



Annual Research & Review in Biology

24(6): 1-12, 2018; Article no.ARRB.39614
ISSN: 2347-565X, NLM ID: 101632869

The Role of TNF-Alpha, Beta, XRCC1 and p53 Gene Predisposition in Cancers

Swapna Kollabathina^{1*} and G. Sudhakar¹

¹Department of Human Genetics, Andhra University, Visakhapatnam, India.

Authors' contributions

This work was carried out in collaboration between both authors. Author SK designed the study, performed the statistical analysis, wrote the protocol, wrote the first draft of the manuscript. Author GS managed the analyses of the study. Both the authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ARRB/2018/39614

Editor(s):

- (1) J. David Puett, Professor, Department of Biochemistry and Molecular Biology, University of Georgia, Athens, USA.
(2) George Perry, Dean and Professor of Biology, University of Texas at San Antonio, USA.

Reviewers:

- (1) Lawrence M. Agius, University of Malta, Malta.
(2) Tabe Franklin Nyenty, University of Ngaoundere, Cameroon.
(3) Bensu Karahalil, Gazi University, Turkey.

Complete Peer review History: <http://www.sciencedomain.org/review-history/23592>

Review Article

Received 8th December 2017
Accepted 26th February 2018
Published 12th March 2018

ABSTRACT

Cancer is a group of diseases characterized by abnormal and uncontrolled cellular growth and differentiation. The major cause of Cancer consists of different types of genetic alterations that include gene substitutions, point mutations and gene amplification leading to damage in molecular pathways responsible for cell growth, survival, and metastasis. These changes develop in the majority of cases with a specific type of tumor. Genes can be used as biomarkers for identification and development of targeted treatments and also for predicting various responses to therapies. The aim of the present review is to reveal the role of tumorigenesis in TNF alpha, beta, xrcc1 and p53 genes in cancer susceptibility to gene predisposition.

Keywords: Tumorigenesis; tumor necrosis factor; XRCC1; p53; predisposition; SNPs.

1. INTRODUCTION

*Corresponding author: E-mail: kollabathinaswapna@yahoo.in, kollabathinaswapna@gmail.com;

Cancer is a group of deadly diseases it is characterized by abnormal and uncontrolled cellular growth and differentiation. Multiple genetic expression confers a survival advantage and unlimited proliferative potential to somatic or germ cells. Complex relations like genetic and environmental factors cause cancer to any part of the body due to damage to DNA. Novel techniques in biomedical research make an easy way to identify several genes as biomarkers of cancers for the identification, screening, diagnosis, and treatment. Another use of biomarkers is to find the early perception of cancer, design sporadic therapies for the diseases including signaling molecules associated with cellular development, metabolism, and apoptosis, inculcating to observe the functional damage with molecular and cellular obstructions. Major exposition of genetic alterations exhibited in three classes of genes proto-oncogenes, tumor suppressor genes, and DNA repair genes grouping together for the contribution of the development of cancer genotypically and phenotypically. These genetic alterations include gene substitutions, point mutations and gene amplifications that lead to damage in molecular pathways responsible for cell growth, survival, and metastasis. When such changes develop in the majority of cases with a specific type of tumor, genes can be used as biomarkers for identification and development of targeted treatments besides predicting gene responses to various therapies [1]. The present review showed the role of tumorigenesis in TNF alpha, beta, xccc1 and p53 genes with cancer susceptibility and genetic predisposition.

2. PRO-TUMORIGENIC-TUMOR NECROSIS FACTOR-ALPHA (TNF- α)

The central premise of this review article correctly identifies as the axial involvement of inflammation and chronic inflammation in carcinogenesis. Chronic inflammatory mediators exhibit multiple effects in the development of tumors, the growth of the tumor depends on intrinsic and extrinsic pathways of inflammation. On the other hand, inflammation highly supports carcinogens, malignant formation, tumor growth invasion, and metastasis. Among all other Cancers, the 20% of cancers are combined with chronic inflammation and most of it contains solid tumors with inflammatory infiltrates. Tumor initiation, growth, and progression of many immune cell effects are mediated by pro-inflammatory cytokines. The established pro-tumorigenic cytokine is a tumor necrosis factor alpha (TNF- α). Tumorigenesis modifies these

cytokines as attractive targets for accurate treatment of cancer. The TNF- α gene acts as master regulator gene of tumor-associated inflammation and defense against infection. This cytokine is involved in multiple inflammatory and immune responses and plays role in the pathogenesis of many autoimmune and infectious diseases. TNF- α is also called TNF, cachexin, or cachectin, it regulates the transcription levels [2,3]. The human TNF gene (TNF- α) was discovered in 1985 [4]. It is situated on chromosome 6p21, contains 4 exons. The last exon consists of more than 80% of the secreted protein [5]. TNF actively binds two receptors, TNFR1 and TNFR2. TNFR1 is expressed in many tissues, and totally operate the membrane-bound and soluble trimeric forms of TNF, and the other hand TNFR2 is observed in the immune cellular system and respond to the membrane-bound form of the TNF homotrimer. The death domain of TNFR-1 indicates the capacity of inducing apoptotic cell death, the major differences between these two receptors are the absence of death domain in TNFR2 [6].

The process of growth stimulating, inhibitory and self-regulatory properties of the body organs are influenced by TNF- α . The functional role of TNF- α is to control the immune cells, inflammation, represses tumorigenesis, viral replication and responds to Interleukin 1 and Interleukin 6 producing cells and TNF- α also declined to regulate their production leading to a variety of human cancers [7] and inflammatory bowel disease (IBD) [8]. TNF- α promotes chronic impacts along with acute pathological regulations, causes necrosis of some types of tumors. It stimulates the growth of other types of tumor cells. The increasing levels of TNF- α correlate with high risk of mortality [9]. TNF- α is involved in inflammatory disorders of inflammatory and non-inflammatory origin [10]. The storage of TNF- α in the body for a long time loses its antitumor activity with polymerization of the cytokine due to the shedding of TNF receptors by tumor cells. However, large production of anti-TNF antibodies is found in patients with carcinomas or chronic infection. It also leads to disruptions in the alpha-2 macroglobulin proteinase system cytokine deregulation. Continuous overproduction of TNF- α leads to a condition called as cachexin, characterized by anorexia, catabolism, weight loss and anemia and this large production of TNF- α occurs in the illness of cancer.

3. TNF- α ROLE IN DIFFERENT CANCERS

The polymorphism at -308 positions in G to A transition of the TNF- α gene is located within the binding site of the AP-2 repressive transcription factor. The less common allele A2 (-308A) results from a lack of AP-2 binding and, leads to increased TNF- α gene expression [11]. The A2 allele is correlated with some malignancies like breast, gastric cancer and possibly with thrombosis [12,13]. Microsatellite polymorphisms situated in the TNF- α locus reports association in increased risk of gastric and colorectal cancer [14]. Elevated levels of serum with TNF- α protein was observed in European patients with oral cancer, but no association was found in Taiwanese patients [15,16]. Several reports indicated that serum concentrations with TNF are increased in various cancer patients [17,18]. TNF expression levels also higher in different pre-neoplastic and tumor tissues [17-19] and the increased TNF expression levels associated with the susceptibility of malignant diseases like prostate cancer, breast cancer, and cervical carcinoma, chronic lymphocytic leukemia, Barrett's adenocarcinoma [17,18,20] colorectal neoplasia [21]. The decreased levels of serum TNF concentration was observed in breast and prostate cancer patients at the time of chemotherapy treatment [22]. These findings showed that TNF involved in a major tumor progression and it is considered as a prime risk of prognosis for cancer patients. Single nucleotide polymorphisms (SNPs) at position -308 in the TNF promoter (-308G/A) is found to be predisposed to various types of cancer. The person carrying higher TNF expression levels with TNF-308G/A had an increased cancer risk, mainly invasive cervical cancer [23], gastric cancer [24], non-Hodgkin's lymphoma [25,26], ulcerative colitis-associated colorectal cancer [27], hepatocellular carcinoma [28], and non-small cell lung cancer [29]. A study performed in a north European population, showed significance with the vascular invasion of breast cancer [30] and the results suggest that the -308A allele promotes and associates with cancer development and progression. Another study mentioned that the -238A allele in the TNF-promoter has decreased susceptibility to lung cancer, gastric cancer, uterine cervical cancer, colorectal cancer, or renal cell carcinoma [31]. The TNF-857T allele increases the risk of B-cell lymphoma [32]. These reports indicated that TNF has a tumor-promoting character and TNF promoter SNPs are the major associates of cancer risk. Nowadays TNF is used in cancer treatment in the isolated limb perfusion (ILP)

adjustment of soft tissue sarcoma (STS), and metastases confined to the limb [33].

4. TUMOR NECROSIS FACTOR BETA (TNF- β)

TNF- β also known as Lymphotoxin- α is belonging to a major histocompatibility complex-related with TNF family. It is obtained from the Th1 type T-cells and promotes vascular endothelial cells to modify their surface adhesion molecules to bind the phagocytic cells. It is a potent multifunctional cytokine regulates T and B lymphocytes. The functional activities are similar to TNF- α , but TNF- β is most important for the origin of lymphoid organs. These cytokines provide essential coordinated pathways which lead to communication for inflammatory and immune responses. Reactions from the specific cellular receptors, these cytokines act as a signaling pathway caused by cell death, survival, and differentiation. The Lymphotoxin also consists of similar activities like TNF produced by lymphocytes, other than macrophages or neutrophils. The protein consists of a specific potent cytotoxic activity on tumor cells with multiple target cells. The recombinant TNF-beta contains a 171 amino acid fragment, for the mature TNF-beta chain protein and is elevated in *E. coli* combined with amino-terminal hexahistidine tag [34,35]. The protein binds the lymphotoxin-alpha and lymphotoxin-beta. The encoded protein-ligand plays a role in the growth and operation of lymphoid tissue and transformed cells. The Function of the encoded protein leads to apoptosis and release of the cytokine interleukin 8.

5. TNF - β ROLE IN DIFFERENT CANCERS

The first intron of the TNF - β gene with G/A polymorphism located at position 252, showed elevated expressions in the concentration of TNF- α and TNF- β proteins in plasma [36,37]. Increasing levels of TNF- β expressions of the mRNA and the protein levels were also exhibited with the correlation of the less common allele B1 (252G) [36]. The TNF- β (G252A) polymorphism is also associated with increased risk of development in a breast, esophageal, gastric and colorectal cancer [38]. A study reveals that the allele frequency of the B1 homozygotes is higher than 16% in Europeans and 13% in Asians [38]. The combinational effects of TNF- α and TNF- β polymorphisms were obtained in patients with breast or esophageal cancer, the genotypes

exhibited significant overexpression levels and associated with an increased risk of cancer development [38]. TNF- β with high expression of B1 allele was also observed with a significantly increased risk in patients with osteosarcoma, breast, colorectal and bladder cancer [39]. The TNF- α and TNF- β genotypes together detected in a high ratio of genotypes with A2A2/B1B1, A1A2/B1B2, and A1A2/B1B1, it indicates that the TNF- α gene, TNF- β also plays an important role in the significant risk of oral cancer, though B1 increases TNF- β production [40]. The genotypes of A1A2/B1B2 reported a low risk of oesophageal and gastric cancer in Chinese, and the major risk of myeloma in English Caucasians [41].

6. DNA REPAIR GENES-XRCC1

The responsibilities of Genomic stability and integrity are maintaining the exact DNA replication. DNA repair systems play a major role in regulating the human genome integrity. Loss of ability in the repair capacity due to either mutations or inherited polymorphisms in DNA repair genes leads to differentiation of the DNA repair ability. Modifications in DNA repair genes are associated with various cancers [42]. A cell that has obtained a large amount of DNA damage and no longer capacity to repair damage incurred to its DNA, leads to one of the three possible states including an irreversible state of dormancy, and cell suicide or apoptosis or programmed cell death, unregulated cell division, which can form cancerous tumors. DNA repair systems are accumulating several multienzymes, multistep processes, are fundamental to the controlling of genomic stability and integrity in the replication errors, environmental depreciation and the cumulative changes of aging. The DNA-repair enzymes influence the DNA adducts levels through genetic polymorphisms [43]. DNA repair capacity may increase the risk of the disease. Most of the DNA repair genes, X-ray repair cross-complementing group1 (XRCC1) associations elevated in various cancers like breast [42] lung [44], head and neck [45] Bladder cancer [46] Squamous cell carcinoma of head and neck [47], and XRCC1 is a protein present in humans and it is a major gene involved in DNA repair. It is located on chromosome 19q13, involving 17 exons and encodes 633 amino acids. It is important for mammalian viability with efficient repair of single-strand breaks and damaged bases in DNA [48]. The major function of XRCC1 is a modulator of the multiple activities involved in base excision repair in mammalian

cells through interacting with and combining together of DNA polymerase β , DNA ligase III and forms a repair complex [49] and poly (ADP-ribose) polymerase in the N-terminal, C terminal and central regions of XRCC1 gene. This pathway repairs the damaged DNA caused by a different variety of endogenous and exogenous factors involving oxidation, alkylating agents and ionizing radiation [50].

This protein binds directly to nicked DNA, the association of gapped DNA with DNA polymerase Beta, resulting from protein directly involved in independent DNA damage recognition [51]. Although mutations in XRCC1 have been showed to disrupt the protein function by modifying binding sites or catalytic domain of the protein [52]. The deficiency of XRCC1 causes reduced a capacity of genetic stability and increases the spontaneous or induced chromosome translocations and deletions [53]. Base excision DNA repair (BER) is one of the major proactive DNA repair pathways for important cellular defense mechanism against the harmful effects of the regularly damaged or improper bases in DNA [54]. DNA repair enzymes regulate the DNA to repair the damaged nucleotide residues formed by replication or carcinogen exposures and compounds of cytotoxic mutations in carcinogenesis [55]. Major damage to DNA and its repair is a cause for various types of cancer [56-59]. Lack of DNA repair efficiency leads to mutations in genes, because of the functional loss of the DNA repair protein. Several mutations and polymorphisms are expressed in the number of genes encoding for DNA repair enzymes. DNA damage is a form of cell stress and injury that has been involved in many cancers. The relation between oxidative induced stress and different cellular defense mechanisms leads to oxidative injury. Sometimes oxidative stress results in DNA or protein damage leading to cell suicide (apoptosis). Cell injuries and cell stress are repaired by antioxidant enzymes and DNA repair mechanisms.

7. XRCC1 ROLE IN DIFFERENT CANCERS

The studies suggested that the XRCC1 gene having 300 SNPs, the extensively studied SNPs are XRCC1 Arg194Trp and XRCC1 Arg399Gln [60]. Many studies have indicated that the XRCC1 Arg194Trp polymorphism increases the development of different kinds of cancers like glioma and thyroid cancer, but several meta-

analyses revealed that no correlation between XRCC1 Arg194Trp polymorphism and risk of head and neck cancer, hepatocellular carcinoma, lung cancer [61] on the other hand XRCC1 399Gln allele showed with multiple DNA damages in phenotypes of human cells and tissues in different types of cancers. The variant 399A allele was correlated with an increased risk of stomach cancer [62,63] lung [64,65] prostate cancer [66] and moreover, this allele has shown a decreased risk of esophageal cancer and bladder [67]. A study indicates that the 194 Arg/Arg genotype is associated cancer risk factor for oral cavity and pharynx [68]. There was a study of differences observed in NNK induced sister chromatid exchange between cells with the codon 194 Arg/Arg genotype and codon 194 Arg/Trp genotype in different types of cancers [69]. A hypothetical analysis reveals that the XRCC1-399 Gln allele is correlated with elevated risk of smoking-dependent cancers including a major risk of lung cancer [70], moreover inconsistent reports also exist [71,72]. In the case of Breast cancer, the XRCC1-399 Gln allele was associated significantly in non-smokers with detectable PAH-adducts [73].

8. TUMOR SUPPRESSOR GENES p53

The gardener gene of the genome p53 is located on chromosome 17p13 and is the single most common target for genetic variations in human tumors. An oncoprotein MDM2 with over-amplification showed in sarcomas are reported to be associated with p53 [74]. Two different mechanisms are involved in tumor formation. In the first, p53 stimulates the phosphatase 21 cyclin-dependent kinase inhibitor gene in response to DNA damage. Secondly, the loss of p53 gene in cells avoids the p21 gene transcription to the increased expression of the multiple Cdks normally turned off p21, results from cellular multiplication. P53 restraints are involved in inducing apoptosis. Activation of the p53 transcriptional factor responses starts with cellular signaling which mainly causes DNA damage. Inactive p53 and MDM2 triggers post-translational modification, apoptosis lead to altering the cell cycle through p21 and triggering the caspase cascade reactions [75]. Phosphorylation of p53 damages the ability of MDM2 to inhibit p53-dependent transactivation, mostly due to alteration of p53 [76]. Dephosphorylation of p53 enhances the ability at a consensus binding site for DNA [77]. Mutation in the arginine residue abolishes transport of p53 to the nucleus. The nuclear protein p53 binds to

single, double-stranded DNA non-specifically, along with that the mutant p53 showed the high affinity towards single-stranded DNA compared with the double-stranded DNA [78]. The pleiotropic effects of p 53 act as transcription-dependent and transcription-independent mechanism. p53 activation leads to homotetramerization with its functions of transcriptional activations also induced by transactivating target genes in intransient cell cycle arrest at G1 or G2 phase [79]. Cell cycle checkpoint arrest mainly in the stress conditions, inhibiting the accumulation of genetic errors p53 can also take action in permanent arrest, or senescence, with the help of transactivation genes like *p21CIP1/WAF1* [80]. Worldwide genome studies revealed that mutant p53 are dynamic of modulating the expression of transcripts. p53 acts as a transcription factor capable of on and off mechanism of genes in response to different gene expressions [81].

The P53 gene is a transactivation domain. The wild-type p53 gene is able to be down-regulated and amplified with proliferating cell nuclear antigen (PCNA) and can interfere with the induction of c-fos mRNA during serum stimulation [82]. The decreasing levels of p53 are observed in normal cells, but the overexpression in many tumors of different sites proposed that p53 may be a transformation-related antigen [83]. The ability of the wild-type p53 to suppress the transformation of cells in culture by other oncogenes by the growth of transformed cells in culture [84,85], the tumor formation in animals and humans is mainly due to deletions or point mutations of the wild-type p53 allele [86-88]. P53 oncogene functionally blocks the interaction of DNA polymerase viral replication like SV40 [89], and it may bind to an analogous cellular replication protein to prevent entry into S phase [90]. The p53 gene plays a critical role in cell cycle regulation and induction of apoptosis, DNA repair, and genomic instability, all of which are central to the prevention of human cancer forming agents. Inactivation of the wild-type p53 function is frequently seen during the development of cancer. Inactivation of p53 results mutation, inhibition, nuclear DNA damage, degradation protein levels through the MDM2 and aberrant transcription. The p53 tumor suppressor gene mutations associated with over 50% of all human tumors [91]. Regulations of the cell cycle arrest by the activation of p53 protein alterations due to mutations provide evidence for clonal expansion of neoplastic cells [92]. Failure of the tumor

suppressor responses, inactive p53 also incapable to activate expression of MDM2. These p53 mutant proteins are profusely unstable and accumulate in high levels in the tumor cells [93]. Functional p53 may also be inactivated through over expression of the mdm2 gene in some tumors, especially sarcomas since the MDM2 protein speedy degrades and inhibits p53 by binding to the protein [94]. p53 tumor suppressor gene essential for the DNA damage checkpoint, but it was simultaneously observed to have a larger function after cellular stress, such as oncogenes activation or hypoxia [95]. The p53 protein functions as a tetrameric transcription factor found at very low levels in normal unstressed cells. The most common genetic alterations noticed in human cancers with p53 gene mutations [96]. p53 also plays a major role in multiple DNA repair pathways including Wild-type p53 nucleotide excision repair [97]. And regulator post-transcriptional activity included phosphorylation, acylation, proliisomer aisation, and sumoylation and post-translation changes.

9. p53 ROLE IN DIFFERENT CANCERS

The polymorphism at codon 72 variant is an intragenic modifier of mutant p53 [98]. Other studies in colorectal [99], lung [100], and head and neck cancers [99] have also revealed that in Arg/Pro germline heterozygotes, the Pro allele is specifically lost and the Arg allele turns to mutated. The codon 72 Pro/Pro genotype was correlated with an increased risk of lung cancer [101]. Another study showed that the Arg/Pro genotype plays a role in the heritable incidence of smoke-related lung carcinoma [102], but inconsistent results were reported in six other case-control studies [103]. A huge study revealed that there is no association with frequent P53 polymorphisms in breast cancer [104] and ovarian cancer [105]. A study reveals an eightfold elevated risk with the intron 3 variant in ovarian cancer patients [106] and 1.93 fold risk association showed with intron 6 polymorphism in the rare Msp1 allele carried ovarian cancer patient [107]. One more study explains intron 3 polymorphism as a causative factor in sporadic and familial breast cancer [108], these results also show inconsistent association with other studies [109-111]. Although a statistically significant association in p 53 intron 6 polymorphisms showed major risk in breast and colon cancers [112]. Rare C allele polymorphism detected in sporadic breast cancer patients in Americans with high-level mutant protein p 53

[113]. But there was no association observed in Polish cancer patients with p53 germline mutations [114]. The mutations of p53 gene of all cancers are regulated by C to T transitions with the mutational methylated CpG sequences. The increased risk of G to T transversions was expressed in lung cancers of smokers than the nonsmokers and with other cancers [115]. Subcellular disruptions of wild-type p53 was highly expressed in the cytoplasm of the undifferentiated neuroblastoma [116]. Irregular cytoplasmic p53 locations are mainly seen in human primary breast cancers, colon cancers and hepatoblastomas [117].

10. CONCLUSION

This review represents that the chronic inflammation involved in tumor development and progression, TNF acts as a major pro-inflammatory cytokine and also acts as an endogenous tumor promoter to regulate inflammation and carcinogenesis in different cancers. Tumor necrosis factor constitutes a representative index of inflammation activity and hence implicates cell damage in the evolving defects in a resolution of inflammation in carcinogenesis. Even though several cell culture and animal studies are available but natural chronic inflammatory agents and their true potential will be identified from the study of the genes with their functional mechanisms in cancer development is needed for future experimental, clinical trials. The inactivation of XRCC1 DNA repair genes leads to an important aspect in cancer initiation and progression by lack of genomic stability, with multiple effects leading to malignancies. The p53 gene regulates the tumor suppressions with a nuclear transcriptive action including a pro-apoptotic function in oncogenic potential. DNA damage by cellular stresses exhibits in pro-apoptotic function. Activated p53 influences cell cycle arrest to allow DNA repair and/or apoptosis to prevent the major DNA damage through the transactivation of its target genes. Thus, the DNA-binding activity of p53 is strongly linked to its tumor suppressive action. The regulatory mechanisms of p53 and also p53-mediated therapeutic strategies is helpful to cure different cancers.

Individual differences in mechanism of these genes are to be more crucial for the development of malignancies. However, the study of these particular genes and their functions is a greater pathway for future clinical aspects and molecular

therapies to improve the survival rate of the victims.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Voorzanger-Rousselot N, Garnero P. Biochemical markers in oncology. Part I: molecular basis. Part II: clinical uses. *Cancer Treat Rev.* 2007;33:230-283.
2. Tsytsykova AV, Goldfeld AE. *Mol Cell Biol.* 2002;22:2620–2631.
3. Carswell EA, Old LJ, Kassel RL, Green S, Fiore N, Williamson B. An endotoxin-induced serum factor that causes necrosis of tumor. *Proc Natl Acad Sci, USA.* 1975;72:3666
4. Old LJ. Tumor necrosis factor (TNF). *Science.* 1985;230(4726):630-2.
5. Nedwin GE, Naylor SL, Sakaguchi AY, Smith D, Jarrett-Nedwin J, Pennica D, Goeddel DV, Gray PW. Human lymphotoxin and tumor necrosis factor genes: structure, homology and chromosomal localization. *Nucleic Acids Res.* 1985;13(17):6361-73.
6. Ashkenazi A, Dixit VM. Death receptors: Signaling and modulation. *Science.* 1998;281:1305–1308.
7. Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell.* 2001;104:487-501.
8. Mikocka-Walus AA, Turnbull DA, Moulding NT, Wilson IG, et al. Controversies surrounding the comorbidity of depression and anxiety in inflammatory bowel disease patients: a literature review. *Inflamm. Bowel Dis.* 2007;13:225-34.
9. Rink L, Kirchner H. Recent progress in the tumor necrosis factor –alpha field. *Archiv Allergy Immunol.* 1996;111:199-209.
10. Strieter RM, Kunkel SL, Bone RC. Role of tumor necrosis factor alpha in diseases states and inflammation. *Crit Care Med.* 1993;21:S447-S463.
11. Wilson AG, Symons JA, McDowell TL, Mc evitt HO, Duff GW. Effects of a polymorphism in the human tumor necrosis factor alpha promoter on transcriptional activation. *Proc Natl Acad Sci USA.* 1997;94:3195-99.
12. Guo W, Wang N, Li Y, Zhang JH. Polymorphisms in tumor necrosis factor genes and susceptibility to esophageal squamous cell carcinoma and gastric cardiac adenocarcinoma in a population of high incidence region of North China. *Chin Med J.* 2005;118:1870-78.
13. Azmy IA, Balasubramanian SP, Wilson AG, Stephenson TJ, Cox A, Brown NJ. Role of tumor necrosis factor gene polymorphism (-308 and -238) in breast cancer susceptibility and severity. *Breast Cancer. Res.* 2004;6:395-400.
14. Saito S, Kasai Y, Nomoto S, Fujiwara M, Akiyama S, Ito K, Nakao A: Polymorphism of tumor necrosis factor in esophageal, gastric or colorectal carcinoma. *Hepatogastroenterol.* 2001;48:468-470.
15. Jablonska E, Piotrowski L, Grabowska Z. Serum Levels of IL-1b, IL-6, TNF-a, sTNF-RI and CRP in patients with oral cavity cancer. *Pathol Oncol Res.* 1997;3:126-9.
16. Chen WC, Tsai MH, Wan L, Chen WC, Tsai CH, Tsai FJ. CYP17 and tumour necrosis factor-alpha gene polymorphisms are associated with risk of oral cancer in Chinese patients in Taiwan. *Acta Otolaryngol.* 2005;125:96-99.
17. Ferrajoli A, Keating MJ, Manshoury T, Giles FJ, Dey A, Estrov Z, et al. The clinical significance of tumor necrosis factor-alpha plasma level in patients having chronic lymphocytic leukemia. *Blood.* 2002;100(4):1215–9.
18. Ahmed MI, Salahy EE, Fayed ST, El-Hefnawy NG, Khalifa A. Human papillomavirus infection among Egyptian females with cervical carcinoma: relationship to spontaneous apoptosis and TNF-alpha. *Clin Biochem.* 2001;34(6):491–8.
19. Szlosarek PW, Grimshaw MJ, Kulbe H, Wilson JL, Wilbanks GD, Burke F, et al. Expression and regulation of tumor necrosis factor alpha in normal and malignant ovarian epithelium. *Mol Cancer Ther.* 2006;5(2):382–90.
20. Garcia-Tunon I, Ricote M, Ruiz A, Fraile B, Paniagua R, Royuela M. Role of tumor necrosis factor-alpha and its receptors in human benign breast lesions and tumors (in situ and infiltrative). *Cancer Sci.* 2006;97(10):1044–9.
21. Kim S, Keku TO, Martin C, Galanko J, Woolsley JT, Schroeder JC, et al. Circulating levels of inflammatory cytokines

- and risk of colorectal adenomas. *Cancer Res.* 2008;68(1):323–8.
22. Berberoglu U, Yildirim E, Celen O. Serum levels of tumor necrosis factor alpha correlate with response to neoadjuvant chemotherapy in locally advanced breast cancer. *Int J Biol Markers.* 2004;19(2): 130–4.
 23. Duarte I, Santos A, Sousa H, Catarino R, Pinto D, Matos A, et al. G-308A TNF-alpha polymorphism is associated with an increased risk of invasive cervical cancer. *Biochem Biophys Res Commun.* 2005;334(2):588–92.
 24. Machado JC, Figueiredo C, Canedo P, Pharoah P, Carvalho R, Nabais S, et al. A proinflammatory genetic profile increases the risk for chronic atrophic gastritis and gastric carcinoma. *Gastroenterology.* 2003;125(2):364–71.
 25. Bel Hadj Jrad B, Chatti A, Laatiri A, Ahmed SB, Romdhane A, Ajimi S, et al. Tumor necrosis factor promoter gene polymorphism associated with increased susceptibility to non-Hodgkin's lymphomas. *Eur J Haematol.* 2007;78(2):117–22.
 26. Rothman N, Skibola CF, Wang SS, Morgan G, Lan Q, Smith MT, et al. Genetic variation in TNF and IL10 and risk of non-Hodgkin lymphoma: a report from the InterLymph Consortium. *Lancet Oncol.* 2006;7(1):27–38.
 27. Garrity-Park MM, Loftus EV Jr, Bryant SC, Sandborn WJ, Smyrk TC. Tumor necrosis factor-alpha polymorphisms in ulcerative colitis-associated colorectal cancer. *Am J Gastroenterol.* 2008;103(2):407–15.
 28. Ho SY, Wang YJ, Chen HL, Chen CH, Chang CJ, Wang PJ, et al. Increased risk of developing hepatocellular carcinoma associated with carriage of the TNF2 allele of the -308 tumor necrosis factor-alpha promoter gene. *Cancer Causes Control.* 2004;15(7):657–63.
 29. Shih CM, Lee YL, Chiou HL, Chen W, Chang GC, Chou MC, et al. Association of TNF-alpha polymorphism with susceptibility to and severity of non-small cell lung cancer. *Lung Cancer.* 2006;52(1): 15–20.
 30. Azmy IA, Balasubramanian SP, Wilson AG, Stephenson TJ, Cox A, Brown NJ, et al. Role of tumour necrosis factor gene polymorphisms (-308 and -238) in breast cancer susceptibility and severity. *Breast Cancer Res.* 2004;6(4):R395–400.
 31. Jang WH, Yang YI, Yea SS, Lee YJ, Chun JH, Kim HI, et al. The -238 tumor necrosis factor-alpha promoter polymorphism is associated with decreased susceptibility to cancers. *Cancer Lett.* 2001;166(1):41–6.
 32. Hellmig S, Fischbach W, Goebeler-Kolve ME, Folsch UR, Hampe J, Schreiber S. A functional promoter polymorphism of TNF-alpha is associated with primary gastric B-Cell lymphoma. *Am J Gastroenterol.* 2005;100(12):2644–9.
 33. Eggermont AM, de Wilt JH, ten Hagen TL. Current uses of isolated limb perfusion in the clinic and a model system for new strategies. *Lancet Oncol.* 2003;4:429–437.
 34. Crowe PD, VanArsdale TL, Walter BN, Ware CF, Hession C, Ehrenfels B, et al. A lymphotoxin-beta-specific receptor. *Science.* 1994;264:707–10.
 35. Cannella B, Sizing ID, Benjamin CD, Browning JL, Raine CS. Antibodies to lymphotoxin alpha (LT alpha) and LT beta recognize different glial cell types in the central nervous system. *J Neuroimmunol.* 1997;78:172–9.
 36. Messer G, Spengler U, Jung MC, Honold G, Blomer K, Pape GR, et al. Polymorphic structure of the tumor necrosis factor (*TNF*) locus: An NcoI polymorphism in the first intron of the human TNF-beta gene correlates with a variant amino acid in position 26 and a reduced level of TNF-beta production. *J Exp Med.* 1991;173:209–19.
 37. Pociot F, Briant L, Jongeneel CV, Mölvig J, Worsaae H, AbbalM, Thomsen M, Nerup J, Cambon-Thomsen A: Association of tumor necrosis factor (TNF) and class II major histocompatibility complex alleles with the secretion of TNF- α and TNF- β by human mononuclear cells: A possible link to insulin dependent diabetesmellitus. *Eur J Immunol.* 1993;23:224–231.
 38. Guo W, Wang N, Li Y, Zhang JH. Polymorphisms in tumor necrosis factor genes and susceptibility to esophageal squamous cell carcinoma and gastric cardiac adenocarcinoma in a population of high incidence region of North China. *Chin Med.* 2005;J118:1870–78,
 39. Oliveira ID, Petrilli AS, Tavela MH, Zago M A, deToledo SR: TNFalpha, TNF-beta, IL-6, IL-10, PECAM-1 and the MPO inflammatory gene polymorphisms in osteosarcoma. *J Pediatr Hematol Oncol.* 1993;29:293–97.

40. Messer G, Spengler U, Jung MC, Honold G, Blomer K, Pape GR, et al., Polymorphic structure of the tumor necrosis factor (TNF) locus: An NcoI polymorphism in the first intron of the human TNF-beta gene correlates with a variant amino acid in position 26 and a reduced level of TNF-beta production. *J Exp Med.*1991;173:209-219.
41. Davies FE, Rollinson SJ, Rawstron AC, Roman E, Richards S, Drayson M, Child JA, Morgan GJ: High-produced haplotypes of tumor necrosis factor alpha and lymphotoxin alpha are associated with an increased risk of myeloma and have an improved progression-free survival after treatment. *J Clin Oncol.* 2000;18:2843-51.
42. Duell EJ, Millikan RC, Pittman GS, Winkel S, Lunn RM, Tse CK. Polymorphisms in the DNA repair gene XRCC1 and breast cancer. *Cancer Epidemiol Biomark Prev.* 2001;10:217-222.
43. Hu JJ, Smith TR, Miller M, Lohman K, Case LD. Genetic regulation of ionizing radiation sensitivity and breast cancer risk. *Environ Mol Mutagen.* 2002;39:208-215.
44. Chen S, Tang D, Xue K, Xu L, Ma G, Hsu Y, et al., DNA repair gene XRCC1 and XPD Polymorphisms and risk of lung cancer in a Chinese population. *Carcinogenesis.* 2002;23(8):1321-5.
45. Flores-Obando RE, Gollin SM, Ragin CC. Polymorphisms in DNA damage response genes and head and neck cancer risk. *Biomarkers.* 2010;15:379-399.
46. Matullo G, Guarrera S, Carturan S, et al. DNA repair gene polymorphisms, bulky DNA adducts in white blood cells and bladder cancer in a case control study, *Int. J. Cancer.* 2001;92:562-567.
47. Sturgis EM, Castillo EJ, Li L, Zheng R, Eicher SA, Clayman GL, Strom SS, Spitz MR, Wei Q. Polymorphisms of DNA repair gene XRCC1 in squamous cell carcinoma of the head and neck. *Carcinogenesis.* 1999;20(11):2125-9.
48. Goode EL, Ulrich CM, Potter JD. Polymorphisms in DNA repair genes and associations with cancer risk. *Cancer Epidemiol Biomarkers Prev.* 2002;11:1513-30.
49. Caldecott KW, Aoufouchi S, Johnson P, and Shall S. XRCC1 polypeptide interacts with DNA polymerase beta and possibly poly (ADP-ribose) polymerase, and DNA ligase III is a novel molecular 'nick-sensor' *in vitro.* *Nucleic Acids Res.* 1996;24:4387-94.
50. Skjelbred CF, Svendsen M, Haugan V, Eek AK, Clausen KO, Svendsen MV, et al. Influence of DNA repair gene polymorphisms of hOGG1, XRCC1, XRCC3, ERCC2 and the folate metabolism gene MTHFR on chromosomal aberration frequencies. *Mutat Res.* 2006;602:151-162.
51. Marintchev A, Mullen M.A, Maciejewski MW, Pan B, Gryk MR, Mullen GP. Solution structure of the single-strand break repair protein XRCC1 N-terminal domain. *Nat Struct Biol.* 1999;6:884-93.
52. Caldecott KW. XRCC1 and DNA strand break repair. *DNA Repair (Amst).* 2003;2:955-69.
53. Hitomi K, Iwai S, Tainer JA. The interact structural chemistry of base excision repair machinery: Implications for DNA damage recognition, removal and repair. *DNA Repair (Amst).* 6(4):410-28.
54. Fan J, Wilson DM. Protein-protein interactions and post translational modifications in mammalian base excision repair. *Free Radic Biol Med.* 2005;38:1121-38.
55. Loeb KR, Loeb LA. Significance of multiple mutations in cancer. *Carcinogenesis (Lond.).* 2000;21:379-85.
56. Rajewsky MF, Engelbergs J, Thomale J, Schweer T. Relevance of DNA repair to carcinogenesis and cancer therapy. *Recent Results Cancer Res.* 1998;154:127-46.
57. Millar AL, Pal T, Madlensky L, Sherman C, Temple L, Mitri A, et al. Mismatch repair gene defects contribute to the genetic basis of double primary cancers of the colorectum and endometrium. *Hum. Mol. Genet.* 1999;8:823-29.
58. Yao X, Buermeyer AB, Narayanan L, Tran D, Baker SM, Prolla et al. Different mutator phenotypes in Mlh1-versus Pms2-deficient mice. *Proc. Natl. Acad. Sci. USA.* 1999;96:6850-55.
59. Benhamou S, Sarasin A. Variability in nucleotide excision repair and cancer risk: a review. *Mutat. Res.* 2000;462:149-158.
60. Feng YZ, Liu YL, He XF, Wei W, Shen XL, Xie DL. Association between the XRCC1 Arg194Trp polymorphism and risk of cancer: Evidence from 201 case-control studies. *Tumour Biol.* 2014;35(11):10677-10697.

61. Li W, Yang F, Gui Y, Bian J. DNA repair gene XRCC1 Arg194Trp polymorphism and susceptibility to hepatocellular carcinoma: A meta-analysis. *Oncol Lett.* 2014;8(4):1725–30.
62. Shen H, Xu Y, Qian Y, Yu R, Qin Y, Zhou L, et al. Polymorphisms of the DNA repair gene XRCC1 and risk of gastric cancer in a Chinese population. *Int J Cancer.* 2000;88(4):601-6.
63. Sigurdson AJ, Land CE, Bhatti P, Pineda M, Brenner A, Carr Z, et al. Thyroid nodules, polymorphic variants in DNA repair and RET-related genes, and interaction with ionizing radiation exposure from nuclear tests in Kazakhstan. *Radiat Res.* 2009;171(1):77-88.
64. Divine KK, Gilliland FD, Crowell RE, Stidley CA, Bocklage TJ, Cook DL, et al. The XRCC1 399 glutamine allele is a risk factor for adenocarcinoma of the lung. *Mutat Res.* 2001;461(4):273-8.
65. Zhou W, Liu G, Miller DP, Thurston SW, Xu LL, Wain JC, et al. Polymorphisms in the DNA repair genes XRCC1 and ERCC2, smoking, and lung cancer risk. *Cancer Epidemiol Biomarkers Prev.* 2003;12(4):359-65.
66. Berhane N, Sobti RC, Mahdi SA. DNA repair gene polymorphism (XPG and XRCC1) and association of prostate cancer in a north Indian population. *Mol Biol Rep.* 2012;39(3):2471–79.
67. Lee SG, Kim B, Choi J, Kim C, Lee I, Song K. Genetic polymorphisms of XRCC1 and risk of gastric cancer. *Cancer Lett.* 2002;187(1-2):53-60.
68. Sturgis EM, Castillo EJ, Li L, Zheng R, Eicher SA, Clayman GL, Strom SS, Spitz MR, Wei Q. Polymorphisms of DNA repair gene XRCC1 in squamous cell carcinoma of the head and neck. *Carcinogenesis (Lond.).* 1999;20:2125–2129.
69. Abdel-Rahman SZ, El-Zein RA. The 399Gln polymorphism in the DNA repair gene XRCC1 modulates the genotoxic response induced in human lymphocytes by the tobacco-specific nitrosamine NNK. *Cancer Lett.* 2000;159:63–71.
70. Divine KK, Gilliland FD, Crowell RE, Stidley CA, Bocklage TJ, Cook DL, Belinsky SA. The XRCC1 399 glutamine allele is a risk factor for adenocarcinoma of the lung. *Mutat Res.* 2001;461:273–78.
71. Ratnasinghe D, Yao SX, Tangrea JA, Qiao YL, Andersen MR, Barrett MJ, Giffen CA, Erozan Y, Tockman MS, Taylor PR. Polymorphisms of the DNA repair gene XRCC1 and lung cancer risk. *Cancer Epidemiol Biomarkers Prev.* 2001;10:119–123.
72. Chen S, Tang D, Xue K, Xu L, Ma G, Hsu Y, Cho SS. DNA repair gene XRCC1 and XPD polymorphisms and risk of lung cancer in a Chinese population. *Carcinogenesis.* 2002;23:1321–1325.
73. Shen J, Gammon MD, Terry MB, Wang L, Wang Q, Zhang F, et al. Polymorphisms in XRCC1 modify the association between polycyclic aromatic hydrocarbon-DNA adducts, cigarette smoking, dietary antioxidants, and breast cancer risk. *Cancer Epidemiol Biomarkers Prev.* 2005;14:336–342.
74. Momand J, Zambetti GP, Olson DC, George D, Levine AJ. The mdm-2 oncogene product forms a complex with the p53 protein and inhibits p53-mediated transactivation. *Cell.* 1992;69:1237-45.
75. Tavernarakis N, Pasparaki A, Tasedemir E, Maiuri MC, Kroemer G. The effects of p53 on whole organism longevity are mediated by autophagy. *Autophagy.* 2008;4(7):870- 3.
76. Shieh SY, Ikeda M, Taya Y and Prives C. *Cell.* 1997;91:325–34.
77. Waterman MJF, Stavridi ES, Waterman JLF, Halazonetis TD. ATMdependent activation of p53 involves dephosphorylation and association with 14-3-3 proteins. *Nature Genet.* 1998;19: 175- 8.
78. Foord OS, Bhattacharya P, Reich Z, Rotter V. A DNA binding domain IS contained in the C-terminus of wild type p53 protein. *Nucleic Acids Res.*1991;19:5191-8.
79. Hermeking H, Lengauer C, Polyak K, He TC, Zhang L, Thiagalingam S, Kinzler KW, Vogelstein B. *Molec. Cell.*1997;1:3–11.
80. Itahana K, Dimri G, Campisi J. *Eur. J. Biochem.* 2001;268:2784–91.
81. Mizuaray S, Yamanaka K, Kotani H. Mutant p53 induces the GEF-H1 oncogene, a guanidine nucleotide exchange factor H1 for RhoA, resulting in accelerated cell proliferation n tumor cells. *Cancer Res.* 2006;66:6319–26.
82. Ginsberg D, Mechta F, Yaniv M, Oren M. Wild-type p53 can down-modulate the activity of various promoters. *Proc. Natl. Acad. Sci. USA.*1991;88:9979-9983.
83. DeLeo AB, Jay G, et al. Detection of a transformation-related antigen in chemically induced sarcomas and other

- transformed cells of the mouse. Proceedings of the National Academy of Science, USA. 1979;76:2420-24.
84. Finlay CA, Hinds PW, Levine AJ. The p53 proto-oncogene can act as a suppressor of transformation. Cell. 1989;57:1083-93.
 85. Michalovitz D, Halevy O, Oren M. Conditional inhibition of transformation and of cell proliferation by a temperature-sensitive mutant of p53. Cell. 1990;62(4): 671-80.
 86. Chen PL, Chen YM, Bookstein R, Lee WH. Genetic mechanisms of tumor suppression by the human p53 gene. Science. 1990; 250(4987):1576-80.
 87. Mowat M, Cheng A, Kimura N, Bernstein A, Benchimol S. Rearrangements of the cellular p53 gene in erythroleukaemic cells transformed by Friend virus. Nature. 1985; 314:633-636.
 88. Baker SJ, Fearon ER, Nigro JM, Hamilton SR, Preisinger AC, Jessup JM, vanTuinen P, Ledbetter DH, Barker DF, Nakamura Y, et al. Chromosome 17 deletions and p53 gene mutations in colorectal carcinomas. Science. 1989;244(4901):217-21.
 89. Gannon JV, Lane DP. p53 and DNA Polymerase alpha compete for binding to SV40 T antigen. Nature. 1987;329:456-58.
 90. Mercer WE, Shields MT, Lin D, Appella E, Ullrich SJ. Growth suppression induced by wild-type p53 protein is accompanied by selective down-regulation of proliferating-cell nuclear antigen expression. Proc Natl Acad Sci USA. 1991;88(5):1958-62.
 91. Hollstein M, Rice K, Greenblatt MS. Database of p53 gene somatic mutations in human tumors and cell lines. Nucleic Acids Res. 1994;22(17):3551-5.
 92. Vogelstein B, Lane D, Levine AJ. Surfing the p53 network. Nature. 2000;408:307-10.
 93. Kubbutat M, Vousden KH. Keeping an old friend under control: regulation of p53 stability. Mol. Med. Today. 1998;4:250-6.
 94. Haupt Y, Maya R, et al. Mdm2 promotes the rapid degradation of p53. Nature. 1997;387:296-99.
 95. Vousden KH and Lu X. Cancer. Nat. Rev. 2002;2:594-604.
 96. Soussi T. p53 Antibodies in the sera of patients with various types of cancer: A review. Cancer Res. 2000;60:1777-88.
 97. Wang XW, Yeh H, Schaeffer L, Roy R, Moncollin V, Egly JM, Wang Z, Freidberg EC, Evans MK, Taffe BG, Bohr VA, Weeda G, Hoeijmakers JH, Forrester K, Harris CC. Nat. Genet. 1995;10:188-195.
 98. Marin MC, Jost CA, Brooks LA, Irwin MS, O'Nions J, Tidy JA, James N, McGregor JM, Harwood CA, Yulug IG, Vousden KH, Allday MJ, Gusterson B, Ikawa S, Hinds PW, Crook T, Kaelln WG., Jr A common polymorphism acts as an intragenic modifier of mutant p53 behaviour. Nat Genet. 2000;25:47-54.
 99. Schneider-Stock R, Mawrin C, Motsch C, Boltze C, Peters B, Hartig R, Buhtz P, Giers A, Rohrbeck A, Freigang B, Roessner A. Retention of the arginine allele in codon 72 of the p53 gene correlates with poor apoptosis in head and neck cancer. Am J Pathol. 2004;164(4): 1233-41.
 100. Nelson HH, Wilkojmen M, Marsit CJ, Kelsey KT. TP53 mutation, allelism and survival in non-small cell lung cancer. Carcinogenesis. 2005;26:1770-3.
 101. Wu W, Tang X, Hu W, Lotan R, Hong W, Mao L. Identification and validation of metastasis-associated proteins in head and neck cancer cell lines by two-dimensional electrophoresis and mass spectrometry. Clin Exp Metastasis. 2002; 19:319-26.
 102. Fan R, Wu MT, Miller D, Wain JC, Kelsey KT, Wiencke JK, et al. The p53 codon 72 polymorphism and lung cancer risk. Cancer Epidemiol Biomarkers Prev. 2000;9:1037-42.
 103. Weston A, Pan CF, Ksieski HB, Wallenstein S, Berkowitz GS, Tartter PI, et al. p53 haplotype determination in breast cancer. Cancer Epidemiol Biomarkers Prev. 1997;6:105-12.
 104. Buyru N, Tigli H, Dalay N. P53 codon 72 polymorphism in breast cancer. Oncol Rep. 2003;10:711-4.
 105. Klug SJ, Wilmotte R, Santos C, Almonte M, Herrero R, Guerrero I, et al. TP53 polymorphism, HPV infection, and risk of cervical cancer. Cancer Epidemiol Biomarkers Prev. 2001;10:1009-12.
 106. Runnebaum IB, Tong XW, Konig R, Zhao H, Korner K, Atkinson EN, et al. p53-based blood test for p53PIN3 and risk for sporadic ovarian cancer. Lancet. 1995; 345:994.
 107. Wang-Gohrke S, Weikel W, Risch H, Vesprini D, Abrahamson J, Lerman C, et al. Intron variants of the p53 gene are associated with increased risk for ovarian cancer but not in carriers of BRCA1 or

- BRCA2 germline mutations. Br J Cancer. 1999;81:179–83.
108. Costa S, Pinto D, Pereira D, Rodrigues H, Cameselle-Teijeiro J, Medeiros R, et al. Importance of TP53 codon 72 and intron 3 duplication 16bp polymorphisms in prediction of susceptibility on breast cancer. BMC Cancer. 2008;8:32.
109. Campbell IG, Eccles DM, Dunn B, Davis M, Leake V. p53 polymorphism in ovarian and breast cancer. Lancet. 1996;347:393–4.
110. Kim JM, Lee OY, Lee CG, Kwon SJ, Kim KS, Moon W, et al., p53 Codon 72 and 16-bp duplication polymorphisms of gastric cancer in Koreans] Korean J Gastroenterol. 2007;50:292–8.
111. Lancaster JM, Brownlee HA, Wiseman RW, Taylor J. p53 polymorphism in ovarian and bladder cancer. Lancet. 1995;346:182.
112. Peller S, Kopilova Y, Slutzki S, Halevy A, Kvitko K, Rotter V. A novel polymorphism in intron 6 of the human p53 gene: A possible association with cancer predisposition and susceptibility. DNA Cell Biol. 1995;14:983–90.
113. Lehman TA, Haffty BG, Carbone CJ, Bishop LR, Gumbs AA, Krishnan S, et al. Elevated frequency and functional activity of a specific germ-line p53 intron mutation in familial breast cancer. Cancer Res. 2000;60:1062–9.
114. Fiszer-Maliszewska L, Kazanowska B, Kusnierczyk P, Manczak M, Niepieklo W, Pochron-Zeman B, et al. Is p53 intronic variant G13964C associated with predisposition to cancer? J Appl Genet. 2003;44:547–52.
115. Hainaut P, Pfeifer GP. Carcinogenesis. 2001;22:367–74.
116. Moll UM, LaQuaglia M, Benard J, Riou G. Wild-type p53 protein undergoes cytoplasmic sequestration in undifferentiated neuroblastomas but not in differentiated tumors. Proc. Natl. Acad. Sci. USA. 1995;92:4407–11.
117. Yamamoto H, Ozaki T, Nakanishi M, Kikuchi H, Yoshida K, Horie H, et al. Oxidative stress induces p53- dependent apoptosis in hepatoblastoma cell through its nuclear translocation. Genes Cells. 2007;12:461–71.

© 2018 Kollabathina and Sudhakar; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

*The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history/23592>*