

International Journal of Environment and Climate Change

Volume 13, Issue 11, Page 2609-2621, 2023; Article no.IJECC.109202 ISSN: 2581-8627 (Past name: British Journal of Environment & Climate Change, Past ISSN: 2231–4784)

Studies on Genetic Diversity and Stress Indices for Salinity Tolerance in *Triticum dicoccum*

Srivijay S. Malipatil ^{a*}, Suma S. Biradar ^{b*}, S. A. Desai ^b, **S. S. Gundlur ^c, Rohit kumar ^d, Karthik Narayanaswamy ^a, T. Harish Vikram ^a and Vishwasgowda C. ^a**

^aDepartment of Genetics & Plant Breeding, Keladi Shivapppa Nayaka University of Agricultural and Horticultural Science, Navile, Shivamogga-577204, India. ^bDepartment of Genetics & Plant Breeding, University of Agricultural Sciences, Dharwad-580005, India.

c Department of Soil Science & Agricultural Chemistry, University of Agricultural Sciences, Dharwad-580005, India.

^dDepartment of Genetics & Plant Breeding, University of Agricultural Sciences, Raichur-584104, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJECC/2023/v13i113429

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/109202

> *Received: 03/09/2023 Accepted: 07/11/2023 Published: 09/11/2023*

Original Research Article

ABSTRACT

Salinity stands out as a crucial abiotic stress factor that significantly impacts both crop yield and quality. Wheat, considered a moderately salt-tolerant crop, offers a wealth of variability and diversity within its species, presenting an accessible avenue for enhancing wheat's salt tolerance. Consequently, this study investigated the extent of genetic diversity for salt tolerance among sixty *Triticum dicoccum* germplasm accessions. In both saline and control condition, the genotypes were

Int. J. Environ. Clim. Change, vol. 13, no. 11, pp. 2609-2621, 2023

^{}Corresponding author: E-mail: srivijay@uahs.edu.in, biradarss@uasd.in, srivijay7560@gmail.com;*

divided into three and nine separate clusters respectively, showing that there was a significant level of genetic variability among the genotypes by multivariate analysis. It was observed that biomass was the largest contributor (85.19 %) to the divergence in the saline situation. Among the yield parameters under saline condition, tillers per meter followed by thousand-grain weight and spike length were observed to be important. Thirteen genotypes were salt-tolerant, according to the Stress susceptibility index (SSI). The stress tolerance index (STI) was defined as a useful tool for determining the high yield and potential stress tolerance of genotypes. Based on the stress tolerance index, only eight genotypes were said to be tolerant.

Keywords: Dicoccum wheat; saline; diversity; clusters and stress index.

1. INTRODUCTION

The nutrient-rich cereal wheat is farmed in a wide range of conditions; it covers over 217 million hectares globally, ranking first among all crops in terms of acreage, and produces around 731 million tonnes per year [1]. Wheat (*Triticum aestivum* L.) is one of the world's most frequently farmed cereal crops and one of the world's most important staple foods for about 2.5 billion people. Cereals are essential in satisfying the expanding global food demand, especially in underdeveloped nations where cereal-based farming is the only source of nutrients and calories [2,3]. Wheat is the most significant food crop in North Africa, West Asia, and Central Asia, accounting for more than half of the calories consumed. In terms of calories and dietary consumption, wheat, along with rice, is one of the most important sources of protein in the leastdeveloped and middle-income nations.

Triticum dicoccum, commonly known as emmer wheat, holds significant importance due to its historical, nutritional, and genetic contributions. Emmer wheat is one of the oldest cultivated grains, dating back thousands of years, and played a crucial role in the development of early human civilizations. Its nutritional value, including dietary fiber, protein, and essential nutrients, makes it a valuable dietary component [4]. Moreover, emmer wheat's genetic diversity is a vital resource for wheat breeding programs, enhancing the resilience of modern wheat varieties. Its adaptability and resistance to pests and diseases promote sustainable agriculture. Emmer wheat remains a source of dietary variety and cultural heritage in regions where it is cultivated, contributing to local food security and traditions. Furthermore, the grain's lower gluten content makes it suitable for individuals with mild gluten sensitivities, though it's not recommended for those with celiac disease Lupo L. [5].

Biological and abiotic stress, often known as environmental stresses, have a significant impact on wheat grain yield [6,7]. Salinity is one of the most important abiotic stresses affecting crop yield and quality [8], affecting around 7 percent of the world's total land area [9]. Salt affects a higher percentage of cultivated land, with 23 percent of cultivated land being saline and 20 percent of irrigated land suffering from secondary salinization. Furthermore, there is a perilous tendency of a 10 percent annual rise in the saline area all over the world [10]. In India, around 6.73 million hectares of land are affected by salt, with 3.77 million hectares affected by sodicity and 2.96 million hectares affected by salinity, respectively [11].

Wheat morphophysiological and agronomic traits play a key role in assessing the value of each trait in enhancing yield, therefore these qualities were employed in breeding projects that resulted in higher yields and the introduction of commercial varieties that can survive salt stress. The selection of genotypes with all conceivable yield contributing traits is the most critical requirement in any crop improvement program. The main factor to consider when making a selection is genotype variability for grain yield and yield component characteristics. Further, such information may be great to set the future path for the salt tolerance breeding program in wheat.

2. MATERIALS AND METHODS

The present study included pre-tested 60 dicoccum wheat germplasm accessions under both saline (natural saline soils) and control plot, out of which, 46 local germplasm accessions, 4 advanced breeding lines, and ten checks (Table 1) were evaluated in alpha lattice design with four blocks and two replications. Each block consisted of 15 genotypes with two rows per genotype and 3- meter length with a spacing of 20 cm between rows. The present investigation was carried out during *rabi* 2020-21 at Ugar Sugars Pvt. Ltd, Ugar Khurd, Tq- Chikkodi, DtBelagavi, Karnataka, which is situated in the northern transitional tract of Karnataka with 16° 38' N latitude and 74° 49' E longitude at an altitude of 537 m above mean sea level (AMSL). Two independent experiments were carried out in both salt stress and controlled condition. Under saline condition, pH of less than 8 and EC of more than 4 dS/m and under control condition pH of 6-8 and EC of less than 4 dS/m was maintained. Table 2 represents the pH and EC dS/m (1 soil:2.5 water) of top layer (0-20 cm) and bottom layer (20-40 cm) of the control and saline field.

Morphological traits like germination percent, days to fifty percent flowering, days to maturity and plant height, physiological traits (SPAD and NDVI) at booting, anthesis and grain filling stages, yield and yield attributes *viz.,* number of productive tillers per meter row, spike length (cm), number of grains per spike, number of spikelets per spike, harvest index, biomass and thousand-grain weight (g) were measured under the study. The statistical analysis for genetic divergence was done using Mahalanobis-D*²*

statistics [12] and the clustering of genotypes was done using the Tocher method [13]. Character contribution towards genetic divergence was computed using the method given by Singh and Chaudhary [14]. Stress indices *viz.,* Stress Susceptibility Index [15] and Stress Tolerance Index [16] were calculated to classify genotypes into salt tolerant and salt susceptible. The genotypes were grouped in three different tolerance categories based on stress susceptibility index as shown under [17]. The genotypes were grouped in three different tolerance categories based on stress susceptibility index as shown under [18]. Following are the formulas to calculate the indices, where Ys is the yield under saline condition and Yp is the control condition, - indicated their respective means.

$$
SSI = \frac{1 - (Ys/Yp)}{1 - (Ys/Yp)}
$$

$$
STI = \frac{(Ys \times Yp)}{(Yp^{2})}
$$

3. RESULTS AND DISCUSSION

Salinity affected all the traits under study and grain yield has reduced significantly. Traits like spikelets per spike, days to maturity, and days to fifty percent flowering are least affected by salt stress (Table 3). All the traits showed a significant difference for all the traits under saline and control condition. Salt-tolerant wheat varieties are considered to be a viable and effective strategy for reducing the harmful effects of salt stress [19-21]. Many efforts have been made in the past to develop salinity-tolerant wheat genotypes. Using traditional breeding procedures, successful results have been reached in the form of varieties such as KRL 19, KRL 1-4, KRL 210, and others [22], no such efforts are made to breed for a salt-tolerant dicoccum wheat genotype or to identify a salt tolerant dicoccum wheat genotype. However, a significant increase in wheat grain production under salinity stress has yet to be achieved.

The sum of squares of the difference between the mean values obtained between any two genotypes for all morpho-physiological, yield, and yield attributing traits in sixty wheat genotypes under two environments *viz.,* saline and control condition were used for the final grouping of the genotypes. The genetic diversity analysis was carried out using Tocher's [13] approach, which was used to arrange 60 wheat genotypes into various clusters by considering the estimated $D²$ value as the square of the generalized distance. The pattern of distribution of genotypes into various clusters under saline and control condition was presented in Tables 4 and 5 respectively. Under saline condition, three clusters were created by combining all 60 wheat genotypes in such a way that the genotypes in each cluster had a lower D^2 value than the genotypes in the other clusters. Cluster I had the most genotypes *i.e.*, 58 genotypes, followed by Cluster II and III the ones that were solitary with genotypes DIC 18 and DIC 94 respectively. Nine clusters were formed under control condition, cluster I was the largest with 16 genotypes, followed by cluster III and cluster V with 12 genotypes each, cluster II had 7 genotypes which included the checks. Cluster IV had six genotypes and Cluster VI had four genotypes, cluster VII, VIII, IX were solitary clusters with genotypes KRL 210, UAS 334 and DIC103 respectively.

The divergence analysis showed that all sixty genotypes were grouped into three clusters in saline condition and seven clusters, under control condition. A maximum number of genotypes were grouped into cluster I under both saline and control condition. Genotypes namely DIC 18 of cluster II and DIC 94 of cluster III in saline environment were showing high intercluster distance. Hybridization between the genotypes of these clusters might yield desirable segregants (Table 6). These results are supported by those of Mundiyara et al. [23], Vinod et al. [24], Singh et al. [25], Ibrahim and Arafa [26] and Srivijay et al. [27].

The highest intra-cluster distance was obtained by cluster I (167.61), and between clusters II and III highest inter-cluster distance (863.27) was obtained under saline condition, followed by clusters I and III (589.92), represented in Table 6. Under non-stress condition (Table 7), cluster IV (44.50) showed the highest intra-cluster distance, clusters IX and VII were showing the highest inter-cluster (222.80) distance, followed by cluster IX and cluster II (211.02). The cluster means for all the characters under saline conditions are represented in Table 8. The solitary cluster III had early flowering having the genotype DIC-94 (55 days), high grain yield (2,735 kg/ha) and the highest mean values for biomass (18,977 kg/ha), number of tillers per meter (157.00), spike length (11.41cm), germination percent (96.50 percent), plant height (104.73 cm), NDVI at anthesis stage (0.70), lowest NDVI at booting stage (0.37), chlorophyll content at anthesis stage (34.68), number of spikelets per spike (15.00) and least harvest index of 14.40 percent were recorded by this cluster. Under control condition (Table 9), cluster IX with genotype DIC 103 had the highest grain yield (5428.62 kg/ha), biomass (12965.64 kg/ha), late flowering (69 days), late maturing (118 days), highest harvest index (42.06%) and highest NDVI at grain filling stage (0.74). Cluster VII with genotype KRL 210 was showing the highest mean values for number of spikelets per spike (51), spike length (11.08 cm) and highest grains per spike (49.50).

The percent contribution of each character towards total genetic divergence under saline and control condition is presented in Table 10. It was observed that under saline condition, biomass (85.19%) was the largest contributor to the divergence. Days to maturity (6.28%), tillers per meter row (4.32%) ranking III and IV contributed the highest to the diversity among various yield attributes. On the contrary, the contribution of other yield attributes such as

SI.no	Trait	Saline							Control		
		Mean	CV	SE	SS	MSS	Mean	c_{V}	SE	SS	MSS
	GY	19.88	7.57	10.65	300780.77	5013.01	36.23	0.54	13.78	909783.54	15163.05
$\overline{2}$	TPM	93.81	2.21	1.47	126717.16	2111.95	122.02	2.00	1.72	78828.97	1313.82
3	BM	84.01	0.25	14.56	8196229.52	136603.82	114.24	13.17	10.64	11144858.62	185747.64
4	SPS	19.43	4.81	0.66	6589.48	109.82	20.27	5.48	0.79	10811.57	180.19
5	SL	6.63	1.42	0.07	144.67	2.41	8.58	4.77	0.29	195.92	3.27
6	GPS	29.28	6.20	1.28	3666.52	61.11	34.23	5.71	1.38	5732.57	95.54
7	PН	81.35	1.79	1.03	15328.41	255.47	94.79	3.47	2.32	8652.79	144.21
8	GP	92.89	4.21	2.76	1307.11	21.79	94.44	4.02	2.68	826.10	13.77
9	TGW	29.92	1.52	0.32	2456.77	40.95	35.42	1.58	0.40	724.39	12.07
10	DFF	60.01	0.81	0.35	3500.49	58.34	62.15	0.63	0.28	3426.25	57.10
11	DM	107.37	0.37	0.28	4329.60	72.16	109.52	0.37	0.28	4360.24	72.67
12	HI	25.25	7.97	1.42	10426.69	173.78	33.38	15.95	3.76	9622.01	160.37
13	NDVI-I	0.53	3.10	0.01	1.23	0.02	0.65	13.92	0.06	0.84	0.01
14	NDVI-II	0.59	3.19	0.01	1.30	0.02	0.65	14.30	0.07	0.79	0.01
15	NDVI-III	0.62	17.95	0.08	1.04	0.02	0.60	19.03	0.08	0.98	0.02
16	SPAD-I	43.50	13.15	4.04	1818.11	30.30	43.88	11.61	3.60	2203.52	36.73
17	SPAD-II	37.71	12.31	3.28	1646.48	27.44	41.23	10.46	3.05	1905.92	31.77
18	SPAD-III	36.69	19.55	5.07	1823.46	30.39	41.48	12.29	3.60	2128.23	35.47

Table 3. ANOVA for alpha lattice design for different traits under saline and control condition

GY- Grain yield (q/ha), TPM-Tillers per meter, BM- Biomass (q/ha), SPS- Spikelets per spike, SL-Spike length (cm), GPS-Grains per spike, PH-Plant height (cm), GP-Germination percentage (per cent), TGW-Thousand grain weight (g), DFF- Days to 50 per cent flowering, DM-Days to maturity, HI-Harvest index, NDVI- anthesis stage booting <i>Germination percentage (per cent), TGW-Thousand gr and grain filling stages, SPAD- Chlorophyll content before anthesis, booting and grain filling stages

Table 4. Clustering of wheat genotypes based on morpho-physiological, yield and yield attributing traits under saline condition

Table 5. Clustering of wheat genotypes based on morpho-physiological, yield and yield attributing traits under control condition

Table 6. Intra and inter-cluster D² values in wheat genotypes under saline condition

	Cluster I	Cluster II	Cluster III	Cluster IV	Cluster V	Cluster VI	Cluster VII	Cluster VIII	Cluster IX
Cluster I	33.46								
Cluster II	121.21	36.88							
Cluster III	62.88	78.11	37.87						
Cluster IV	53.79	106.30	64.74	44.18					
Cluster V	67.63	167.30	113.63	81.41	44.50				
Cluster VI	100.58	62.88	56.75	94.01	153.03	40.96			
Cluster VII	136.01	51.14	90.81	130.53	185.17	75.55	0.00		
Cluster VIII	71.58	79.80	58.08	76.65	111.12	86.55	83.88	0.00	
Cluster IX	99.13	211.02	150.29	125.55	64.01	191.64	222.80	146.63	0.00

Table 7. Intra and inter-cluster D² values in wheat genotypes under control condition

Table 8. Cluster means for various quantitative traits in wheat genotypes under saline situation

GY- Grain yield (q/ha), TPM-Tillers per meter, BM- Biomass (q/ha), SPS- Spikelets per spike, SL-Spike length (cm), GPS-Grains per spike, PH-Plant height (cm), GP-Germination percentage (per cent), TGW-Thousand grain weight (g), DFF- Days to 50 per cent flowering, DM-Days to maturity, HI-Harvest index, NDVI- anthesis stage booting and grain filling stages, SPAD- Chlorophyll content before anthesis, booting and grain filling stages

	GY	TPM	BM	SPS	SL	GPS	PH	GP	TGW
Cluster I	3940.76	117.06	12199.62	15.91	7.89	31.56	96.47	94.38	36.32
Cluster II	2480.07	112.56	9990.45	35.56	9.74	34.63	86.52	93.94	35.30
Cluster III	3138.67	116.42	11040.57	18.00	8.51	33.46	94.82	94.46	34.79
Cluster IV	3653.37	157.08	12281.30	19.33	9.08	39.75	99.50	94.08	35.89
Cluster V	4786.99	131.04	12743.90	17.50	8.66	36.38	100.49	95.08	35.55
Cluster VI	2383.67	125.63	7994.25	13.63	7.52	27.75	89.38	94.63	34.22
Cluster VII	2429.39	81.00	6599.81	51.00	11.08	49.50	76.33	92.50	31.33
Cluster VIII	3727.00	74.50	11160.66	33.00	7.79	31.50	78.00	94.50	34.54
Cluster IX	5428.62	99.50	12965.64	17.00	9.95	37.50	94.17	95.00	34.52
	DFF	DM	HI	NDVI-I	NDVI-II	NDVI-III	SPAD-I	SPAD-II	SPAD-III
Cluster I	65.62	113.31	35.38	0.67	0.67	0.61	43.79	40.95	40.51
Cluster II	52.06	98.75	25.79	0.59	0.60	0.57	49.02	44.69	45.44
Cluster III	63.13	110.62	30.35	0.68	0.68	0.58	42.90	40.54	41.24
Cluster IV	61.32	108.96	32.01	0.58	0.62	0.56	40.90	38.01	41.18
Cluster V	63.91	111.14	39.18	0.63	0.61	0.62	42.87	40.68	41.38
Cluster VI	64.36	112.94	30.88	0.63	0.63	0.67	42.95	39.93	37.76
Cluster VII	52.84	95.20	37.95	0.72	0.71	0.60	44.46	42.75	42.22
Cluster VIII	52.98	97.62	33.98	0.74	0.74	0.63	46.16	53.00	42.35
Cluster IX	68.92	118.04	42.06	0.70	0.68	0.74	46.99	44.45	44.13

Table 9. Cluster means for various quantitative traits in wheat genotypes under control situation

GY- Grain yield (q/ha), TPM-Tillers per meter, BM- Biomass (q/ha), SPS- Spikelets per spike, SL-Spike length (cm), GPS-Grains per spike, PH-Plant height (cm), GP-Germination percentage (per cent), TGW-Thousand grain weight (g), DFF- Days to 50 per cent flowering, DM-Days to maturity, HI-Harvest index, NDVI- anthesis stage booting and grain filling stages, SPAD- Chlorophyll content before anthesis, booting and grain filling stages

thousand-grain weight (1.69%), spikelets per spike (0.33%), spike length (0.93%) and grains per spike (0.05%) was less. Among the morphological traits, days to flowering 0.27 percent to diversity. The NDVI reading at the booting stage (0.16%) and anthesis (0.11) were observed to be important. Under control condition, it was observed that among various groups of traits in general, the contribution of grain weight (66.12%) was found highest, followed by NDVI at the anthesis stage (12.24%), days to maturity (10.49%), tillers per meter (5.14%), days to 50 percent flowering (3.39%) and spikelets per spike (1.97%) respectively.

The utility of the D^2 statistic is enhanced by its applicability to estimate the relative contribution of various characters to the total genetic divergence. The feature that contributes the most to divergence is utilized to place a higher emphasis on selecting a cluster for further selection and selecting parents for hybridization. It has been noted that under saline condition, biomass (85.19%) was the largest contributor to the divergence. This suggests that genotype DIC-94 belonging to cluster III exhibiting a high mean for biomass, grain yield, tillers per meter, spike length, thousand-grain weight, plant height, and germination percent can be a potential

source for hybridization program to obtain transgressive segregants. These results are in concordance with Sunilyadav et al. [28] wherein they observed a higher contribution of thousandgrain weight, tillers per meter, days to maturity, days to flowering, and grain yield towards diversity.

The majority of wheat genotypes are vulnerable to salt stress, resulting in low and unpredictable yields. Because current screening approaches for salinity-related traits are time-consuming and impracticable for most researchers, it is critical to estimate genotype yield under saline and control condition, and tolerance indices are currently being used to quantify the effect of salt on yield. These salt tolerance indices indicate whether the genotypes are salt-tolerant or susceptible to the saline condition. To investigate suitable stress resistance indices for screening of genotypes under salt condition, the grain yield of cultivars under both non-stress and stress condition were measured for calculating different sensitivity and tolerance indices. Among them, stress susceptibility (Table 12) and stress tolerance index (Table 12) have been considered for
grouping of genotypes. These indices grouping of genotypes. These indices categorized the sixty wheat genotypes into three categories *viz.,* tolerant, moderately tolerant, and

Table 10. Per cent contribution of morpho-physiological traits, yield and yield attributes to total genetic diversity under control and saline condition

SI. No.	Source	Per cent contribution					
		Saline condition	Control condition				
1	GY		66.12				
$\overline{2}$	TPM	4.32	5.14				
3	ВM	85.19	0.05				
4	SPS	0.33	1.97				
5	SL	0.93					
6	GPS	0.05	0.11				
$\overline{7}$	PH	0.66					
8	GP						
9	TGW	1.69	0.38				
10	DFF	0.27	3.39				
11	DM	6.28	10.49				
12	HI						
13	NDVI-I	0.16					
14	NDVI-II	0.11	12.24				
15	NDVI-III						
16	SPAD-I						
17	SPAD-II		0.11				
18	SPAD-III						

GY- Grain yield (q/ha), TPM-Tillers per meter, BM- Biomass (q/ha), SPS- Spikelets per spike, SL-Spike length (cm), GPS-Grains per spike, PH-Plant height (cm), GP- Germination percentage (per cent), TGW-Thousand grain weight (g), DFF- Days to 50 per cent flowering, DM-Days to maturity, HI-Harvest index, NDVI- anthesis stage booting and grain filling stages, SPAD- Chlorophyll content before anthesis, booting and grain filling stages

Table 11. Grouping of wheat genotypes based on Stress susceptibility index (SSI)

Table 12. Grouping of wheat genotypes based on stress tolerance index (STI)

susceptible. The tolerant category of SSI included 14 genotypes. Among 13 tolerant genotypes, eight dicoccum wheat genotypes (DIC-9, DIC-13, DIC-68, DIC-73, DIC-88, DIC-94, DIC-95, DDK 50505), along with five checks *viz.,* KRL 210, UAS 334, Kharchia 65, KRL 3-4, and IC 0408331. Based on the stress tolerance index (STI), only 10 genotypes *viz.,* DIC-26, DIC-48, DIC-92, DIC-101, DIC-104, DIC-107, DDK 50507 and UAS 334 were said to be tolerant.

The stress susceptibility index was evaluated based on the yield ratio of each variety in stressed conditions to non-stressed conditions as compared with the proportions in total varieties. Thus, tolerant cultivars identified with high yield or low in both conditions can have the same amount of SSI, so the selection process based on this index led reformers to make a mistake [29]. So, according to the stress-sensitive index, it was observed that 13 genotypes *viz*., eight dicoccum wheat (DIC-9, DIC-13, DIC-68, DIC-73, DIC-88, DIC-94, DIC-95, DDK 50505) and five checks (KRL 210, UAS 334, Kharchia 65, KRL 3- 4, IC 0408331) as a tolerant. A similar classification of genotypes using SSI was done for 97 bread-wheat genotypes by Sardouie et al. [30]. The stress tolerance index (STI) was defined as a useful tool for determining the high yield and stress tolerance potential of genotypes [16]. Based on the stress tolerance index, only 8 genotypes were identified as salt tolerant *viz.,* seven dicoccum wheat genotypes (DIC-26, DIC-48, DIC-92, DIC-101, DIC-104, DIC-107, DDK 50507) and check UAS 334. Mevlut and Sait [31] indicated that the genotypes with >0.9 STI value, usually have high-yielding potential under stress condition even greater differences in two different conditions. Hence, there is still scope for both the indices and generating super salt tolerant varieties.

With respect to dicoccum wheat genotype (DIC-88) based on SSI and few genotypes (DIC-101, DIC-92, DIC-48, DIC-26, DDK 50507) based on STI accept the genotype DIC 101 which is the most promising considering both SSI and STI can be involved in biparental mating to complement the mechanisms operating in both categories to improve both salt tolerance and yield simultaneously. It is clear from the present study that there is a lot of scope for improvement of all these genotypes using the potential donors identified. Genome 'A' has the next highest genes or QTLs followed by 'D' genome of hexaploid wheat. Mechanisms like tissue level compartmentalization *i.e.*, accumulation of toxic

ions in the lower leaves of the plant is noted in dicoccum wheat. Cell level compartmentalization in the cell vacuoles was noted by Munns and Tester [32].

4. CONCLUSION

The study's findings indicate that dicoccum wheat germplasms possess significant potential as sources of salt tolerance. Additionally, a handful of promising accessions could be officially recognized as national genetic resources or developed into new wheat varieties. This research underscores the untapped possibilities offered by dicoccum wheat species in addressing global challenges related to hunger and growing populations, particularly in regions characterized by unproductive, saline soils.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. USDA, United States Department of Agriculture.
	- Available:http://www.fas.usda.gov 2018
- 2. Alexandratos N, Bruinsma J. World agriculture towards 2030/2050: the 2012 revision.
- 3. Shiferaw B, Smale M, Braun HJ, Duveiller E, Reynolds M, Muricho G. Crops that feed the world 10. Past successes and future challenges to the role played by wheat in global food security. Food Secur. 2013; 5(3):291-317.
- 4. D'Antuono LF, Scazzina F, Zhang Y. Physical properties and chemical composition of *Triticum monococcum* L. and Triticum dicoccum Schübler wheat bran. Journal of Cereal Science, 2002; 36(2):207-216.
- 5. Lupo L. Rediscovering the tradition of *Triticum dicoccum* (farro): A review. Journal of Nutritional Science. 2017;6:e48.
- 6. Shamsi K, Petrosyan M, Noor-mohammadi G, Haghparast A, Kobraee S, Rasekhi B. Differential agronomic responses of bread wheat cultivars to drought stress in the west of Iran. Afr. J. Biotechnol. 2011; 10(14): =2708-2715.
- 7. Khavarinejad MS, Karimov M. Study of Genetic diversity among spring wheat genotypes in drought stress by advanced

statistical analysis. Int. J. Agron. Plant Prod. 2012;3(12):590-598.

- 8. Greenway H, Munns. Mechanism of salt tolerance in non-halophytes. Annu. Rev. Plant. Physiol. 1980;31:146-190.
- 9. Flowers TJ, Garcia A, Koyama M, Yeo AR. Breeding for salt tolerance in crop plants, the role of molecular biology. Acta Physiol. Plant. 1997;19(4):427-433.
- 10. Ponnamieruma PN. Role of cultivars tolerance in increasing rice production on saline land. In R. C. Staples & G. H. Toenniessen (Eds.), Salinity tolerance in plants strategies for crop improvement. Wiley, New York. 1984;255-71.
- 11. Mondal AK, Sharma RC, Singh G, Dagar JC. Computerised database on salt affected soils in India. Tech. Bull. 2010;2/2010. CSSRI, Karnal.
- 12. Mahalanobis PC. on the generalised distance in statistics. Proceed. Nat. Instit. Sci. 1936;2:49-55.
- 13. Rao CR. Advanced Statistical Methods in Biometrieal Research. John Wiley and Sons, Inc., New York. 1952;357- 363.
- 14. Singh R K, Chaudhary BD. Biometrical methods in quantitative genetic analysis, Kalyani Publishers, New Delhi. pp. 1977; 57-58.
- 15. Fischer RA, Maurer R. Drought resistance in spring wheat cultivars. I. Grain responses. Australian J. Agric. Res. 1978; 29:897–912.
- 16. Fernandez GCJ., Effective selection criteria for assessing plant stress tolerance. In: Kuo CG (ed) Proceedings of the International Symposium on Adaptation of Vegetables and other Food Crops in Temperature and Water Stress, Publication, Tainan, Taiwan. 1992;257- 270.
- 17. Khanna-chopra R, Vishwanathan C. Evaluation of heat stress tolerance in irrigated environment of *T. aestivum* and related species. I. Stability in yield and yield components. Euphytica, 1999;106(2): 169-180.
- 18. Sang HK, Dae YK, Yacoubi I, Seo YW. Phenotypic and genotypic analyses of drought tolerance in Korean and Tunisian wheat cultivars. Biotechnol. 2014;2:139- 150.
- 19. Ashraf M, O'Leary JW. Responses of some newly developed salt-tolerant genotypes of spring wheat to salt stress. Yield components and ion distribution.

J. of Agron. and Crop Sci. 1996;176:91– 101.

- 20. Flowers TJ, Yeo AR. Breeding for salinity resistance in plants. Aust. J. Plant Physiol. 1995;22:875-884.
- 21. Singh KN, Rana RS. Genetic variability and character association in wheat varieties grown on sodic soil. Indian J. of Agri Sci. 1985;55:723-726.
- 22. Shahbaz M, Ashraf M. Improving salinity tolerance in cereals. Crit. Rev. Plant Sci. 2013;32(4):237-249.
- 23. Mundiyara R, Kerkhi SA, Jakhar ML, Ganga R. D2 analysis in bread wheat (*Triticum aestivum* L.). J. Plant Sci. Res. 2013;29(2):215-219.
- 24. Vinod K, Devendra KP, Saiprasad SV. Genetic divergence analysis in durum wheat (*Triticum durum* Desf.). Int. J. Cur. Res. 2014;6(6):7001-7005.
- 25. Singh S, Sengar RS, Kulshreshtha N, Datta D, Tomar RS, Rao VP, Garg D, Ojha A. Assessment of multiple tolerance indices for salinity stress in bread wheat (*Triticum aestivum* L.). J. Agric. Sci. 2015; 7(3):49.
- 26. Ibrahim HM, Arafa SA. Effect of Chitosan on Growth, Yield and Certain Salinity Stress-Related Metabolites in Two Barley Cultivars Contrasting in Salt Tolerance. J. Plant Prod. Sci. 2020;11(10): 899-906.
- 27. Srivijay Malipatil, SS Biradar, SA Desai, SS Gundlur. Genetic variability studies for yield and its attributes under salt affected soils in wheat species. J. Farm Sci. 2022; 35(2):169-173.
- 28. Sunilyadav K, Singh A K, Baghel SS, Mahesh J. Assessment of Genetic variability and Diversity for Yield and its Contributing Traits among CIMMYT Based Wheat Germplasm. J. Wheat Res. 2014; 6(1):154-159.
- 29. Naeimi M, Akbari GA, Shiranirad AH, Modares SS, Sadat NS, Jabari H. Evaluation of drought tolerance in different canola cultivars based on stress evaluation indices in terminal growth Duration. 2008;83-98.
- 30. Sardouie Nasab S, Mohammadi Nejad G, Nakhoda B. Field screening of salinity tolerance in Iranian bread wheat lines. Crop Sci. 2014;54(4):1489- 1496.
- 31. Mevlut A, Sait C. Evaluation of drought tolerance indices for selection of Turkish oat (*Avena sativa* L.) landraces under

Malipatil et al.; Int. J. Environ. Clim. Change, vol. 13, no. 11, pp. 2609-2621, 2023; Article no.IJECC.109202

various environmental conditions. Zemdirbyste Agriculture. 2011;98(2):157- 166.

32. Munns R, Tester M. Mechanisms of salinity tolerance. Annu. Rev. Plant Biol. 2008;59: 651–681.

© 2023 Malipatil et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License [\(http://creativecommons.org/licenses/by/4.0\)](http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/109202