates in tree species, this review will analyze the benefits, drawbacks, and role of seed treatment with arbuscular mycorrhizal fungi. It will also consider potential AMF that may be applied through seed treatment or inoculation for better growth.

2. EFFECT OF MYCORRHIZA ON GROWTH AND GERMINATION OF DIFFERENT TREE SPECIES

Although arbuscular mycorrhiza fungi are naturally present in the roots of higher tree species, this literature includes the effect of some AM fungi on the growth and germination of tree species (Table 1). Khaiper et al. [20] conducted an experiment on Melia azedarach, in which she treated seeds with different pre-sowing treatments and then sow these seeds into soils inoculated with *Glomus mosseae* and reported that seeds showed maximum seed germination percentage in seeds treated with cow dung slurry for 6 days within the soil inoculated with Glomus mosseae as compared to control. Shukla et al. [21] investigated the effect of AM fungi (*Glomus cerebriforme* and *Acaulospora scrobiculata*) on two multipurpose tree species i.e., Eucalyptus tereticornis and Albizia procera and found that shoot length, dry weight and nutrient uptake increased significantly after inoculations with AM fungi. The best results were obtained with *G. cerebriforme* in both the tree species than other inoculants.

Muthukumar et al. [22] concluded that the seedlings of Casuarina equisetifolia inoculated with Glomus aggregatum and Acaulospora scrobiculata had significantly higher plant growth and measured. nutrient parameters Nevertheless, the response was higher for seedlings inoculated with G. aggregatum compared to those inoculated with Α scrobiculata.

Basumatary et al. [23] reported that an increase in length, diameter, circumference and biomass yield along with nutrient availability in the rhizosphere i.e. Nitrogen, Phosphorus, Potassium and carbon in Rubber tree (*Havea brasiliensis*) seedlings inoculation with Acaulospora sp. and Glomus sp. over control.

Singh et al. [24] revealed that the maximum biomass accumulation, phosphorous and nitrogen content in Acacia nilotica seedlings were achieved under soil treated with *Glomus mosseae* along with phosphate and nitrogenous fertilizers.

Reena and Bagyaraj [25] conducted an experiment on effect of 13 different VAM fungi on the seedlings of Acacia nilotica and Calliandra calothyrsus and reported that inoculated plants had greater plant height. leaf number, stem girth, biomass and phosphorus zinc content. They also had more and mycorrhizal root colonization, spores and external hyphae in soil. A. nilotica seedlings responded best at inoculation with G. mosseae followed by G. caledonicum whereas, C. calothyrsus responded best with G. velum and G. merredum.

Herrmann et al. [26] revealed that the management of Arbuscular mycorrhizal fungi (AMF) would contribute to maintaining or restoring soil fertility, leading to a better tree growth and optimized latex yield of Rubber trees (*Hevea brasiliensis*).

Chen et al. [27] reported that Liqurice (*Glycyrrhiza uralensis*) plant inoculated with *G. mosseae* improved the features of the root system and increased in photosynthetic efficiency. The uptake of P and K in plant was also increased when inoculated with *G. mosseae*.

Filho et al. [28] found that the application of *Rhizofagus clarum* and *Glomus etunicatum* in soil grown with star fruit (*Averrhoa carrambola* L), provided increments of 49% in height, 99% in dry matter production and 86, 129 and 108% in the content of N, K and calcium respectively, in relation to the control.

Jamaluddin and Shukla [29] conducted a laboratory experiment with three treatments *viz.*, AM fungi (*Glomus mosseae*, *Acaulospora* sp. and *Gigaspora* sp.) + root exudates, AM fungi alone and root exudates alone. Out of which the treatment containing AM fungi + root exudates showed maximum colonization, phosphatase enzyme activity, phosphorus uptake and leaf protein as compared with control and other treatments under studied.

Hamidi et al. [30] studied the effect of endomycorrhizae *i.e.Glomus etunicatum* on the seedlings of cork oak plants (*Quercus suber*) and observed that plants inoculated with mycorrhizae showed the stimulatory effect on aerial part, aerial part weight, average root length and average fresh weight and the leaves number as compared to control.

Jha et al. [31] studied the effect of AM fungi namely *Glomus* fascicualtum, *Glomus* aggregatum, *Glomus* cerebriforme, *Glomus* diaphanum and *Glomus* etunicatum on *Dalbergia* sissoo, Acacia procera and Acacia nilotica seedlings and revealed that the application of AM fungi improves the uptake of phosphorus, minor elements and water and enhances plant growth and resistance to root diseases as compared to non-mycorrhizal plants.

An experiment conducted by Young [32] showed the effect of single and mixed inoculations with phosphorus solubilizing bacteria and VAM fungi on the growth of Leucaena leucocephala, Acacia confuse. mangium Liquidamber Α. and formosana. Growth of L. leucocephala tree increased by 22-99% with VAM fungal inoculation. The growth of Acacia confuse (14-63%), A. mangium (7-88%) and Liquidamber formosana (24-280%) promoted by mixed inoculants.

Sumana and Bagyaraj [33] conducted a experiment and used eight different VAM fungi (Acaulospora Gigaspora laevis. margarita. Glomus caleldonicum, Glomus fasciculatum, Glomus intraradices, Glomus lepototichum, Glomus macrocarpum and Glomus mosseae) for selecting best symbiont for Dalbergia latifolia and reported that inoculated plants had greater plant height, stem girth, dry weight and phosphorus content when compared to noninoculated plants. D. latifolia responded best to inoculation with Glomus leptotichum as well as

Glomus fasciculatum as compared to the other AM fungi.

Youpensuk et al. [34] conducted an experiment of Seedlings Macaranga denticulate on inoculated with spores of fungi (Glomus species. Glomus fasciculatum. Acaulospora species and mixed species of AM fungi) in pot and concluded that Nutrient contents (N, P and K) of plant inoculated with Acaulospora species or mixed species of AM fungi were higher than plants inoculated with Glomus spp. and G. fasciculatum.

S. No.	Tree spp.	Mycorrhizal spp.	Description	Reference
	Melia azedarach	Glomus mosseae	The germination percentage (75.87) and other such as mean daily germination, germination value and speed of germination were recorded highest in treatment with cow dung slurry for 6 days + <i>Glomus mosseae</i> .	Khaiper et al. [20]
	Eucalyptus tereticornis and Albizia procera	Glomus cerebriforme and Acaulospora scrobiculata	Shoot length, dry weight and nutrient uptake increased significantly after inoculations with AM fungi. Best results were obtained with <i>G. cerebriforme</i> in both the tree species then other inoculants	Shukla et al. [21]
	Azadirachta indica	<i>G. intraradices</i> and <i>G. gegosporum</i>	Neem (<i>Azadirachta indica</i>) seedlings and inoculate with <i>G.</i> <i>intraradices</i> and <i>G. gegosporum</i> have increase mycorrhizal colonization, greater plant height, leaf area and number of root, biomass, phosphorus, nitrogen, collar diameter and potassium content and seedling quality.	Muthukumar et al. [22]
	Casuarina equisetifolia	Glomus aggregatum and Acaulospora scrobiculata	Higher plant growth and nutrient parameters measured. Nevertheless, the response was higher for seedlings inoculated with <i>G. aggregatum</i> compared to those inoculated with <i>A. scrobiculata</i> .	Muthukumar et al. [35]
	Havea brasiliensis	Acaulospora sp. and Glomus sp.	Increase in length, diameter, circumference and biomass yield along with nutrient's availability in rhizosphere <i>i.e.</i> Nitrogen, Phosphorus, Potassium and carbon in seedlings inoculated with <i>Acaulospora</i> sp. and <i>Glomus</i> sp. over control.	Basumatary et al. [23]
	Acacia nilotica	Glomus mosseae	Maximum biomass accumulation, phosphorous and nitrogen content in seedlings were achieved under	Singh et al. [24]

S. No.	Tree spp.	Mycorrhizal spp.	Description	Reference
			soil treated with <i>Glomus mosseae</i> along with phosphate and nitrogenous fertilizers.	
	Hevea brasiliensis	Arbuscular mycorrhizal fungi	Arbuscular mycorrhizal fungi (AMF) would contribute to maintaining or restoring soil fertility, leading to a better tree growth and optimized latex yield of Rubber trees (<i>Hevea</i> <i>brasiliensis</i>).	Herrmann et al. [26]
	Glycyrrhiza uralensis	G. mosseae	Plant inoculated with <i>G. mosseae</i> improved the features of the root system and increased in photosynthetic efficiency. The uptake of P and K in plant was also increased when inoculated with <i>G.</i> <i>mosseae</i> .	Chen et al. [27]
	Averrhoa carrambola L	Rhizofagus clarum and Glomus etunicatum	Increments of 49% in height, 99% in dry matter production and 86, 129 and 108% in the content of N, K and calcium respectively, in relation to the control.	Filho et al. [28]
	Quercus suber	Glomus etunicatum	Plants inoculated with mycorrhizae showed the stimulatory effect on aerial part, aerial part weight, average root length and average fresh weight and the leaves number as compared to control.	Hamidi et al. [30]

Khaiper et al.; Int. J. Plant Soil Sci., vol. 34, no. 24, pp. 930-936, 2022; Article no.IJPSS.95860

3. CONCLUSION

Trees act as a sink for carbon dioxide (CO2) by absorbing it during photosynthesis and naturally storing it as biomass, which helps to mitigate the consequences of climate change and global warming. the relationship that develops between arbuscular mycorrhizal fungus (AMF) and tree roots, enabling the plant to germinate and grow more successfully. A disadvantage in a plantation program can be the dormancy of seeds or a delay in growth. We have focused on mycorrhizal association and its effects on several tree species in this work with the aim that this information would progress the field of research for tree species with lower biomass or slower growth.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Mousavi L, Wan Ishak WR, Mousavi M. Evaluation of physicochemical methods for dormancy breakage and germination of *Datura stramonium* seeds. Journal of Chemical Health Risks. 2019;9:217-224.

- 2. Baskin CC, Baskin JM, Guerrant EO, Havens K, Maunder M. Determining dormancy-breaking and germination requirements from the fewest seeds. Ex situ plant conservation: supporting species survival in the wild. 2004;162-179.
- 3. Alamgir M, Hossain MK. Effect of presowing treatments on germination and initial seedling development of *Albizia saman* in the nursery. Journal of Forestry Research. 2005;16:200-204.
- 4. Azad MS, Islam MW, Matin MA, Bari MA. Effect of pre-sowing treatment on seed germination of *Albizia lebbeck* (L.) Benth. South Asian Journal of Agriculture. 2006;12:32-34.
- Azad MS, Matin MA, Islam MW. Effect of pre-sowing treatment on seed germination of Lohakath (*Xylia kerrii Craib & Hutch*). Khulna University Studies. 2006; 22:33-36.
- Bonner FT. Glossary of seed germination terms for tree seed workers. US Department of Agriculture, Forest Service, Southern Forest Experiment Station; 1984.

- Kramer P, and Kozlowski T. Physiology of woody plants. *New York Academic Press*, 1979; 811.
- 8. Das N. The effect of different pre-sowing treatments on the germination of *Aquilaria agallocha* and *Shorea robusta* seeds in the nursery. Indian Forester. 2015;141:285-292.
- Sylvia DM, Williams SE. Vesiculararbuscular mycorrhizae and environmental stress. Mycorrhizae in sustainable agriculture. 1992;54:101-124.
- Goussous SJ, Mohammad MJ. Comparative effect of two arbuscular mycorrhizae and N and P fertilizers on growth and nutrient uptake of onions. International Journal of Agriculture and Biology. 2009;11:463-467.
- 11. Rotor AV, Delima PC. Mycorrhizal association, N fertilization and biocide application on the efficacy of bio-N on Corn (Zea mays L.) growth and productivity. *International Journal of Scientomeric Research* 2010;2:267-290.
- 12. Herrera-Medina MJ, Steinkellner S, Vierheilig H, Ocampo Bote JA, Garcia Garrido J. Abscisic acid determines arbuscule development and functionality in the tomato arbuscular mycorrhiza. New Phytologist. 2007;175:554-564.
- Bakhtiar Y, Yahya S, Sumaryono W, Sinaga MS, Budi SW, Tajuddin T. Isolation and identification of Mycorrhizosphere bacteria and their antagonistic effects towards *Ganoderma boninense* in vitro. Microbiology Indonesia. 2010;4:9-9.
- 14. Duponnois R, Colombet A, Hien V, Thioulouse J. The mycorrhizal fungus Glomus intraradices and rock phosphate amendment influence plant growth and microbial activity in the rhizosphere of *Acacia holosericea*. Soil biology and Biochemistry. 2005;37:1460-1468.
- Berruti A, Lumini E, Balestrini R, Bianciotto V. Arbuscular mycorrhizal fungi as natural biofertilizers: let's benefit from past successes. Frontiers in microbiology. 2016;6:1559.
- 16. Vierheilig, H. Regulatory mechanisms during the plant–arbuscular mycorrhizal fungus interaction. *Canadian Journal of Botany*. 2004; 82:1166–1176.
- 17. Budi SW, Bakhtiar Y, May NL. Bacteria associated with arbuscular mycorrhizal spores Gigaspora margarita and their potential for stimulating root mycorrhizal colonization and neem (*Melia azedarach*

Linn) seedling growth. Microbiology Indonesia. 2012;6:180-188.

- Barea JM, Azcón R and Azcon-Aguilar C. Mycorrhizosphere interactions to improve plant fitness and soil quality. Antonie Van Leeuwenhoek. 2002:81(1-4):343-351.
- Habte M, Muruleedhara BN, Ikawa H. Response of neem (*Azadirachta indica*) to soil P concentration and mycorrhizal colonization. Arid Land Research and Management. 1993;7:327-333.
- Khaiper M, Dhanda, SK, Ahlawat, KS, Chugh, R. S., Jangra, M. and Verma, P. Effect of pre-sowing treatments on drupe germination of *Melia azedarach* at nursery stage. Agricultural Mechanization in Asia, Africa and Latin America. 2021;52:4569-4577.
- Shukla A, Kumar A, Jha A, Rao DV. Phosphorus threshold for arbuscular mycorrhizal colonization of crops and tree seedlings. Biology and Fertility of Soils. 2012;48:109-116.
- 22. Muthukumar T, Uma E, Priyadharsini P. Arbuscular Mycorrhizal Fungal Strains and Soil Type Influence Growth, Nodulation, and Nutrient Uptake of *Casuarina equisetifolia*. Microbiological Research In Agroecosystem Management. 2013;35-52.
- 23. Basumatary N, Parkash V, Tamuli AK, Saikia AJ, Teron R. Arbuscular mycorrhizal inoculation affects growth and rhizospheric nutrient availability in *Hevea brasiliensis* (Willd. ex A. Juss.) Mull. Arg. clones. International Journal of Current Biotechnology. 2014;2:14-21.
- 24. Singh KP, Yadav R, Kumari P, Bhadauria S. Biomass production, phosphorus and nitrogen content in *Acacia nilotica* under am fungi and nutrient treated soil. Indian Forester. 2014;140:489-493.
- 25. Reena J, Bagyaraj DJ. Response of *Acacia nilotica* and *Calliandra calothyrsus* to different VA mycorrhizal fungi. Arid Land Research and Management. 1990;4:261-268.
- Herrmann L, Bräu L, Robin A, Robain H, Wiriyakitnateekul W, Lesueur D. High colonization by native arbuscular mycorrhizal fungi (AMF) of rubber trees in small-holder plantations on low fertility soils in North East Thailand. Archives of Agronomy and Soil Science. 2016;62:1041-1048.
- 27. Chen M, Yang G, Sheng Y, Li P, Qiu H, Zhou X, Huang L, Chao Z. *Glomus mosseae* inoculation improves the root

system architecture, photosynthetic efficiency and flavonoids accumulation of liquorice under nutrient stress. Frontiers in plant science. 2017;8:1-10.

- 28. Filho JAV, Mendonça FMS, Martins MA, dos SPC, Cordeiro CAJ. Arbuscular mycorrhizal fungi and phosphate fertilization on star fruit tree seedlings. *Revista brasileira de Ciencias agrárias* 2017;12:14-19.
- 29. Jamaluddin M, Shukla R. Effect of root exudates on colonisation of am fungi in *Jatropha* curcas. Indian Forester. 2012;138:113-115.
- Hamidi O, Talbi Z, Chliyeh M, Touhami AO, Selmaoui K, Benkirane R, Douira A. Effect of Endomycorrhizal Inoculation on the Young Cork Oak Plants (*Quercus suber*) Growth. Annual Research & Review in Biology. 2017;13:1-11.
- Jha A, Kumar A, Shukla A, Chakravarty N. Response of four multipurpose tree species to arbuscular mycorrhizal inoculations. Indian Phytopathology. 2012; 65:297.

- Young CC. Effects of phosphorussolubilizing bacteria and vesiculararbuscular mycorrhizal fungi on the growth of tree species in subtropical-tropical soils. Soil Science and Plant Nutrition. 1990; 36:225-231.
- Sumana DA, Bagyaraj DJ. Selection of efficient VA-mycorrhizal fungi for inoculating Neem. In Proceedings of the National conference of Mychorrhiza. Singh S (ed) Barkatulla University, Bhopal 1999;10.
- Youpensuk S, Rerkasem B, Dell B, Lumyong S. Effects of arbuscular mycorrhizal fungi on a fallow enriching tree (*Macaranga denticulata*). Fungal Diversity. 2005;18:189-199.
- 35. Muthukumar T, Udaiyan K, Rajeshkannan V. Response of neem (*Azadirachta indica* A. Juss) to indigenous arbuscular mycorrhizal fungi, phosphate-solubilizing and asymbiotic nitrogen-fixing bacteria under tropical nursery conditions. Biology and Fertility of Soils. 2001;34:417-426.

© 2022 Khaiper et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/95860