



Seasonal Changes of Some Metabolites in *Hyoscyamus boveanus* (Dunal) Asch. & Schweinf – Saint Katherine, South Sinai, Egypt

Karima Mohamed El-Absy^{1*}

¹Eco-Physiology Unit, Plant Ecology and Ranges Department, Desert Research Center, Cairo, Egypt.

Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

Article Information

DOI: 10.9734/AJOB/2021/v13i230184

Editor(s):

(1) Prof. Jehad M. H. Ighbareyeh, Al-Quds Open University, Palestine.

Reviewers:

(1) Sergeeva Natalya Nikolaevna, Federal State Budget Scientific Institution, Russia.

(2) Nasiru Abdullahi, Bayero University Kano, Nigeria.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/75256>

Original Research Article

Received 12 August 2021
Accepted 27 October 2021
Published 01 November 2021

ABSTRACT

This study was conducted to investigate the effects of seasonal fluctuations on some metabolites and to explore the correlation between soil and plant analysis in *Hyoscyamus boveanus* (*H. boveanus*) at the Wadi El-Sheikh Awad in Saint Katherine, South Sinai, Egypt. Significant differences ($p < 0.05$) were observed for physical and chemical properties of the soil associated with *H. boveanus* during 0-20 and 20-40 cm depths, which increased most of them during the first depth. All chemical composition contents of *H. boveanus* are influenced significantly ($p < 0.05$) by seasons studied, except Na^+ , glycosides, total phenol, crude protein contents. A substantial increase in mineral composition (Na^+ , K^+ , P and Fe^{2+}), total alkaloids, glycosides, total phenol, proline, total carbohydrates and all photosynthetic pigments contents were recorded in *H. boveanus* during the summer season. While, the mineral composition (Ca^{2+} , Mg^{2+} , S, N and Cl^-), water content and crude protein contents appeared to be higher in the winter season. The relationships between soil and plant variables were delineated by performing the principal component analysis (PCA). The PC1 and PC2 displayed differences between the soil and plant variables, also, the variables Mg^{2+} , Cl^- , pH, EC, Ca^{2+} and K^+ in the soil associated with *H. boveanus* are variables with better chemical properties of the soil, which affect the plant distribution in Wadi El-Sheikh Awad during the two seasons. The PCA revealed high positive correlations among soil variables as well as among plant variables. Soil magnesium correlated highly and positively with

*Corresponding author: E-mail: karima.mohamed77@yahoo.com;

the plant variables i.e., crude protein, water content, Chl b, Chl a+b, carotenoid and total pigment contents. The pH, EC, and Ca^{2+} in soil were positively correlated with all chemical composition contents of *H. boveanus*. Some metabolites in *H. boveanus* were significantly increased during the summer season compared to the other season, due to the activation of plant physiological stress tolerance mechanisms.

Keywords: Seasonal changes; metabolites; PCA; *Hyoscyamus boveanus*.

1. INTRODUCTION

The wild populations of plant species are exposed to severe danger, which could lead to their extinction due to climate change and human activities in their natural habitats. In Egypt, South Sinai is arid to the extremely arid region and characterized by ecological uniqueness due to its diversity in terrestrial forms, geological and climatic structures, which resulted in a diversity of plant species, which are mainly characterized by the dispersal and dominance of shrubs, sub-shrubs, and tree paucity [1, 2, 3]. Ayyad et al. [4] reported that the South Sinai region was more diverse compared to the entire Sinai peninsula. South Sinai contains 472 plant species including 19 Egyptian endemic species, 115 of medicinal interest, and approximately 170 species used in folk medicine [5].

The family Solanaceae is considered one of the largest and most important families of flowering plants. It contains about 102 genera and about 2460 species in the global flora [6]. In Egyptian flora, according to Hepper [7], the family Solanaceae comprises about 25 genera and about 91 wild and cultivated species, while, this family comprises about eight genera and 30-33 wild species according to Boulos [8]. *Hyoscyamus* genus belonging to the family Solanaceae have 26 species in the flora of the world [9, 10]. While Boulos [11] mentioned that there are fifteen species distributed in North Africa and Western Europe to Central Asia. Seven species of *Hyoscyamus* genus are found in Egypt [8].

Hyoscyamus boveanus (Dunal) Asch. & Schweinf (*H. boveanus*) is a flowering plant belonging to the genus *Hyoscyamus*. It is a plant species endemic to Egypt. It was found that the optimum height range for this species to grow is between 1300 and 1650 meters above sea level. *H. boveanus* is vulnerable to extinction due to natural factors such as area dryness, climate changes and fragmentation inherent in its habitats, as well as human activities such as overgrazing, cutting, killing, in addition to the

dams and random human construction [12, 13]. All plant species of the genus *Hyoscyamus* which are abundant in deserts and on arid land are rich sources in tropane alkaloids, especially hyoscyamine and scopolamine contents [14, 15], thus these species have high medicinal importance [16], where are widely used as an analgesic, and an anti-spasmodic, mydriasis, and anti-cholinergic, as well as a sedative and painless in popular medicine [17].

Seasonal variations for the chemical composition or biochemical ingredients have been reported in various plant species by several researchers. Different seasons affect the different biochemical attributes of the plant species, as variation significantly was observed in the active ingredients of these plants, and this variation is due to differences in environmental variables such as rainfall, temperature, and other variables [18, 19]. The evaluation of the results concerning the seasonal variation supports a biosynthetic pathway in *Mentha pulegium* L. plants [20]. Seasonal variation on chemical composition in soil and plants of *Lycium showii* was also studied by Kamel and El-Absy [21] and concluded that significant differences were observed to represent genuine environmental variations on chemical composition during seasons, sites, locations, and their interactions.

Due to public concern about increasing soil productivity and crop input efficiency, there must understand a relationship between spatially diverse soil properties and fertility [22]. The principal component analysis (PCA) can be used to summarize the main sources of data variance between correlated variables, as the PCA is a multivariate statistical technique for dimensional reduction that uses correlated variables to recombine and define the orthogonal linear of variables [23]. PCA is one of the statistical tools that can be used to study the relationship between the chemical properties of the soil/plant or the relationship between them. Several studies investigating the relationship between soil properties and vegetation have been conducted using PCA [23, 24, 25, 26].

The major aim of this research was to Quantitative determination and study seasonal variations of some metabolites of *H. boveanus* plants growing under natural conditions in Saint Katherine, South Sinai, Egypt, as well as, to elucidate the relationship between soil chemical properties and plant chemical composition contents by principal component analysis.

2. MATERIALS AND METHODS

2.1 Study area and Plant Material

The field study was conducted in July 2019 and January 2020 in the Wadi El-Sheikh Awad in Saint Katherine, South Sinai, Egypt. The research area is located along an altitudinal gradient ranging from 1120 to 1140m above sea level, within longitude (E) 33° 39' 06.00" and latitude (N) 28° 39' 05.00" [27]. El-Sheikh Awad area is located outside the Ring Dyke at the northern end of Nabq Al Hawa, an area of mixed sandstone and metamorphic rocks with great exposure to slabs, shallow sand and gravel wadis and garden areas [28]. In Wadi El-Sheikh Awad (Table 1), the average air temperature was much higher in the summer than in the winter. While average relative humidity and rainfall increased with the winter than the summer.

H. boveanus is a stout and rare succulent perennial herb, 45 – 60 cm high, with an unpleasant smell. The full plant including the inflorescence spreads densely hairy. Stem erect, green, herbaceous, cylindrical, branched forming a rounded bush, solid. Leaves are petiolate, alternate, apex acute, margin entire. The flowers are crowded, bisexual, arranged in one-sided racemes, its color is white with purple blotches and stripes, filaments and anthers have cream color, it appears in late spring. Fruit is capsule deep brown, ellipsoid. Seeds flat at lateral faces, luster, yellowish-brown [14, 29, 30]. Reproduction by seeds occurs at the late of the summer.

2.2 Soil Analysis

The soil samples associated with *H. boveanus* were collected from three random points at the two successive depths 0-20 cm and 20-40 cm in the Wadi El-Sheikh Awad. Three replicates were taken from each sample and carried to the laboratory in closed tins to be used for soil analyses. Soil samples were air-dried before sieving, then sieved and used for mechanical analysis of soil particles as outlined by Jackson [31] and Rowell [32] for soil texture. Soil moisture content was estimated by the method described by Rowell [32]. For each soil sample, Electrical conductivity (EC) and pH value were carried out using soil-water paste, according to Jackson [33], EC was expressed as mmhos/cm. The mineral contents in soil i.e., chloride (Cl⁻), calcium (Ca²⁺), manganese (Mn²⁺), sodium (Na⁺) and potassium (K⁺) were determined using a saturation paste [34].

2.3 Plant Analysis

From Wadi El-Sheikh Awad, three fresh samples of *H. boveanus* were randomly collected during the summer (July, 2019) and the winter (January, 2020) seasons, and sealed in plastic bags, and stored at 4°C under dark conditions. Drying of collected plant materials was done in the oven at 70°C to a constant weight after which dried samples were milled to fine powder and stored in brown bags at room temperature pending chemical analyses. The plant water content is the difference between fresh weight (Wf) and dry weight (Wd) on the basis of fresh weight, and it can be calculated from the equation; water content% = [(Wf-Wd)/Wf]x100 [35]. Sodium (Na⁺), potassium (K⁺) and calcium (Ca²⁺), magnesium (Mg²⁺), chloride (Cl⁻), sulfur (S), iron (Fe²⁺), phosphorus (P) concentrations were determined by atomic absorption spectrophotometry (GBC Avanta E, Victoria, Australia) [36]. Whilst, the chlorides were estimated following the AgNO₃ method according

Table 1. Seasonal mean temperature, Relative humidity and Rainfall at Wadi El-Sheikh Awad in Saint Katherine, South Sinai, Egypt

Meteorological	Air	Relative	Rainfall
Seasons	Temperature (°C)	Humidity (%)	(mm)
Winter	12.92	59.47	0.91
Summer	30.14	69.39	0.00

to Jackson and Thomas [37] after extraction from the ashed powdered samples. Total nitrogen (N) content was determined using the micro-Kjeldahl method [38]. The photosynthetic pigments parameters were quantified spectrophotometrically, and using the wavelengths of 663, 645 and 470 nm, the chlorophyll a, chlorophyll b and total carotenoids were calculated by equations of Lichtenthaler [39], respectively. The total alkaloids, glycosides and the total phenols contents were determined following the methods described by Harborne [40], Hikino et al. [41] and Singleton and Rossi [42], respectively. The crude protein % was determined by multiplying the total nitrogen by 6.25 according to Allen [43]. The proline and the total available carbohydrate contents were determined according to the methods of Bates et al. [44] and Chaplin and Kennedy [45], respectively.

2.4 Statistical Analysis

To perform statistical analyses in this study, the XLSTAT version 2020.5.1.1075 software for Windows was used. The water content % in soil was treated statistically by applying the two-way ANOVA test. The Student's t-test was used to confirm whether the difference between depths or seasonal variations was significant for soil and plant analysis. The values of $p \leq 0.05$ were considered to be statistically significant [46]. The data of soil analysis are expressed as the mean (\pm standard error) of three replicates. Mean values of soil and plant measurements are shown in the tables or figures and a sign (*) to inform statistical significance for the reported differences. The principal component analysis was done using a computer software program PAST version 2.17c.

3. RESULTS AND DISCUSSION

3.1 Soil Analysis

The physical properties (%) for the soil associated with *H. boveanus* from the two depths in Wadi El-Sheikh Awad are given in Table 2. According to Student's t-test, significant differences in all mechanical properties were observed between the 0-20 and 20-40 cm depths except clay %. The same tendency was observed by Kamel and El-Absy [21] as well as El-Absy and Kamel [47] who reported that most physical properties were significantly affected by depths in *Teucrium polium* and *Lycium showii* species during different habitat conditions, respectively. Compared to other mechanical properties (%), coarse sand, followed by fine

sand was registered the highest percentage at the two depths in Wadi El-Sheikh Awad. Fine sand and silt plus clay showed significant differences in the simple alluvial fan vegetation as *H. boveanus* of southern Sinai [48]. Thus, the soil physical properties indicated that the soil associated with *H. boveanus* at the two depths in Wadi El-Sheikh Awad is characterized by sandy to fine gravelly in texture. Poor fertility conditions to sandy texture are associated with low inputs of organic matter in arid environments [49], as Wadi El-Sheikh Awad environment.

According to two-ways ANOVA, the water content % was significantly different ($P < 0.05$) between seasons and season x depths interaction, while, the difference between the two depths was not statistically significant (Fig. 1). Kamel and El-Absy [21] and El-Lamey [50] also noticed significant variations in the water content of *Retama raetam* and *Lycium showii* species between seasons at different depths under different environmental conditions, respectively. The values of water content % in the winter season were notably higher than in the summer season during the two depths. The seasonal effect can lead to beneficial changes in the water content of the soil, such as an increase in the availability of phosphorus content needed for plants nutrition [51]. The soil associated with *H. boveanus* was characterized by a high water content % in the surface layer (depth 0-20 cm). The seasonal variation was found in water content by Kamel and El-Absy [21] and Ahmed et al. [52] who reported maximum values in the winter season compared to the summer season, due to rainfall, which leads to normal plant growth. While, Larcher [53] attributed the reason to the increase in total ions accumulation due to increased soil salinity and soil moisture stress. Moustafa and Zayed [48] indicated that plant species as *H. boveanus* are richer in drier habitats, that is, in the absence of high moisture. Also they added the moisture gradient is complex and associated with many environmental factors, which are elevation, slope, climatic drought, soil texture and soil surface nature.

The chemical analysis comparison of the soil associated with *H. boveanus* between the two depths showed in Table 3. The Student's t-test showed that the chemical analysis of the soil associated with *H. boveanus* has significant differences ($P < 0.05$) between the 0-20 and 20-40 depths. This result was consistent with Kamel and El-Absy [21], El-Lamey [50] and El-Absy et al. [54].

Table 2. The values of mean and standard deviations (\pm SD) of physical properties at the soil associated with *H. boveanus* in the two depths during Wadi El-Sheikh Awad

Depths (cm)	Physical properties (%)							Soil Texture Class
	Very Coarse Sand	Coarse Sand	Medium Sand	Fine Sand	Very Fine Sand	Slit	Clay	
0-20	8.30 \pm 0.11	50.12 \pm 0.45	2.21 \pm 0.09	28.03 \pm 0.19	2.08 \pm 0.07	4.03 \pm 0.12	5.23 \pm 0.10	Sandy
20-40	6.72 \pm 0.08	46.11 \pm 0.31	4.52 \pm 0.06	30.40 \pm 0.15	3.21 \pm 0.03	3.90 \pm 0.10	5.14 \pm 0.08	Sandy
Probability	0.002*	0.004*	0.001*	0.003*	0.001	0.002*	NS	

According to Student's t-test, the asterisks (*) denote a significant difference ($p < 0.05$) between the two depths.

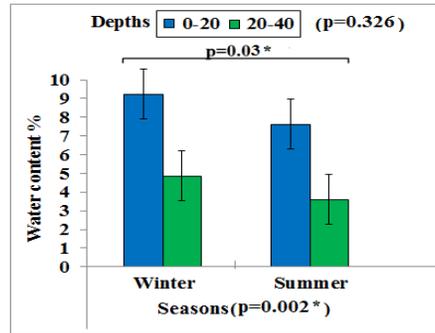


Fig. 1. Water content % at the soil associated with *H. boveanus* in the two depths and the two seasons during Wadi El-Sheikh Awad. According to two-way ANOVA test, the asterisks (*) denote a significant difference ($p < 0.05$) between the two depths and the two seasons as well as their interactions

Table 3. The values of mean and standard deviations (\pm SD) of chemical properties at the soil associated with *H. boveanus* in the two depths during Wadi El-Sheikh Awad

Depths (cm)	Chemical Properties						
	pH	EC (dS/m)	Cl ⁻ (meq/L)	Ca ²⁺ (meq/L)	Mg ²⁺ (meq/L)	Na ⁺ (meq/L)	K ⁺ (meq/L)
0-20	7.35 \pm 2.35	3.18 \pm 1.21	23.16 \pm 2.67	8.55 \pm 1.99	180.12 \pm 6.95	16.82 \pm 3.84	2.41 \pm 1.01
20-40	7.55 \pm 5.16	2.91 \pm 1.40	10.21 \pm 3.59	8.91 \pm 2.61	171.23 \pm 7.14	12.41 \pm 4.92	0.91 \pm 0.68
Probability	0.0001*	0.003*	0.0001*	0.002*	0.0001*	0.0001*	0.0001*

According to Student's t-test, the asterisks (*) denote a significant difference ($p < 0.05$) between the two depths.

The values of electrical conductivity (EC), Cl^- , Mg^{2+} , Na^+ and K^+ have increased significantly in 0-20 cm depth compared with their values in 20-40 cm depth. While, pH and Ca^{2+} values in 20-40 cm depth were significantly increased in comparison with the 0-20 cm depth. Of all chemical analysis in the soil associated with *H. boveanus* at the two depths in Wadi El-Sheikh Awad, Mg^{2+} content showed higher values relative to the other chemical analysis. Based on the pH values, the soil associated with *H. boveanus* in Wadi El-Sheikh Awad tended to be somewhat alkaline.

In general, our results agreed with Moustafa and Zayed [48] who mentioned that the soil associated with the *H. boveanus* plants was sandy to loamy sand, alkaline and not saline to slightly saline, also with a low content of basic nutrients and cation exchange capacity. Also, Labib et al. [55] reported that the soil properties indicated the weak organic nature of the study area in Saint Katherine. The results by Salama et al. [56] revealed significant differences for clay, calcium, magnesium, chlorides, electric conductivity, potassium and sulphates at the separated vegetation groups in South Sinai. Lower calcium, magnesium and potassium levels as well as total bases increased the acidic the soil [57]. Along the same lines, Zhao et al. [58] found that a higher concentration of calcium reduces the reaction of soil acids, in addition to affecting soil availability nutrient recycling and utilization. In agreement and disagreement with our results, they reported that soil pH and moisture content increased with increasing soil depth across the succession of vegetation cover, respectively. Often an inverse and adverse relationship was found between the high concentration of one cation in the soil and the availability of other cations for the plant to uptake [59]. As the availability of potassium depends on the availability of the relative quantities of potassium, calcium and magnesium in addition to the potassium content of the soil [60]. Hence, Laekemariam et al. [60] mentioned that the magnesium concentration is considered a potentially important factor in controlling potassium availability. The decrease in nutrient ion activities and the production of extreme ratios of $\text{Na}^+/\text{Ca}^{2+}$, Na^+/K^+ , $\text{Ca}^{2+}/\text{Mg}^{2+}$ and $\text{Cl}^-/\text{NO}_3^-$ in the soil solution may be due to the high concentrations of Na^+ and Cl^- [61].

3.2 Plant Analysis

The effects of different seasons on photosynthetic pigments contents are presented

in Fig. 2. All photosynthetic pigments contents of *H. boveanus* plant displayed statistically significant differences during the winter and summer seasons in Wadi El-Sheikh Awad, using Student's t-test. These results are in line with earlier reports by Uvalle Saucedo et al. [62] and Devi et al. [63] who find significant differences in all photosynthetic pigments under different seasons. The concentrations of chlorophyll a (chl a), chlorophyll b (chl b), chl a+b, chl a/b, total carotenoids and total pigment were higher values in the summer season than in the winter season. Total carotenoids content was the highest of *H. boveanus* plant, followed by chl b and chl a contents during the two studied seasons. In contrast, Uvalle Saucedo et al. [62] mentioned that chlorophyll was higher than the carotenoids in all plants. There was a perceptible trend for seasonal variation in all photosynthetic pigments contents and were higher in the summer season than in the winter season [64], which might have been related to seasonal water deficits in summer and extremely low temperatures in winter [62]. The reasons for the decrease in the chlorophyll content during the winter season may be due to the chloroplasts degradation, the oxidative degradation of chlorophyll by the peroxidase enzymes degrading chlorophyll [65], low irradiance [66] and changing the pigment concentration [67]. The chlorophyll content of each leaf area reflects the adaptation of the different plant species to the environmental conditions of the area [68].

Based on Student's t-test, statistically significant differences determined with respect to all concentrations of the mineral compositions (%) and water content (%) of *H. boveanus* in winter and summer seasons during Wadi El-Sheikh Awad, except Na^+ and N concentrations (Fig. 3). Significant differences trends among seasons in the minerals concentrations were in accordance with the results of Kamel and El-Absy [21], Glavac et al. [69] and Al-Qahtani et al. [70], but an opposite tendency was found by Williams and Chadwick [71]. During the two seasons, Ca^{2+} , Mg^{2+} , S, Cl^- and N concentrations and water content of *H. boveanus* plants increased in the winter season compared to the summer season. Whilst, the values of Na^+ , K^+ , P and Fe^{2+} concentrations in the summer season were higher than in the winter season of *H. boveanus* plants. These findings are in agreement with Kamel and El-Absy [21] and El-Lamey [50].

The water content % in plants is the most important physiological measure that affects the

efficiency of photosynthesis, plant growth and biomass productivity, in addition to being an indicator of plant tolerance to drought and salinity, because water stress restricts transpiration, including stomata closure and water evaporation from the leaf surface [35]. Some chemical compositions such as K^+ and Na^+ revealed higher variation under salinity stress while some other chemical compositions like nitrogen showed higher variation under drought stress [72]. When stress intensity increases, the minerals uptake by the plant may decrease [73] and it was also found that some minerals in plants are reduced due to other minerals [21]. To adapt to diverse environmental stresses, the plants have evolved complex physiological and biochemical adaptations [74], and mineral concentrations in the tissues of the plant species have been positively correlated with their habitats [75]. The concentrations of Na^+ and K^+ and ion balance play important roles in plant salt tolerance [76].

In Fig. 4, the results of statistical analysis by Student's t-test exhibited that there was a significant influence on proline and total carbohydrates contents in *H. boveanus* plant between the two studied seasons. On the contrary, no statistically significant differences in crude protein % were noticed between the winter and summer seasons (Fig. 4). Similarly, results were reported by Gonzalez-Hernandez et al. [77] for crude protein, by Pouris et al. [78] for proline and by Al-Qahtani et al. [70] for total carbohydrates.

The contents of proline and total carbohydrates in *H. boveanus* plant in the summer season were higher than in the winter season, but crude protein % was lower in the summer season than in the winter season. Other studies have also the highest contents for total carbohydrates [70] and proline [79] in the summer season and for the crude protein in the winter season [50].

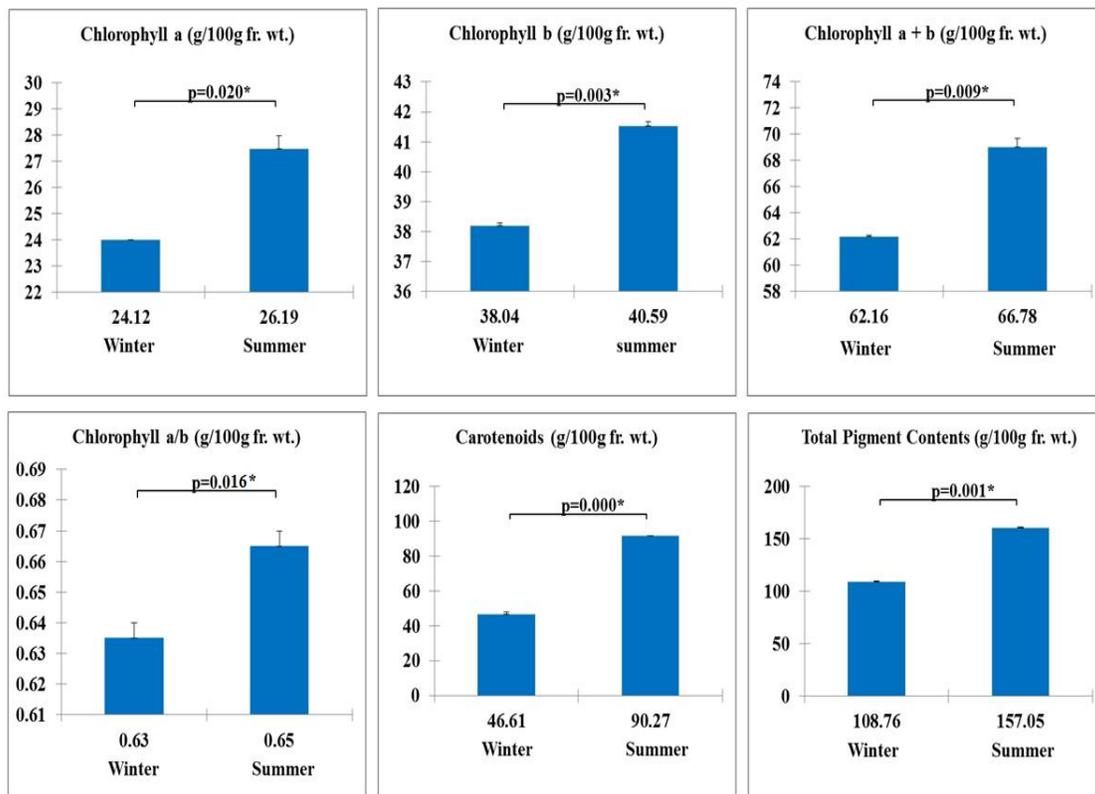


Fig. 2. Mean values of photosynthetic pigments contents in *H. boveanus* during different seasons. According to Student's t-test, the asterisks (*) denote a significant difference ($p < 0.05$) between the winter and summer seasons

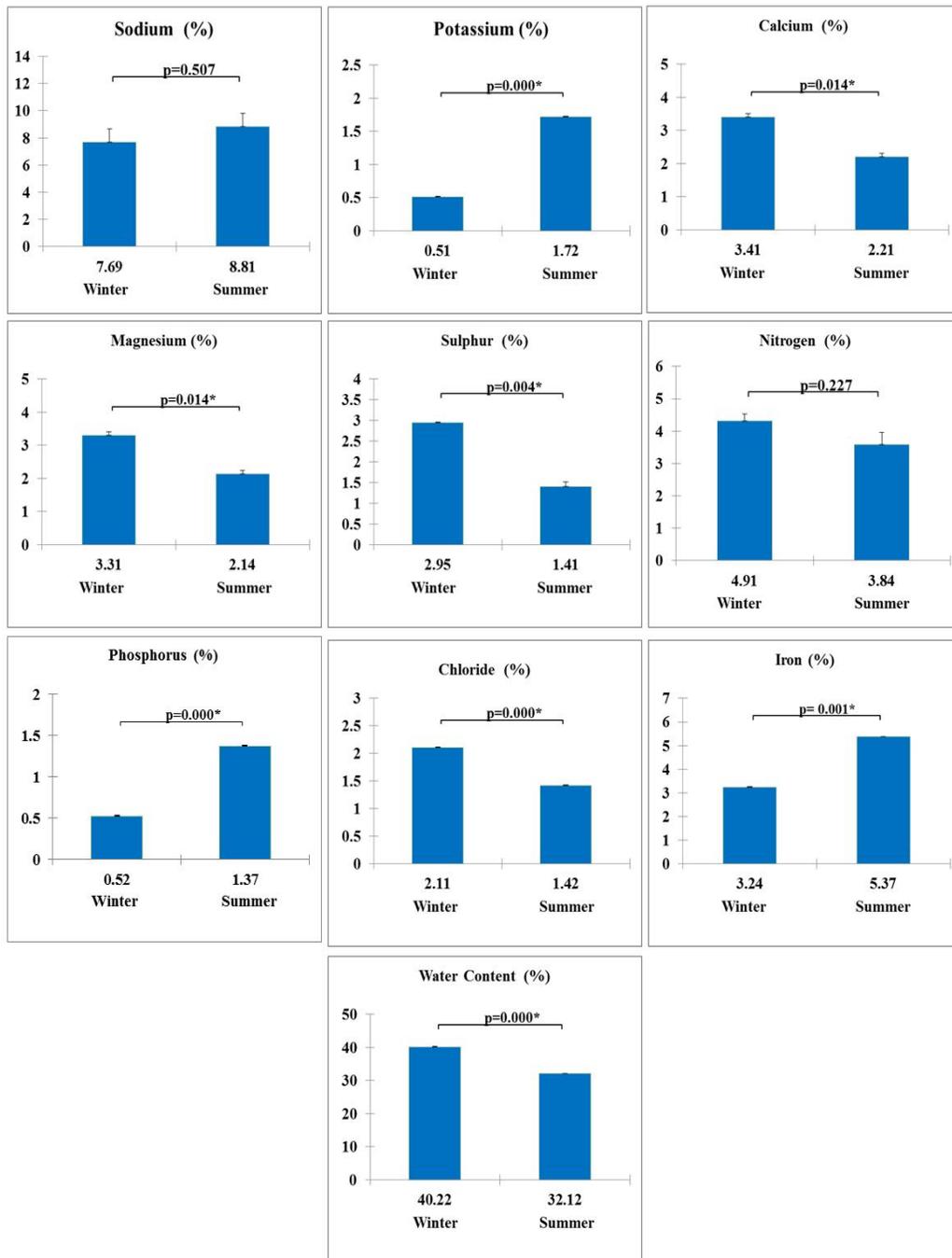


Fig. 3. Mean values of mineral composition and water content in *H. boveanus* during different seasons. According to Student's t-test, the asterisks (*) denote a significant difference ($p < 0.05$) between the winter and summer seasons

Increased proline content can be used as an indicator of disturbed physiological conditions as drought and salinity stresses in most plant species [79, 80]. Proline accumulation in plants

increased under drought and salinity conditions but was more under drought conditions [81, 82]. The increase in proline is due to the high total soluble salts in the soil [83].

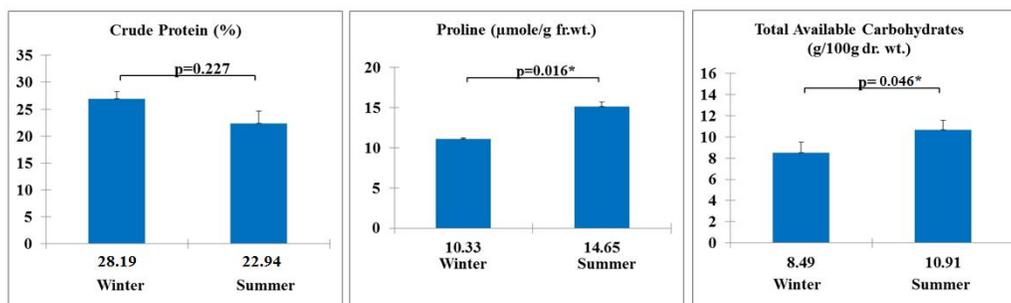


Fig. 4. Mean values of crude protein %, proline and total carbohydrates contents in *H. boveanus* during different seasons. According to Student's t-test, the asterisks (*) denote a significant difference ($p < 0.05$) between the winter and summer seasons

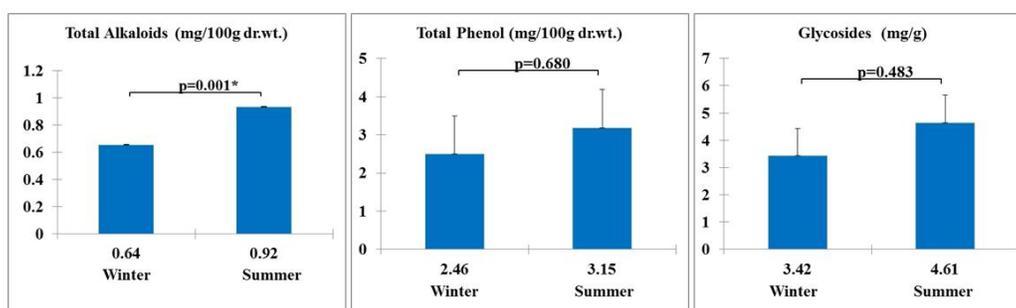


Fig. 5. Mean values of total alkaloids, glycosides and total phenol contents in *H. boveanus* during different seasons. According to Student's t-test, the asterisks (*) denote a significant difference ($p < 0.05$) between the winter and summer seasons

Differences between the winter and summer seasons were statistically significant for the total alkaloids, but not statistical significance for the glycosides content and total phenol contents (Fig. 5). The studies carried out by Soni et al. [82], Salminen et al. [84], Elaloui et al. [85], Lin et al. [86] and Aoussar et al. [87] showed that secondary metabolites in some plant species are influenced significantly by seasonal variations. When comparing the two seasons, the total alkaloids, glycosides and total phenol contents in *H. boveanus* plant were higher in the summer season than in the winter season. El-Shazly et al. [14] detected 23 alkaloids in *H. boveanus*. Several studies noticed the effect of seasonal variation on secondary metabolites in different plants. Significantly higher total alkaloids and total phenolic compounds were higher in winter than summer [88], while the total alkaloids increased during winter than other seasons [89]. The difference in the secondary metabolites is due to the effect of the conditions of soil growth in addition to the humidity and temperature [90]. Cold weather decreases biosynthesis and consequently decreases some secondary

metabolites compounds [91], while increasing with increasing light intensity [92]. On the basis of these results and the results of Soni et al. [82] and Deshmukh [93] secondary metabolites compounds increase with increasing water stress, especially the phenolic content through hydrolysis of glycosides.

3.3 Relationships between the Soil and Plant Variables

The PCA simplifies the complex data by transforming the number of correlated variables into a smaller number of variables in the data collection called principal components (PCs). Consequently, it has been used to understand the similarities and dissimilarities relationships between the soil and plant chemical variables during the winter and summer seasons of *H. boveanus*, which are graphically displayed in a biplot of PC1 and PC2 (Fig. 6). When comparing the winter and summer seasons, the PC1 and PC2 showed that the soil and plant chemical variables were distributed in different regions and formed different groups, and therefore these

results indicate that there are differences between these variables during the winter and summer seasons. According to this biplot, the *H. boveanus* performance during the winter and summer seasons displayed a positive correlation among most soil and plant chemical variables, but, they differed in their degree and consistency in quantity. In this study, within PC1 and PC2, the soil and plant chemical variables that recorded the highest factor loading were selected as the most important contributor to the two PCs. In previous studies, the first two PCs indicated to those relationships between the variables, which were interpreted as a related response to some of the soil variables by Islabão et al. [24] as well as to some soil and plant variables by Juhos et al. [25], Ferraz et al. [26] and Gil et al. [94]. PCA indicated that changes in the relationship between variables under different stresses are reasonable [72].

The first PC1 had positively and highly correlated with soil Mg^{2+} as in the Fig. 6, and positively correlated with soil Cl^- (Fig. 6B). As for plant chemical variables, the PC1 showed positive and highly association with total pigment, Chl a+b (Fig. 6A) and water content (Fig. B), and low positive association with carotenoids, Chl b (Fig. 6A) and crude protein (Fig. 6C). The PC2 had a high positive correlation with pH and Ca^{2+} (Fig. 6ABCD) and with EC (Fig. 6D), and moderate/low positive correlation with EC (Fig. 6ABC), Mg^{2+} (Fig. 6BCD) and K^+ (Fig. 6CD) in the soil of *H. boveanus*. Whilst, it had highly positively correlated with all photosynthetic pigments (Fig. 6A), with N, Cl^- (Fig. 6B), with crude protein (Fig. 6C) and with phenol (Fig. 6D), as well as low positive association with other variables in *H. boveanus*. On the other hand, the PC1 and PC2 under the winter and summer seasons were negatively correlated with the other soil and plant chemical variables. These results indicated that the PC1 had affected by soil magnesium and Chl b, Chl a+b, carotenoids and total pigment contents in plant, while, the PC2 had affected by most soil variables and all plant variables under study. The PC1 and PC2 in the Fig. 1 indicated that the chemical variables (Mg^{2+} , Cl^- , pH, EC, Ca^{2+} and K^+) of the soil associated with *H. boveanus* during the winter and summer seasons are variables with better soil chemical characteristics, which effect on the distribution of plants in the Wadi El-Sheikh Awad, South Sinai. This result was consistent with Omar [95] and Salama et al. [96]. The PC1 was highly positively correlated with EC and pH [25], with pH, K^+ , Ca^{2+} and Mg^{2+} [26] and with N, P

and K^+ [23], and highly negatively correlated with other soil variables. The PC2 showed a high positive correlation with soil pH [23, 25] and soil P [26], and displayed a negative correlation with other soil variables. Gil et al. [94] stated that the first two PCs has a correlation with some soil and plant variables studied. The PC1 correlation was low for crude protein content [97].

In soil variables, the high positive correlations were found among pH, EC and Ca^{2+} , among water content, Na^+ and K^+ as well as Na^+ , water content and Cl^- under the winter and summer seasons (Fig. 6ABCD). This result was consistent with Islabão et al. [24], Ferraz et al. [26], who reported that the most properties of soil were significantly positively or negatively associated with each other. Regarding plant variables, high positive correlations were observed among all photosynthetic pigments (Fig. 6A), among all mineral contents (Fig. 6B), among crude protein, proline and total carbohydrates (Fig. 6C) as well as among alkaloids, phenol and glycosides (Fig. 6D) at the summer and winter seasons. A highly positive correlation between Na^+ and K^+ , between proline and total protein, between proline and soluble carbohydrates, between chlorophyll a and carotenoids, between chlorophyll b and carotenoids as well as total chlorophyll and carotenoids were observed by Uvalle Saucedo et al. [62], Nejat and Sadeghi [72], Sims and Gamon [98], Murakeózy et al. [99], Kaspary et al. [100] and Sadeghi and Robati, [101]. Hendry and Price [102] confirm that the association between total chlorophyll and carotenoid concentrations plays an important role in protecting plants from stresses through photo-oxidation.

Soil magnesium had a highly positive correlation with plant variables i.e., crude protein (Fig 1C), WC (Fig 1B), Chl b, Chl a+b, carotenoids and total pigment contents (Fig. 6A). The pH, EC, and Ca^{2+} in soil were highly or moderately positively correlated with all photosynthetic pigments contents (Fig. 6A), with all the minerals contents (Fig. 6B), with crude protein, proline and carbohydrates (Fig. 6C) as well as with alkaloids, phenol and glycosides (Fig. 6D). A very low positive correlation was observed with crude protein, proline and carbohydrates (Fig. 6C) as well as with alkaloids, phenol and glycosides (Fig. 6D) at soil potassium. Non-correlations were noticed between other soil variables and studied plant variables. High correlation results between soil and plant variables indicate an effect of soil chemical variables on plant

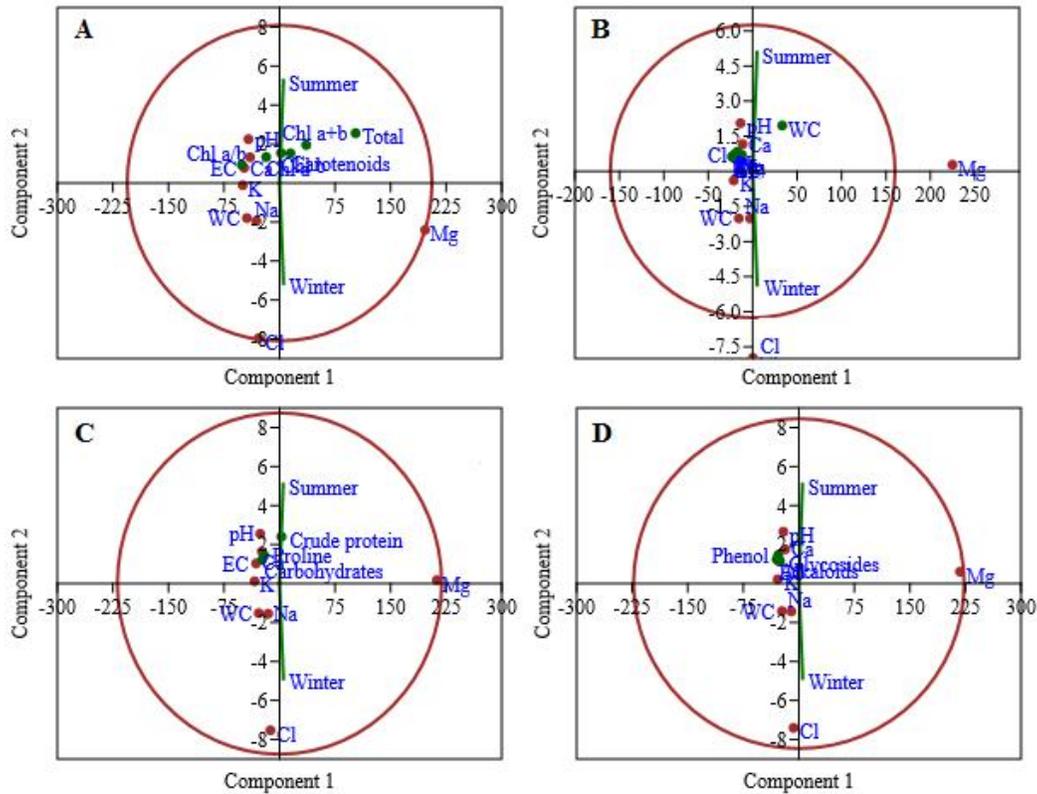


Fig. 6. Biplot diagram based on first two PCs axes of soil chemical variables (brown points) with (A) Photosynthetic pigments, (B) Mineral compositions and water content, (C) Crude protein %, proline and total carbohydrates, as well as with (D) total alkaloids, glycosides and total phenol contents in *H. boveanus* (green points) during different seasons. Symbols: EC: electrical conductivity; Cl-: chloride; Na+: sodium; K+: potassium; Ca²⁺: calcium; Mg²⁺: magnesium; S: sulfur; Fe²⁺: iron; P: phosphorus; N: total nitrogen; WC: water content; Total: total pigments

variables. Based on these correlations results, increasing these soil variables will increase plant variables. According to the PCA, the plant variables: Ca²⁺, Mg²⁺, proline, carbohydrate contents correlated significantly and positively with the most soil variables, but negatively with water content [94]. Carbohydrate contents show strong positive correlations with EC, but did not display any clear interactions with other soil variables studied [103]. While the plant species richness showed significant negative associations with EC, Na⁺, K⁺, Ca²⁺, Mg²⁺, Cl⁻ [56]. The relationship between tropane alkaloids and nitrogen availability in *H. niger* was confirmed by Nassar et al. [104]. Gil et al. [94] reported that increased calcium and magnesium accumulation within plant cells could contribute to physiological salinity tolerance mechanisms. The harmful effect of sodium ions could be counteracted by increasing calcium and

magnesium concentrations [105] and magnesium [106] to the extent that is non-toxic within cells, which gives plants a certain degree of tolerance. Gil et al. [94] cleared that the significant correlation between plant proline content and soil variables associated with environmental stress indicates the functional role of proline in the stress tolerance mechanisms of the plant species under study. Seasonal changes are the main factor effecting on the chemical composition of plants [107], as these reflect seasonal changes in physiological needs and efforts, rather than availability in plant content [108].

4. CONCLUSION

The results of the present study show that the soil and plant analysis of *H. boveanus* varied significantly during depths and seasons studied. Seasonal variation was observed for some plant

chemical compositions, which increased significantly during the summer season than the winter season. The principal component analysis formed different groups from soil and plant variables based on the variation in the two seasons studied. According to the biplot diagram, clear and reasonable correlations were found among soil and plant variables, which may lead to the maintenance of cellular osmotic balance to protect the plant during different stress conditions.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- Said R. The geology of Egypt. Elsevier Publishing Company. Amsterdam. 1990.
- Moustafa AA, Klopatek JM. Vegetation and landforms of the St. Katherine area, Southern Sinai, Egypt. Journal of Arid Environments. 1995;30:385-395.
- El-Absy KM, Kamel AM. Physiological and anatomical responses of *Teucrium polium* L. growing under different habitat conditions at North West Coast and South Sinai. Journal of Biodiversity and Environmental Sciences. 2019;15(6):40-52.
- Ayyad MA, Fakhry AM, Moustafa ARA. Plant biodiversity in the Saint Catherine area of the Sinai peninsula, Egypt. Biodiversity and Conservation. 2000;9: 265-281.
<https://doi.org/10.1023/A:1008973906522>
- Fayed A, Shaltout K. Conservation and sustainable use of medicinal plants in arid and semiarid eco-systems project, Egypt (GEF, UNDP) (project no: 12347/12348), Flora of Saint Katherine Protectorate, and Floristic Survey of the Mountainous Southern Sinai: Saint Katherine Protectorate, final report. 2004;13(3):13. Available:<http://www.idosi.org/aejaes/jaes>
- Satil F, Aslan M, Erdoğ an E, Polat R, Selvi S. Comparative anatomical studies on some species of *Hyoscyamus* L. (Solanaceae) growing in turkey. Bangladesh Journal Botany. 2015;44(1): 37-43.
DOI:<https://doi.org/10.3329/bjb.v44i1.22721>
- Hepper FN. Flora of Egypt. Family 159: Solanaceae. Täckholmia Additional Series. 1998;6:1-168.
- Boulos L. Flora of Egypt: Checklist Revised annotated edition. Al -Hadara Publishing, Cairo, Egypt, 2009;410.
- Rechinger KH. Flora des Iranischen hochlandes und der umrahmenden Gebirge (Persien, Afghanistan, Teile Von West-Pakistan, Nord-Iraq, Azerbaidjan, Turkmenistan), Graz-Austria. 1963;570-573.
- Yousaf Z, Masood S, Shinwari ZK, Khan MA, Rabani A. Evaluation of taxonomic status of medicinal species of the genus *Hyoscyamus*, *Withania*, *Atropa* and *Datura* based on polyacrylamide gelelectrophoresis. Pakistan Journal of Botany. 2008;40(6): 2289-2297.
- Boulos L. Flora of Egypt; Al Hadara Publishing Cairo: Cairo, Egypt, Volume 4. 2005.
- Mosallam HAM. Assessment of target species in St. Catherine protected area, Sinai, Egypt. Journal of Applied Sciences Research. 2007;3(6):456-469.
- Khafagi O, Hatab EE, Omar K. Challenges towards *Hypericum sinaicum* conservation in south Sinai, Egypt. Jordan Journal of Biological Sciences. 2012;6(2):116-126. DOI:10.12816/0000269
- El-Shazly A, Tei A, Witte L, El-Domiaty M, and Wink M. Tropane Alkaloids of *Hyoscyamus boveanus*, *H. desertorum*, *H. muticus* and *H. albus* from Egypt. Zeitschrift für Naturforschung C. 1997;52:729-739.
<https://doi.org/10.1515/znc-1997-11-1203>
- Bourebaba L, Saci S, Touguit D, Gali L, Terkmane S, Oukil N, Bedjou F. Evaluation of antidiabetic effect of total calystegines extracted from *Hyoscyamus albus*. Biomed Pharmacother. 2016;82:337-44. DOI:10.1016/j.biopha.2016.05.011
- Mateus L, Cherkaoui S, Christen P, Veuthey JL. Capillary electrophoresis for the analysis of tropane alkaloids: pharmaceutical and phytochemical applications. Journal Pharm Biomed Anal. 1998;18(4-5):815-25. DOI:10.1016/s0731-7085(98)00223-4
- Al-Snafi AE. Therapeutic importance of *Hyoscyamus* species grown in Iraq (*Hyoscyamus albus*, *Hyoscyamus niger* and *Hyoscyamus reticulatus*)-A review. IOSR Journal Pharmacy. 2018;8:18-32.

18. Reddy RK, Reddy SJ. Elemental concentrations in medicinally important leafy materials, *Chemosphere*. 1997;34(9-10): 2193-2212. DOI:[https://doi.org/10.1016/S0045-6535\(97\)00078-7](https://doi.org/10.1016/S0045-6535(97)00078-7)
19. Kumar A, Kaul MK, Bhan MK, Punit K, Khanna PK, Suri KA. Morphological and chemical variation in 25 collections of the Indian medicinal plant, *Withania somnifera* (L.) Dunal (Solanaceae). *Genetic Resources and Crop Evolution*. 2007;54:655-660. DOI:<https://doi.org/10.1007/s10722-006-9129-x>
20. Stengele M, Stahl-Biskup E. Seasonal variation of the essential oil of european pennyroyal (*Mentha pulegium* L.). *Acta Hort.* 1993;344:41-51. <https://doi.org/10.17660/ActaHortic.1993.344.5>
21. Kamel AM, El-Absy KM. Seasonal variations in protein patterns and mineral contents of *Lycium showii* under different habitat conditions. *Asian Plant Research Journal*. 2020;6(4): 91-103. DOI:<https://doi.org/10.9734/aprj/2020/v6i430141>
22. Abd-Elmabod SK, Bakr N, Muñoz-Rojas M, Pereira P, Zhang Z, Cerdà A, Jordán A, Mansour H, De La Rosa D, Jones L. Assessment of soil suitability for improvement of soil factors and agricultural management. *Sustainability*. 2019;11(6): 1588. <https://doi.org/10.3390/su11061588>
23. Metwally MS, Shaddad SM, Liu M, Yao R-J, Abdo AI, Li P, Jiao J, Chen X. Soil properties spatial variability and delineation of site-specific management zones based on soil fertility using fuzzy clustering in a hilly field in Jianyang, Sichuan, China. *Sustainability*. 2019;11(24):7084. <https://doi.org/10.3390/su11247084>
24. Islabão GO, Pinto MAB, Selau, LPR, Vahl LC, Timm LC. Characterization of soil chemical properties of strawberry fields using principal component analysis. *R. Bras. Ci. Solo*. 2013;37:168-176. DOI:<https://doi.org/10.1590/S0100-06832013000100017>
25. Juhos K, Szabó S, Ladányi M. Influence of soil properties on crop yield: a multivariate statistical approach. *International Agrophysics*. 2015;29:433-440. DOI:10.1515/intag-2015-0049
26. Ferraz GAS, PFerra PFP, Martins FB, Silva FM, Damasceno FA, Barbari M. Principal components in the study of soil and plant properties in precision coffee farming. *Agronomy Research*. 2019;17(2):418-429. DOI:<https://doi.org/10.15159/AR.19.114>
27. Badr A, El-Shazly H, Ahmed H, Hamouda M, El-Khateeb E, Sakr M. Genetic diversity of *Achillea fragrantissima* in Egypt inferred from phenotypic variations and ISSR markers associated with traits of plant size and seed yield. *Plant Genetic Resources*. 2017;15(3):239-247. doi:10.1017/S1479262115000568
28. Meakin K, de Kort SR, Gilbert H, Gilbert F, Zalat S, Mohi L, Ibrahim S, Griffin J. Monitoring birds, reptiles and butterflies in the St Katherine Protectorate, Egypt. *Egyptian Journal of Biology*. 2005;7:66-95.
29. Täckholm V. *Students Flora of Egypt*. Cairo University, Cooperative printing Co., Beirut, 2nd. Ed., 1974;482-483.
30. Hosni HA, El-Ghamery AA, Sadek AM. Distinction between *Hyoscyamus boveanus* (Dunal) Asch & Schweinf. and *Hyoscyamus muticus* L. (Family: Solanaceae). *Egypt. Journal Biotechnology*. 2017;55: 94-113.
31. Jackson ML. *Soil chemical analysis*. Prtice Hall of India Private., New Delhi., India; 1967.
32. Rowell DL. *Soil science methods and applications*. Longman Publishers, Singapors. 1994;229.
33. Jackson ML. *Soil chemical analysis constable and co. Ltd. London*; 1962.
34. Tuzuner A. *Soil and water laboratory analysis guide*. Ankara: General Directorate of Rural Services Publications. 1990.
35. Jin X, Shi C, Yu CY, Yamada T, Sacks EJ. Determination of leaf water content by visible and near-infrared spectrometry and multivariate calibration in *Miscanthus*. *Front. Plant Science*. 2017;8:721. DOI:10.3389/fpls.2017.00721
36. Chapman H. Cation-exchange capacity. In *Methods of Soil Analysis: Part 2 Chemical and Microbiological Properties*; JohnWiley & Sons: Hoboken, NJ, USA, 1965;9:891–901.
37. Jackson W, Thomas GW. Effect of KCl and Dolomitic Limestone on growth and on uptake of sweat potato. *Soil Science*. 1960;89:347-352.
38. Bremner JM. Total nitrogen and inorganic forms of nitrogen. In: *Methods of Soil Analyses*. (Ed.): C.A. Black. American

- Society of Agronomy, Madison, Wisconsin. 1956;1149-1237.
39. Lichtenthaler HK. Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. In: Packer, L.; Douce, R., ed. Methods in enzymology. London: Academic Press. 1987;148:350-382. DOI:[https://doi.org/10.1016/0076-6879\(87\)48036-1](https://doi.org/10.1016/0076-6879(87)48036-1)
 40. Harborne JB. Phytochemical Methods: A guide to modern techniques of plant analysis. Chapman and Hall Ltd., London. 1973.
 41. Hikino H, Kiso Y, Wagner H, Fiebig M. Antihepatotoxic actions of flavonolignans from *Silybum marianum* fruits. *Planta Medica*. 1984;50(3):248-50. DOI:10.1055/s-2007-969690.
 42. Singleton VL, Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *The American Journal of Enology and Viticulture*. 1965;16:144-158.
 43. Allen SE. Chemical analysis of ecological materials. Blackwell Scientific Publications. Oxford, London Edinburgh. 1989;Pp. 368.
 44. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline for water-stress studies. *Plant and Soil*. 1972;39:205-207.
 45. Chaplin MF, Kennedy JF. Carbohydrate Analysis" A practical approach. 2nd Ed. Oxford Univ., Press Oxford, New York, Tokyo, 324 pp. 1994.
 46. McDonald JH. Handbook of Biological Statistics Sparky House Publishing, Baltimore. 2008.
 47. El-Absy KM, Kamel AM. Physiological and anatomical responses of *Teucrium polium* L. growing under different habitat conditions at North West Coast and South Sinai. *Journal of Biodiversity and Environmental Sciences*. 2019;15(6):40-52.
 48. Moustafa AM, Zayed A. Effect of environmental factors on the flora of alluvial fans in southern Sinai, *Journal of Arid Environments*. 1996;32(4):431-443. DOI:<https://doi.org/10.1006/jare.1996.0036>
 49. Bonifacio E, Morte A. Soil properties in Desert truffles- Phylogeny, Physiology, Distribution and Domestication. Editor: Summerer Verlag Germany. Heidelberg Germany. 2014;57-67.
 50. El-Lamey TM. Changes in some chemical compounds of *Retama raetam* (Forssk.) Webb & Berthel. in response to different environmental conditions. *Journal of Biodiversity and Environmental Sciences*. 2020;16(2):78-91.
 51. Misra A, Tyler G. Influence of soil moisture on soil solution chemistry and concentrations of minerals in the Calcicoles *Phleum phleoides* and *Veronica spicata* grown on a limestone soil. *Annals of Botany*. 1999;84:401-410. DOI:<https://doi.org/10.1006/anbo.1999.0941>
 52. Ahmed FA, Abdallah NM, Ezz MK, El-Azab MM. Seasonal variations and identification of biologically active constituents of *Lycium shawii* Plant Roem. & Shult. (Family Solanaceae). *International Journal of Innovative Science, Engineering & Technology*. 2017;4(11):34-47.
 53. Larcher W. Physiological plant ecology. Summerer-Verlage, Berlin Heidelberg, Germany. 1995.
 54. El-Absy KM, Kasim WA, El-Kady HF, El-Shourbagy MN. Physiological studies on *Achillea fragrantissima* and *Artemisia judaica* in Saint Katherine, South Sinai, Egypt. *International Journal of Scientific Research in Agriculture Science* 2015;2(Proceedings) 127-136.
 55. Labib CM, El-Shatoury SA, Dewedar A. Isolation, characterization and bioactivities of soil actinomycetes from the world heritage site (WHS) of Saint Katherine. Egypt. *Journal Botany*. 2016;56(3,46):769-783.
 56. Salama FM, Abd El-Ghani MM, El-Naggar SM, Aljarroushi MM. Vegetation analysis and species diversity in the desert ecosystem of coastal wadis of South Sinai, Egypt. *Journal of Biology and Earth Sciences*. 2013;(3)2:B214-B227.
 57. Lopes AS, Guilherme LRG. Interpretação de análise de solo - Conceitos e aplicações. 3.ed. São Paulo, Associação Nacional para Difusão de Adubos - ANDA, 2004;50. (Boletim Técnico, 2).
 58. Zhao C, Long J, Liao H, Zheng C, Li J, Liu L, Zhang M. Dynamics of soil microbial communities following vegetation succession in a karst mountain ecosystem, Southwest China. *Sci Rep*. 2019;9: 2160. DOI:<https://doi.org/10.1038/s41598-018-36886-z>
 59. Hoskins BR. Soil testing handbook for professionals in agriculture, horticulture, nutrient and residuals management. 3rd ed. England: Maine Forestry & Agricultural

- Experiment Station, University of Maine. p.119. 1997.
60. Laekemariam F, Kibret K, Shiferaw H. Potassium (K)-to-magnesium (Mg) ratio, its spatial variability and implications to potential Mg-induced K deficiency in Nitisols of Southern Ethiopia. *Agriculture & Food Security*. 2018;7:13. DOI:<https://doi.org/10.1186/s40066-018-0165-5>
 61. Grattan SR, Grieve CM. Salinity±mineral nutrient relations in horticultural crops. *Scientia Horticulturae*. 1999;78:127-157. DOI:[https://doi.org/10.1016/S0304-4238\(98\)00192-7](https://doi.org/10.1016/S0304-4238(98)00192-7)
 62. Uvalle Saucedo JI, Gonzalez Rodriguez H, Ramirez Lozano RG, Silva IC, Gomez Meza MV. Seasonal trends of chlorophylls a and b and carotenoids in native trees and shrubs of Northeastern Mexico. *Journal of Biological Sciences*. 2008;8: 258-267. DOI:10.3923/jbs.2008.258.267
 63. Devi K, Kapila S, Rao A. Seasonal variations in photosynthetic pigments of three species of Marchantiaceae. *International Journal of Advances in Pharmacy, Biology and Chemistry*. 2015;4(3): 713-718.
 64. Banik S, Mukherjee R, Ghosh P, Karmaka S, Chatterjee S. Estimation of plant pigments concentration from tulsi (*Ocimum sanctum* Linn.): a six months study. *Journal of Pharmacognosy and Phytochemistry*. 2018;7(4): 2681-2684.
 65. Shimokawa K, Uchida Y. A chlorophyllide a degrading enzyme (hydrogen peroxide-2, 4 dichlorophenol requiring) of *Citris unshiu* fruits. *Journal Jap. Soc. Hortic. Science*. 1992;61(1):175-181.
 66. Kramer PJ, Kozlowski T. *Physiology of wood plants*. Academic Press, New York 1-811. 1979.
 67. Devmalkar VS, Murumkar CV, Salunkhe SM, Chavan SJ. Studies on pigment chlorophyll isolation and estimation of different bryophytes for their biochemical properties. *Journal Nat. Prod. Plant Res*. 2014;4(2): 56-61.
 68. Ivanov LA, Ronzhina DA, Yudina PK, Zolotareva NV, I. V. Kalashnikova IV, Ivanova LA. Seasonal dynamics of the chlorophyll and carotenoid content in the leaves of steppe and forest plants on species and community level. *Russ Journal Plant Physiology*. 2020;67:453-462. <https://doi.org/10.1134/S1021443720030115>
 69. Glavac V, Koenies H, Ebben U. Seasonal variation of calcium, magnesium, potassium, and manganese contents in xylem sap of beech (*Fagus sylvatica* L.) in a 35-year-old limestone beech forest stand. *Trees*. 1990;4:75-80. <https://doi.org/10.1007/BF00226069>
 70. Al-Qahtani H, Alfarhan AH, Al-Othman ZM. Changes in chemical composition of *Zilla spinosa* Forssk. medicinal plants grown in Saudi Arabia in response to spatial and seasonal variations. *Saudi Journal of Biological Sciences*. 2020;27(10):2756-2769, DOI:<https://doi.org/10.1016/j.sjbs.2020.06.035>
 71. Williams P, Chadwick M. Seasonal variation in the availability of plant nutrients in acid colliery spoil. *Journal of Applied Ecology*. 1977;14(3):919-931. doi:10.2307/2402822
 72. Nejat N, Sadeghi H. Finding out relationships among some morpho-biochemical parameters of christ's thorn (*Ziziphus spina-christi*) under drought and salinity stresses. *Planta Daninha, Viçosa-MG*. 2016;34(4):667-674. DOI:10.1590/S0100-83582016340400006
 73. Akinci S, Lösel DM. Plant water stress response mechanisms. Ismail, M.; Rahman, M. and Hasegawa, H. (Eds). *Rijeka Croatia*. 2012;1:15-42. DOI:10.5772/29578
 74. Osakabe Y, Osakabe K, Shinozaki K and Tran L-SP. Response of plants to water stress. *Frontiers Plant Science*. 2014;5:86. DOI:10.3389/fpls.2014.00086
 75. Morsy AA, Youssef AM, Mosallam HAM, Hashem AM. Assessment of selected species along Al-Alamein-Alexandria international desert road, Egypt. *Journal of Applied Sciences Research*. 2008;4(10):1276-1284.
 76. Zheng H, Zhao H, Liu H, Wang J, Zou D. QTL analysis of Na⁺ and K⁺ concentrations in shoots and roots under NaCl stress based on linkage and association analysis in japonica rice. *Euphytica*. 2014;201:109-121. DOI:<https://doi.org/10.1007/s10681-014-1192-3>
 77. Gonzalez-Hernandez, MP, Starkey EE, Karchesy J. Seasonal variation in concentrations of fiber, crude protein, and phenolic compounds in leaves of red alder

- (*Alnus rubra*): nutritional implications for cervids. *Journal of Chemical Ecology*. 2000;26(1):293-301.
DOI:10.1023/A:1005462100010
78. Pouris J, Meletiou-Christou M-S, Chimona C, Rhizopoulou S. Seasonal functional partitioning of carbohydrates and proline among plant parts of the sand daffodil. *Agronomy*. 2020;10(4):539. <https://doi.org/10.3390/agronomy10040539>
 79. Dhaka V, Meena KL. Seasonal variation in free proline content in some species of family euphorbiaceae of the rajasthan, India. *Journal of Experimental Biology and Agricultural Sciences*. 2018;6(1):249-252.
DOI:10.18006/2018.6(1).249.252
 80. Lansac AR, Zaballos JP, Martin A. Seasonal water potential changes and proline accumulation in mediterranean shrubland species. *Vegetatio*. 1994;113:141-154.
DOI:<https://doi.org/10.1007/BF00044231>
 81. Anosheh HP, Sadeghi H, Emam Y. Chemical priming with urea and KNO₃ enhances maize hybrids (*Zea mays* L.) seed viability under abiotic stress. *Journal Crop Science Biotechnology*. 2011;14:289295. <https://doi.org/10.1007/s12892-011-0039-x>
 82. Soni U, Brar S, Gauttam VK. Effect of seasonal variation on secondary metabolites of medicinal plants. *International Journal of Pharmaceutical Sciences and Research*. 2015;6(9): 3654-3662. DOI:10.13040/IJPSR.0975-8232.6(9).3654-62.
 83. Ahmed AM. Ecophysiological studies on certain wild plants grown in different habitats in the Egyptian desert. Ph. D. thesis. Ain Shams Univ., Cairo, Egypt. 2002.
 84. Salminen JP, Roslin T, Karonen M, Sinkkonen J, Pihlaja K, Pulkkinen P. Seasonal variation in the content of hydrolyzable tannins, flavonoid glycosides, and proanthocyanidins in oak leaves. *Journal of Chemical Ecology*. 2004;30:1693-1711.
DOI:<https://doi.org/10.1023/B:JOEC.0000042396.40756.b7>
 85. Elaloui M, Ghazghazi H, Ennajah A, Manaa S, Guezmir W, Karray NB, Laamouri A. Phenolic profile, antioxidant capacity of five *Ziziphus spina-christi* (L.) Willd provenances and their allelopathic effects on *Trigonella foenum-graecum* L. and *Lens culinaris* L. seeds. *Nat Prod Research*. 2017;31(10):1209-1213.
DOI: 10.1080/14786419.2016.1226830
 86. Lin W, Kuang Y, Wang J, Duan D, Xu W, Tian P, Nzabanita C, Wang M, Li M, Ma B. Effects of seasonal variation on the alkaloids of different ecotypes of *Epichloë endophyte-festuca sinensis* associations. *Frontiers in Microbiology*. 2019;10:1695. <https://doi.org/10.3389/fmicb.2019.01695>
 87. Aoussar N, Rhallabi N, Mhand RA, Manzali R, Bouksaim M, Douira A, Mellouki F. Seasonal variation of antioxidant activity and phenolic content of *Pseudevernia furfuracea*, *Evernia prunastri* and *Ramalina farinacea* from Morocco, *Journal of the Saudi Society of Agricultural Sciences*. 2020;19(1):1-6.
DOI:<https://doi.org/10.1016/j.jssas.2018.03.004>
 88. Ncube B, Nair JJ, Rárová L, Strnad M, Finnie JF, Van Staden J. Seasonal pharmacological properties and alkaloid content in *Cyrtanthus contractus* N.E. Br., South African *Journal of Botany*. 2015;97:69-76, <https://doi.org/10.1016/j.sajb.2014.12.005>
 89. Li JY, Wang XB, Luo JG, Kong LY. Seasonal variation of alkaloid contents and anti-inflammatory activity of *Rhizoma coptidis* based on fingerprints combined with chemometrics methods, *Journal of Chromatographic Science*. 2015;53(7):1131-1139.
DOI:<https://doi.org/10.1093/chromsci/bmu175>
 90. Brito SMO, Coutinho HDM, Talvani A, Coronel C, Barbosa AGR, Vega C, Figueredo FG, Tintino SR, Lima LF, Boligon AA, Athayde ML, Menezes IRA. Analysis of bioactivities and chemical composition of *Ziziphus joazeiro* Mart. using HPLC–DAD. *Food Chemistry*. 2015;186:185-191.
DOI:<https://doi.org/10.1016/j.foodchem.2014.10.031>
 91. Chao MT, Mohamed AR. Roles of titanium dioxide and ion-doped titanium dioxide on photocatalytic degradation of organic pollutants (phenolic compounds and dyes) in aqueous solutions: A review. *Journal of Alloys and Compounds*. 2011;509(5): 1648-1660. <https://doi.org/10.1016/j.jallcom.2010.10.181>
 92. Slimestad R, Verheul B. Review of flavonoids and other phenolics from fruits

- of different tomato (*Lycopersicon esculentum* Mill) Cultivars. Journal Science Food Agriculture. 2009;89:1255-70. DOI:<https://doi.org/10.1002/jsfa.3605>
93. Deshmukh RN. Osmolyte accumulation in sorghum bicolor under water stress. Bionano frontier. 2012;5(2):204-09.
 94. Gil R, Bautista I, Boscaiu M, Lidón A, Wankhade S, Sánchez H, Llinares J, Vicente O. Responses of five Mediterranean halophytes to seasonal changes in environmental conditions. AoB PLANTS. 2014;6:plu049. DOI:<https://doi.org/10.1093/aobpla/plu049>
 95. Omar KA (2014) Ecological and climatic attribute analysis for Egyptian *Hypericum sinaicum*. American Journal of Life Sciences. 2014;2(6):369-381. DOI:10.11648/j.ajls.20140206.17
 96. Salama F, El-Ghani MA, Gadallah M, El-Naggar S, Amro A. Variations in Vegetation Structure, Species Dominance and Plant Communities in South of the Eastern Desert-Egypt. Notulae Scientia Biologicae. 2014;6(1):41-58. DOI:<https://doi.org/10.15835/nsb619191>
 97. Bhattarai K, Douglas A, Johnson, Thomas A, Jones, Kevin J, Connors, Dale R, Gardner. Physiological and Morphological Characterization of Basalt Milkvetch (*Astragalus filipes*): Basis for Plant Improvement. Rangeland Ecology & Management. 2008;61(4):444-455. DOI:<https://doi.org/10.2111/08-011.1>
 98. Sims DA, Gamon JA. Relationships between leaf pigment content and spectral reflectance across a wide range of species, leaf structures and developmental stages. Remote Sens. Environment. 2002;81:337-354. DOI:[https://doi.org/10.1016/S0034-4257\(02\)00010-X](https://doi.org/10.1016/S0034-4257(02)00010-X)
 99. Murakeözy EP, Nagy Z, Duhazé C, Bouchereau A, Tuba Z. Seasonal changes in the levels of compatible osmolytes in three halophytic species of inland saline vegetation in Hungary, Journal of Plant Physiology. 2003;160(4): 395-401. DOI:<https://doi.org/10.1078/0176-1617-00790>
 100. Kaspary TE, Lamego FP, Cutti L, Aguiar ACM, Bellé C. Determination of photosynthetic pigments in fleabane biotypes susceptible and resistant to the herbicide glyphosate. Planta Daninha, Viçosa-MG. 2014;32(2):417-426. DOI: <http://dx.doi.org/10.1590/S0100-83582014000200020>
 101. Sadeghi H, Robati Z. Response of *Cichorium intybus* L. to eight seed priming methods under osmotic stress conditions. Biocatalysis and Agricultural Biotechnology. 2015;4(4): 443-448. DOI:<https://doi.org/10.1016/j.bcab.2015.08.003>
 102. Hendry GAF, Price AH. Stress Indicators: Chlorophylls and Carotenoids. In: Hendry, G.A.F. and Grime, J.P., Eds., Methods in Comparative Plant Ecology, Chapman Hall, London. 1993;148-152. DOI:<http://dx.doi.org/10.1007/978-94-011-1494-3>
 103. Gil R, Lull C, Boscaiu M, Bautista I, Lidón A, Vicente O. Soluble carbohydrates as osmolytes in several halophytes from a mediterranean salt marsh. Notulae Botanicae Horti Agrobotanici Cluj-Napoca. 2011;39(2), 09-17. DOI:<https://doi.org/10.15835/nbha3927176>
 104. Nassar RMA, Seleem EA, Caruso G, Sekara A, Abdelhamid MT. The nitrogen-fixing bacteria-effective enhancers of growth and chemical composition of Egyptian henbane under varied mineral N nutrition. Agronomy. 2020;10(7):921. DOI:<https://doi.org/10.3390/agronomy10070921>
 105. Gul B, Khan MA (2008). Role of Calcium in Alleviating Salinity Effects in Coastal Halophytes. In: Khan M.A., Weber D.J. (eds) Ecophysiology of High Salinity Tolerant Plants. Tasks for Vegetation Science. 2008;vol 40. Summerer, Dordrecht. https://doi.org/10.1007/1-4020-4018-0_6
 106. Grigore MN, Boscaiu M, Llinares J, Vicente O. Mitigation of salt stress-induced inhibition of *Plantago crassifolia* reproductive development by supplemental calcium or magnesium. Notulae Botanicae Horti Agrobotanici Cluj-Napoca. 2012;40(2):58-66. DOI:<https://doi.org/10.15835/nbha4028246>
 107. Sgarbossa J, Schmidt D, Schwerz F, Schwerz L, Prochnow D, Caron BO. Effect of season and irrigation on the chemical composition of *Aloysia triphylla* essential oil. Revista Ceres. 2019;66(2): 85-93. DOI:<https://dx.doi.org/10.1590/0034-737x201966020002>
 108. Estevez JA, Landete-Castillejos T, GarcíaB AJ, Ceacero F, Martínez A,

Gaspar-López E, Calatayud A, Gallego L.
Seasonal variations in plant mineral
content and free-choice minerals

consumed by deer. *Animal Production
Science*. 2010;50(3):177-185.
DOI: <https://doi.org/10.1071/AN09012>

© 2021 El-Absy; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/75256>