

Application of Phosphorus and Calcium to Improve Physiological Response of Groundnut (*Arachis hypogaea* L.)

Henry Tamba Nyuma^{1,2*}, William S. Kollie¹,
Cornel. L. Rweyemamu³ and John S. Fayiah⁴

¹Department of Agronomy, University of Liberia, College of Agriculture and Forestry, Monrovia, Liberia.

²Department of Soil Science, University of Eldoret, Kenya.

³Department of Crop Science and Horticulture, Sokoine University of Agriculture, Morogoro, Tanzania.

⁴National Public Health Institute of Liberia, Monrovia, Liberia.

Authors' contributions

This work was carried out in collaboration among all authors. Authors HTN and CLR designed the study, performed the statistical analysis. Author HTN wrote the protocol and wrote the first draft of the manuscript. Authors CLR and JSF managed the analyses of the study. Author WSK managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/CJAST/2021/v40i931353

Editor(s):

(1) Dr. João Miguel Dias, University of Aveiro, Portugal.

Reviewers:

(1) Riaz Ahmad, Bahauddin Zakariya University, Pakistan.

(2) Tális Pereira Matias, Universidade Federal de Alfenas, Brazil.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/68100>

Review Article

Received 06 March 2021

Accepted 12 May 2021

Published 15 May 2021

ABSTRACT

Genotype, environment and agronomic practices are key determinants of crop growth and productivity which are important to provide food, feed, raw materials and income to small holder farmers in Africa. The study was conducted at Crop museum, Sokoine University of Agriculture, Morogoro, Tanzania to investigate physiological response of groundnut to calcium and phosphorus nutrition among three improved groundnut genotypes *Mangaka* (G_1), *Masasi* (G_2) and *Pendo* (G_3) with three levels of phosphorus and at calcium control (T_0), 125 kg/ha (T_1) and 55 kg/ha (T_2) supplied from Diammonium Phosphate (DAP) and *Minjingu mazao*, respectively. Significant ($P \leq 0.05$) influence of calcium was observed on leaf area index (LAI) 4.03; crop biomass 88.79 g/plant; number of nodules 66.22, and crop growth rate (CGR) 15.05 g m²/day. Whereas phosphorus had

*Corresponding author: E-mail: tnyuma@gmail.com, nyumahht@ul.edu.lr;

significant influence on net assimilation rate (NAR) 10.84 g m²/day. Similarly, groundnut genotype (*Masasi*) had significant influence on LAI (3.95); CGR (13.04 g m²/day); NAR (12.36 g m²/day) and number of nodes (64.93). However, there was no significant effect of genotype on crop biomass recorded from the investigation. Growth parameters were significantly influenced by genotype and fertilizer interactions with Significant (P=0.2) interaction effect for crop biomass observed between G₃ x T₁, while G₃ x T₂ significantly affected number of nodules and G₂ x T₁ had Significant interaction effects on CGR and NAR. This is an indication that investment in improved genotype and appropriate application rate of fertilizers has the potential to enhance yield and income of smallholder farmers.

Keywords: Calcium; genotypes; groundnut; phosphorus; physiology.

1. INTRODUCTION

Groundnut (*Arachis hypogaea* L.) is an important global multi-purpose legume crop that reinforces livelihood strategies meeting food, nutrition, and income security [1]. It is an essential source of protein, calories, essential fatty acids, vitamins, and minerals associated with several health benefits for human, and raw materials for industrial use including products such as food, feed, fuel, paints, lubricants and insecticides and can be used as an ideal crop in rotational systems to improve soil fertility due to its natural ability to fix atmospheric nitrogen [1-6].

However, yield levels of the crop has remained relatively low (964 kg/ha) in Sub Saharan Africa (SSA) far less than potential yields of up to 3500 kg/ha reported elsewhere due to a range of biotic, abiotic and socio-economic constraints including lack of access to quality seeds, poor agronomic practices such as low use of fertilizers and limited access to extension and advisory services [4,7-11].

In Tanzania, groundnut serves as food, feed and raw material for industries, but farmers have limited access to improved genotypes and fertilizer resources due to cost and lack of education on new agronomic practices, thus major limiting production, especially where there are no recommended rates of fertilizers for groundnut whose production is characterized by low external input, limited extension and advisory services, erratic rainfall, hence fluctuating yields despite the economic importance of the crop [6,10,12].

Lack of access to improved groundnut genotype in Tanzania poses numerous challenges to farmers ranging from poor crop performance, crop quality and subsequently low income at household level. Selection of improved

genotypes with superior characteristic under good agronomic practices has been highlighted as key to sustainable production Daudi et al. [10].

Sustainable groundnut production depends on proper selection of variety, fertilizer management and other management practices as optimal rates of fertilizer have positive effect on the performance of groundnut [13, 6]. Applications of micro and macronutrients are effective for increase of crop production. These had capability to mitigate negative effects of environmental stresses [14, 15]. Numerous crop production was improved by balance fertilization [16].

The objective of the study was to evaluate the effect of fertilizer on growth attributes of groundnut and provide information on optimal fertilizer rates for groundnut genotypes with superior performance, hence improved production.

2. MATERIALS AND METHODS

2.1 Experimental Site

Field experiment was laid out at Sokoine University of Agriculture (SUA), Crop Museum situated at latitude 6° 45' South and longitude 37° 40' East at 525 m.a.s.l in Morogoro municipality. The area is situated at the foot of Uluguru Mountain in Morogoro and has a bimodal rainfall pattern, predominantly sub-humid with irregular and unreliable distribution characterized by kaolinitic clay soils, which are well drained and mostly clay [17,18].

2.2 Experimental Design

Three groundnut varieties (*Mangaka*, *Masasi* and *Pendo*) used in the experiment were obtained from Naliedende Agricultural Research Institute

(ARI), in Mtwara, Tanzania whereas fertilizer materials included *Minjingu Mazao*, source of calcium contained N (10%), 20% P₂O₅, 25 % CaO, 5% S, 0.5% Zn, 1.5% MgO and 0.1% Boron [19] and Diammonium Phosphate (DAP), source of phosphorus (NH₄)₂ (HPO₄) was composed of 18% N and 46%P₂O₅.

The experiment was laid out in a split plot Randomized Complete Block Design (RCBD) with four replications and three groundnut genotypes, G₁ = (Mangaka), G₂ = (Masasi), G = (Pendo) as main plot factor (factor A) whereas fertilizer types were applied as sub –plot factor (factor B). Fertilizer types were applied as follows: T₀ = (Control), T₁ = (55 kg P/ha) T₂ = (125 kg Ca/ha).

2.3 Soil Analyses

Soil samples were collected as recommended by Landon [20] at the depth of 0–30cm for physicochemical analyses. Soil pH was determined electrometrically in 1:2.5 soil–water suspensions as described by Thomas [21]. Available P was analyzed using Bray –1 [22]. Organic carbon determination was done by wet digestion method of Walkley and Black [23]. Total N was determined by the micro – Kjeldahl digestion–distillation method [24]. Exchangeable bases, K and Na were analyzed by flame photometer whereas Ca and Mg were determined by Atomic Absorption Spectrophotometer (AAS) as described by Petersen [25]. Available Cu, Zn, Fe and Mn were extracted by DTPA as described by Lindsay et al. [26].

Soil physicochemical properties were done by the Department of Soil and Geological Sciences at Sokoine University of Agriculture. Soil physicochemical properties are presented in Table 1. Agronomic and other management practices were carried out as described by Kanyeka et al. [27].

2.4 Data Collection

Data on growth characteristics of groundnut was done at 4,6,8 and 10 Weeks After Planting (WAP).

Crop biomass (g/plant): Crop Biomass (BM) was determined from three plants by harvesting a whole plant including roots from the penultimate rows of the plot. The harvested

plants were oven dried at 70 °C for 48 hours and the dry weights were recorded using (Doran 7000, Doran Inc.) electronic weighing scale

Leaf area index: Leaf area index (L.A.I) was calculated as described by Brown [28].

$$\text{LAI} = \text{leaf area per plant (cm}^2\text{)} / \text{Ground cover per plant (cm}^2\text{)}$$

Crop growth rate (g.m²/day): crop growth rate (CGR) at different growth stages was determined as recommended by Brown (1984) as follow:

$$\text{CGR} = \text{W}_2 - \text{W}_1 / \text{T}_2 - \text{T}_1 \times 1/\text{GA (g m}^{-2} \text{ day}^{-1}\text{)}$$

W₁ = Weight at T₁ of the period

W₂ = Weight at T₂ of the Period

T₁ = Time in date at the start of the period

T₂ = Time in the date at the end of the period

GA = Ground area

Net assimilation rate (g m² /day): NAR was determined by assessing the ratio of CGR and LAI at various growth stages. Net assimilation rate was calculated as described by Brown [28].

$$\text{NAR} = (1/\text{LAI}) (\text{dw}/\text{dt}) = \text{g m}^2/\text{day}$$

Where;

NAR = net

assimilation

rate dw/dt =

the change in

plant dry

matter per unit

time

LAI = leaf area index

Number of nodules per plant: number of nodules was determined by counting the number of nodes visible on the roots of three plants per plot and the average number of nodules was recorded.

2.5 Data Analysis

Data collected were subjected to analysis of variance (ANOVA) using GENSTAT released version 14th edition and declared significant at P < 0.05 using the following statistical model described by Gomez and Gomez [29]. Mean separation test was done using Duncan Multiple Range Test (DMRT) at P ≤ 0.05.

3. RESULTS AND DISCUSSION

3.1 Soil Characteristics

Soil analyses in the current study revealed that soil textural (sandy clay loam); bulk density ($1.2\text{g}/\text{cm}^3$) and soil pH (5.9) were considered appropriate for root penetration, moisture retention, nodule formation, hence increased growth, development and production [30-32]. Nkot et al. [33] reported poor nodule formation in soil with pH between 3.5 and 5.0 had adverse effect on root penetration, nodule formation, nutrient availability.

A very low status of available P (0.048 mg/kg), as observed in the current study was evident that P was a major limiting factor for groundnut production in the study area. Phosphorus deficiency constitutes a serious limitation to crop production in weathered tropical soils containing high Fe and Al oxides that quickly fix added P. Moazed et al. [34] reported that low quantities of soluble P in Oxisols limit crop production and productivity.

Soil calcium level in the current study was rated very high according to Landon [20], but was inadequate to meet crop growth requirement. High level of soil Ca is affected by amounts of organic matter resulting in the inability of the soil to hold the amount required for crop growth. Calcium availability is crucial for pod formation Calcium application also influences shelling percent, seed and pod yields in groundnut [35]. Results also revealed that macronutrient deficiencies, especially P could be a major constraint to groundnut production because initial soil P was very low for optimum yield [36].

Crop Biomass: results from the current study showed no significant influence of genotype on crop biomass however; fertilizer had significant ($P=0.001$) influence on biomass. The highest biomass (88.79 g /plant) was observed with application of *Minjingu mazao*. Also, an exponential increase in biomass was observed from week 8 to 10 due to the influence of fertilizer Fig. 1.

Leaf Area Index (LAI): Genotypic variation on LAI was recorded during the current study with Masasi having significant influence on LAI. Significant influence of calcium was observed on LAI compared to other treatments. Similar findings were reported by Habib [37], Song et al. [38].

Crop growth rate: groundnut genotype had significant influence on CGR as the lowest CGR was observed in Mangaka ($11.18\text{ g m}^2/\text{day}$) whereas the highest was observed in Masasi ($12.88\text{ g m}^2/\text{day}$). Significant influence of fertilizer type on CGR was observed and recorded in the order of *Minjingu mazao* > DAP > control. Such findings are in conformity with Mahpara et al. [16] who reported increased ground growth with application of macronutrients.

Net assimilation rate: Net assimilation rate was significantly influenced by genotype as Masasi recorded the highest NAR ($10.56\text{ g m}^2/\text{day}$), whereas P significantly improved NAR from 7.79 to $10.84\text{ g m}^2/\text{day}$. Nagaraj et al. [39] attributed increased NAR in groundnut with increase in the amount of P compared to control plots.

Number of nodules: number of nodules was significantly ($P=0.001$) affected by application of P. The current study recorded a 29.0 % increase in number of nodules with the application of P

Table 1. Soil physio-chemical characteristics at experimental site

A. Physical (%)	Chemical Macro Nutrients / Rating ¹	Exchangeable Cations $\text{cmol}_c^{(+)}$	Micronutrients (mg/kg)
Sand 49.2	pH 5.9*	Calcium 27.3 VH	Iron 31.7 VH
Clay 42.72	Organic Carbon 0.07 - VL	Magnesium 186.6 VH	Manganese 92.0 VH
Silt 8.08	Total Nitrogen 0.18 -VL	Potassium 2.16 H	Cooper 13.0 VH
Textural Class: Sandy clay	Organic Matter 0.12-VL	Sodium 5.4 VH	Zinc 24.6 VH
	C : N ratio 1:1.25-VL		
	Extractable P 0.048 - L		

¹Soil nutrient status rating as described by Landon, 1991. L= low, H= high, VL= very low, VH= very high

compared to control plots, similar observations were reported by Mouri et al. [13], Dzomeku et al. [40] with the application of P. Similarly, number of nodules significantly ($P=0.001$) differed among genotypes with Masasi recording

the highest number of nodules (64.93) compared to the other genotypes. An observed increase in the number of nodules was recorded for all genotypes between weeks 6 and 8 with steady decline at week 10 (Fig. 2).

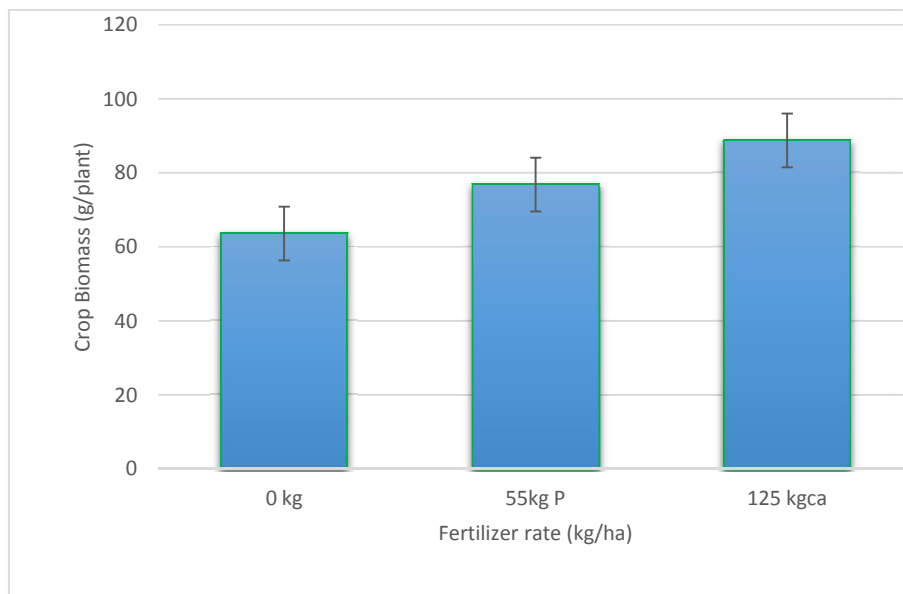


Fig. 1. Response of crop biomass to fertilizer treatment

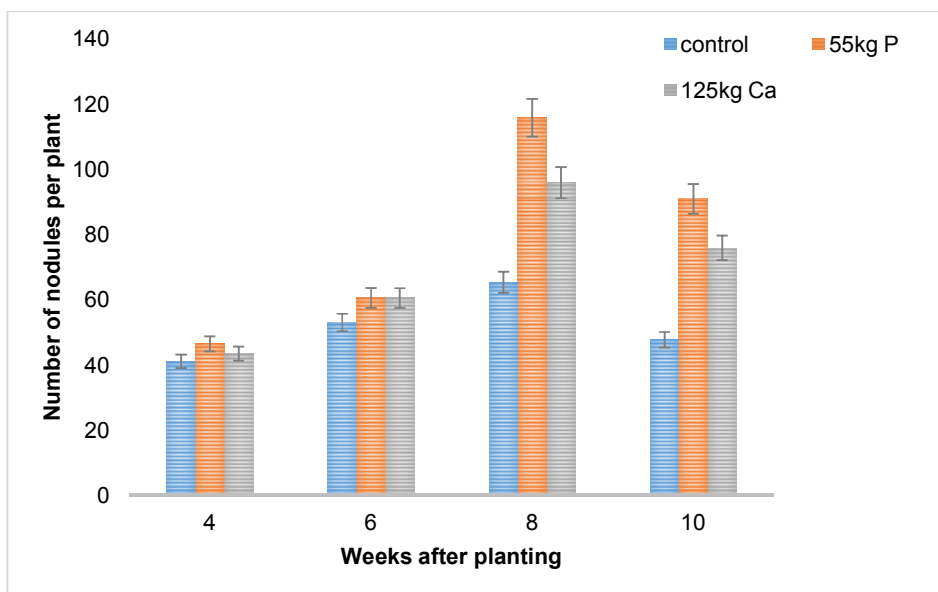


Fig. 2. Effect of fertilizer on number of nodules in groundnut at various growth stages

Table 2. Effect of genotype on growth characteristics of groundnut

Genotype	BM (g/plant)	LAI	CGR (g m ² /day)	NAR (g m ² /day)	Number of Nodules
G ₁	75.00a	3.71b	11.18a	8.53a	42.14a
G ₂	79.25a	3.95b	13.04c	12.36c	64.93b
G ₃	75.01a	2.86a	12.88b	10.56b	63.25b
SE ±	3.33	0.14	1.67	0.16	3.24
CV P value	4.4 0.01	4.60 0.01	5.3 0.03	3.1 0.001	5.700.001

*Means in the same column and factor followed by the same letter are not significantly different at $P \leq 0.05$ according to Duncan Multiple Range Test. Note: BM = plant biomass (g/plant), LAI = Leaf area index; CGR: Crop growth rate (g m²/d,) and NAR: Net assimilation rate (g m²/d)

Table 3. Effect of fertilizer on growth variables of groundnut

Treatment	BM (g/plant)	LAI	CGR (g m ² /day)	NAR (g m ² /day)	Number of Nodules
T ₀	63.62a	3.51a	8.48a	7.79a	47.01a
T ₁	76.85b	3.52a	13.55b	10.84b	57.08b
T ₂	88.79c	4.03b	15.05c	10.81b	66.22c
SE ±	3.17	0.02	3.35	0.076	2.76
CV	4.1	1.8	4.9	3.0	4.9
P value	0.001	0.001	0.001	0.076	0.001

*Means in the same column and factor followed by the same letter are not significantly different at $P \leq 0.05$ according to Duncan Multiple Range Test. Note: BM = plant biomass (g/plant), LAI = Leaf area index; CGR: Crop growth rate (g m²/d,) and NAR: Net assimilation rate (g m²/d)

Table 4. Genotype × Fertilizer interaction effect on growth characteristics of groundnut

Interaction (G x T)	BM (g/plant)	LAI	CGR (g m ² /day)	NAR (g m ² /day)	Number Nodules /plant
G ₁ x T ₀	62.24a *	3.10 c	12.00a	11.65a	27.74a
G ₁ x T ₁	62.51a	3.15 c	13.77a	11.87a	54.79c
G ₁ x T ₂	66.12ab	3.26 c	13.75a	11.74a	63.70d
G ₂ x T ₀	73.63bc	1.95a	22.72c	16.30b	34.99b
G ₂ x T ₁	82.01cd	1.99a	26.83c	16.35b	54.54c
G ₂ x T ₂	74.92bc	1.88a	24.96c	17.32c	51.50c
G ₃ x T ₀	85.26de	2.31b	13.76b	11.41a	52.77c
G ₃ x T ₁	93.24e	2.31b	21.86c	11.57a	85.46e
G ₃ x T ₂	87.87de	2.36b	16.36b	12.52a	85.48e
Mean	76.42	2.47	69.0	15.64	56.77
SE±	6.13	6.06	6.13	0.30	4.46
Cv(ab) %	7.8	13.3	8.9	2.3	7.9
P- value	0.2	0.09	0.04	0.003	0.001

Interaction effects: Significant (P=0.2) interaction effect between G₃ x T₁ was observed on plant biomass; significant (P=0.09) interaction effect for G₂ x T₂ on LAI; Significant (P= 0.04) and (P= 0.003) interaction effect for CGR and NAR was observed with G₂ x T₁ and G₂ x T₂, respectively. Number of nodules was significantly (P=0.001) influenced by genotype x fertilizer interaction with G₃ x T₂ interaction showing positive response.

4. CONCLUSION

Results from the current study showed both genotype and fertilizer had significant (P≤ 0.05) influence on growth characteristics of groundnut with positive influence calcium observed on leaf area index (LAI) 4.03; crop biomass 88.79 g/plant; number of nodules 66.22, and crop growth rate (CGR) 15.05 g m²/day. Whereas phosphorus had significant influence on Net assimilation rate (NAR) 10.84 g m²/day.

Similarly, groundnut genotype (*Masasi*) had significant influence on LAI (3.95); CGR (13.04 g m²/day); NAR (12.36 g m²/day) and number of nodes (64.93). Given the significant interaction effect of genotype x fertilizer on growth parameters, genotype Pendo proved superior performance under both treatments were significantly influenced by genotype x fertilizer interactions with Significant (P=0.2) interaction effect for crop biomass observed between G₃ x T₁, while G₃ x T₂ significantly affected number of nodules and G₂ x T₁ had Significant interaction effects on CGR and NAR. This is an indication that under appropriate agronomic management with 55 kg P/ha and 125 kg Ca/ha, Pendo has the potential to superior performance, hence yield compared to other genotypes used in the experiment.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Ojiewo CO, Janila P, Bhatnagar-Mathur P, Pandey MK, Desmae H, Okori P, et al. Advances in Crop Improvement and Delivery Research for Nutritional Quality and Health Benefits of Groundnut (*Arachis hypogaea* L.) Frontiers in Plant Science; 2020. DOI: 10.3389/fpls.2020.00029. Available:www.frontiersin.org
- Guasch-Ferré M, Liu X, Malik VS, Sun Q, Willett WC, Manson JE, et al. Nut consumption and risk of cardiovascular disease. J. Am. Coll. Cardiol. 2017;70(20):2519–2532. DOI: 10.1016/j.jacc.2017.09.035.
- Jaiswal JK, Levini LA, Dakora FD. Phylogenetically diverse group of native bacterial symbionts isolated from root nodules of groundnut (*Arachis hypogaea* L.) in South Africa. Syst Appl Microbiol. 2017;40:215–226.
- Variath MT, Janila P. Economic and Academic Importance of peanut. In: Varshney, R., Pandey, M., Puppala, N. Editors. The peanut genome. Compendium of plant genomes. Cham: Springer; 2017; Doi: 10.1007/978-3-319-63935-2-2.
- Willett W, Rockstrom J, Loken B, Springmann M, Lang T, Vermeulen S, et al. Food in the Anthropocene: the EAT-Lancet Commission on healthy diets from sustainable food systems. Lancet. 2019;393(10170):447–492. DOI: 10.1016/S0140-6736(18)31788-4.
- Nyuma HT, Rweyemamu CL, Fayiah JS. Effect of Fertilizer and Genotype on Crop Quality and Profitability of Groundnut in Morogoro, Tanzania, International Journal of Advanced Research and Publications (IJARP). 2019;3(11):1-5. Available:http://www.ijarp.org/online-paperspublishing/nov2019.html
- Debele S, Amare A. Integrated management of Cercospora leaf spots of groundnut (*Arachis hypogaea* L.) through host resistance and fungicides in Eastern Ethiopia. Afr J Plant Sci. 2015;9:82–89. DOI:10.5897/AJPS2014.1260.
- Desmae H, Sones K. Groundnut cropping guide. Africa soil health Consortium. Nairobi: CAB International. FAOSTAT; 2017. [accessed 2018 Feb 16] Available:http:www.fao.org/faostat/en/#data/QC
- Mastewal A, Sakhuja PK, Mashilla D. Evaluation of released and local groundnut varieties against groundnut rust (*Puccinia arachidis*) at Babile, Eastern Ethiopia. Open Acc J Agric Res. 2017;2:000-123.
- Daudi H, Shimelis H, Laing M, Okori P, Mponda O. Groundnut production Constraints, Farming Systems, and Farmer-Preferred Traits in Tanzania, Journal of Crop Improvement. 2018;32(6):812-828. DOI: 10.1080/15427528.2018.1531801.
- Abady S, Shimelis H, Janila P, Mashilo J. Groundnut (*Arachis hypogaea* L.) improvement in sub-Saharan Africa: A review, Acta Agriculturae Scandinavia, Section B — Soil & Plant Science; 2019. DOI: 10.1080/09064710.2019.1601252.
- Kamhabwa F. Consumption of fertilizers and fertilizer use by crop in Tanzania. Dar es Salaam, Tanzania. 2014;23.
- Mouri SJ, Sarkar MAR, Uddin MR, Sarker UK, Kaysar MS, Hoque MMI. Effect of variety and phosphorus on the yield components and yield of groundnut. Progressive Agriculture. 2018;29(2):117-126.
- Ahmad R, Hussain S, Anjum MA, Khalid MF, Saqib M, Zakir I, Hassan A, Fahad S, Ahmad S. Oxidative stress and antioxidant defense mechanisms in plants under salt stress. Plant Abiotic Stress Tolerance. 2019;191-205.

15. Shahzad S, Ali S, Ahmad R, Ercisli S, Anjum MA. Foliar Application of Silicon Enhances Growth, Flower Yield, Quality and Postharvest Life of Tuberose (*Polianthes tuberosa* L.) under Saline Conditions by Improving Antioxidant Defense Mechanism. *Silicon*. 2021;1-8.
16. Mahpara S, Shahnawaz M, Rehman K, Ahmad R, Khan FU. 4. Nitrogen fertilization induced drought tolerance in sunflower: a review. *Pure and Applied Biology (PAB)*. 2019;8(2):1675-1683.
17. Mahoo HF, Young MDB, Mziray OB. Rainfall variability and its implication for the transferability of experimental results in semi-arid areas of Tanzania. *Journal of Agricultural Sciences*. 1999;2(2):127–140.
18. Semoka JRM. Research report for progress rock phosphate project. Sokoine University of Agriculture, Morogoro, Tanzania. 2003;24.
19. Minjingu Mines and fertilizer LTD. Developing Indigenous Resources for Fertilizer Production in Africa: Minjingu Mines and Fertilizer Ltd., Mtwara, Tanzania. 2014;20.
20. Landon JR. Booker Tropical Soil Manual. A hand book for soil survey and agricultural land evaluation in the tropics and sub tropics. Longman Publishers, New York. 1991;474.
21. Thomas GW. Exchangeable cations. In: *Method of Soil Analysis*. (Edited by Page AL, Miller RH, Keeney DR), American Society of Agronomy, Madison, Wisconsin. 1996;154–169.
22. Olsen RS, Sommers LE. Phosphorus. In: *Methods of Soil Analysis*. (Edited by Page AL, Miller RM, Keeney DR), American Society of Agronomy, Madison, Wisconsin. 1982;422–430.
23. Nelson DW, Sommers LE. Total carbon and organic matter. In: *Methods of Soil Analysis, Chemical and Microbiology Properties*. (Edited by Page AL, Miller RM, Keeney DR), American Society of Agronomy, Madison, Wisconsin. 1982;539–577.
24. Bremner JM, Mulvaney CS. Total nitrogen. In: *Methods of Soil Analysis Part 2. Chemical and Microbiological Properties*. (Edited by Page, A. L., Miller. R. H. and Keeney, D. R.), American Society of Agronomy and soil Science of America, Madison, Wisconsin. 1982;593–624.
25. Petersen L. *Soil Analytical Methods. Soil Testing Management and Development*. Soil. Resources Development Institute, Dhaka, Bangladesh. 1996;12.
26. Lindsay WL. Development of a DPTA soil test for zinc, iron, manganese and copper. *Journal of American Soil Science Society*. 1978;42:421–428.
27. Kanyeka E, Kamala R, Kasuga R. *Improved Agricultural Technologies in Tanzania*. Ministry of Agriculture Food Security and Cooperatives, Dar es Salaam, Tanzania. 2007;125.
28. Brown RH. *Growth of the green plant, Physiological Basis of Crop Growth and Development*. ASA, CSSA, Madison, Wisconsin, USA. 1984;153–173.
29. Gomez AK, Gomez AA. *Statistical Procedure for Agricultural Research*. John Wiley and Sons, Inc., London. 1984;680.
30. Kollie WS. *Effects of Liming on Nitrogen Fixation and Yields of Soybean in an Acid Soil of Morogoro, Tanzania* Dissertation for Award of MSc Degree in Crop Science at Sokoine University of Agriculture Morogoro, Tanzania. 2016;93.
31. Bakari R, Mungai N, Thuita M, Masso C. Impact of soil acidity and liming on soybean (*Glycine max*) nodulation and nitrogen fixation in Kenyan soils *Acta Agriculturae Scandinavica, Section B — Soil & Plant Science*. 2020;70(8). Available:<https://doi.org/10.1080/09064710.2020.1833976>
32. Anjum MA, Muhammad HMD, Balal RM, Ahmad R. Performance of two onion (*Allium cepa* L.) cultivars under two different planting systems in calcareous soil. *Journal of Horticultural Science and Technology*. 2019;2:54-59.
33. Nkot L, Nwaga D, Ngakou A, Fankem H, Etoa F. Variation in nodulation and growth of groundnut (*Arachis hypogaea* L.) on oxisols from land use systems of the humid forest zone in southern Cameroon. *African Journal of Biotechnology*. 2011;10(20):3996–4004.
34. Moazed H, Hoseini Y, Naseri AA, Abbasi F. Determining phosphorus adsorption isotherm in soil and its relation to soil characteristics. *International Journal of Soil Science*. 2010;5:131–139.
35. Yang S, Wang J, Tang Z, et al. Transcriptome of peanut kernel and shell reveals the mechanism of calcium on peanut pod development. *Sci Rep*. 2020;10:15723. Available:<https://doi.org/10.1038/s41598-020-72893-9>

36. Katsaruware RD, Mabwe C. Response of groundnut (*Arachis hypogea*) to inorganic fertilizer use in smallholder farming of Makonde District, Zimbabwe. *Journal of Biology, Agriculture and Healthcare*. 2014;4:7.
37. Habib A. Calcium Management towards peg and pod development of groundnut thesis submitted in partial fulfillment of the requirements for the degree of Master of Science (MS) in Agronomy at Sher-e-Bangla Agricultural University, Bangladesh. 2014;160.
38. Song Q, Liu Y, Pang J, Yong JWH, Chen Y, Bai C, Gille C, Shi Q, Wu D, Han X, Li T, Siddique KHM, Lambers H. Supplementary Calcium Restores Peanut (*Arachis hypogaea*) Growth and Photosynthetic Capacity Under Low Nocturnal Temperature. *Front. Plant Sci*. 2020;10:1637. DOI: 10.3389/fpls.2019.01637
39. Nagaraj R, Hanumanthappa M, Kamath S. Growth Parameters and Yield of Groundnut as Influenced by Integrated Nutrient Management at Coastal Zone of Karnataka. *Journal of Pharmacognosy and Phytochemistry*. 2018;7(5):2725-2729.
40. Dzomeku IK, Baba SM, Abdulai Mohammed AM, Abdulai AI. Groundnut Response to Phosphorus and Weed Management in Guinea Savanna Zone of Ghana. *Tropicultura [online]*. 2019;37(1). Available: <https://popups.uliege.be/443/2295-8010/index.php?id=253>. DOI: 10.25518/2295-8010.253.

© 2021 Nyuma et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/68100>