

*Full Length Research Paper*

# Identification and mapping QTLs of bolting time in purple cai-tai (*Brassica rapa* L. var. *purpurea*)

Xiao-Hui Deng<sup>1,2\*</sup>, Qi-Jun Nie<sup>1,2</sup>, Zheng-meng Qiu<sup>1,2</sup>, Cai-xia Gan<sup>1,2</sup>, Lei Cui<sup>1,2</sup> and Feng-Juan Zhu<sup>1,2</sup>

<sup>1</sup>Institute of Economic Crops, Hubei Academy of Agricultural Science, Wuhan 430064, People's Republic of China.

<sup>2</sup>Hubei Key Laboratory of Vegetable Germplasm Enhancement and Genetic Improvement, Wuhan 430064, People's Republic of China.

Received 26 June, 2019; Accepted 15 January, 2020

**Bolting time is a crucial agronomic trait for yield and quality in purple cai-tai (*Brassica rapa* L. var. *purpurea*), but the genetic mechanism controlling the procedure remains unknown. In the present study, a double haploid (DH) population derived from two inbred lines of purple cai-tai 4-1 and 040-3 was constructed to identify the quantitative trait loci (QTLs) of bolting time. Genetic linkage map was performed by JoinMap version 3.0 using SSR, SRAP and ESTP molecular markers. A total of one hundred and thirty-eight molecular markers were integrated into ten linkage groups (LGs), which were anchored to the corresponding chromosome of the *B. rapa* reference genome. The genetic linkage map covers 1253.1 cM, with an average distance of 9.08 cM between two adjacent markers. Five quantitative trait loci (QTLs) were identified to control bolting time and explaining variations from 17.7 to 44.2%. The genetic results of bolting time will be useful for future breeding of late bolting in purple cai-tai.**

**Key words:** Purple cai-tai (*Brassica rapa* L. var. *purpurea*), bolting time, quantitative trait loci (QTL).

## INTRODUCTION

*Brassica rapa* is consisted of various vegetables such as Chinese cabbage, non-heading Chinese cabbage, and turnip. Non-heading Chinese cabbage includes economically important vegetable taxa with a wide range of morphologies, such as pakchoi (*Brassica campestris* ssp. *chinensis* Makino var. *communis* Tsen et Lee), purple cai-tai (*B. rapa* L. var. *purpurea*, Canjie et al., 2019), rosette bok choy (*B. campestris* ssp. *chinensis* Makino var. *rosularis* Tsen et Lee), and taicai (*B. campestris* ssp. *chinensis* Makino var. *tai-tsai* Hort). Purple cai-tai is a natural early bolting mutant which bolting earlier without vernalization, and it is an important vegetable in the middle and lower reaches of Yangtze

river.

In *B. rapa*, one of the most important agronomic traits is bolting because premature bolting significantly affects the quality and yield of the economic products (Kitamoto et al., 2014). Bolting times are regulated by multiple genes. In *Arabidopsis thaliana*, over 300 regulatory genes for bolting and flowering time have been isolated (Bouché et al., 2016). Many QTLs of bolting and flowering have been characterized in *B. rapa*. In the past two decades (Nishioka et al., 2005; Lou et al., 2007, 2011; Li et al., 2009; Kakizaki et al., 2011; Li et al., 2015).

During the elucidation of the genomes of crop species, it is crucial to assign molecular markers to the linkage

\*Corresponding author. E-mail: 954266577@qq.com.

groups (LGs) and construct genetic maps. A number of genetic linkage maps have been produced for *B. rapa* based on diverse marker types including Restriction Fragment Length Polymorphisms (RFLPs), Random Amplified Polymorphic DNA (RAPD), Simple sequence Repeats (SSR), Amplified Fragment Length Polymorphisms (AFLPs), and Sequence-related amplified polymorphism (SRAP) (Kim et al., 2006; Suwabe et al., 2006; Soengas et al., 2007; Yan et al., 2009; Honghao et al., 2014; Haidong et al., 2016).

SRAP has some advantages, such as simple, a apposite throughput rate, targeting open-reading frames (ORFs), and so on (Uzun et al., 2009). SSR markers are useful to construct high-density maps because of its high polymorphism levels, its co-dominant character, its abundance and wide distribution during the genome and the utility as convenient anchor points in the integration of intraspecific and interspecific consensus maps (Acher et al., 2004). Expressed sequence tag polymorphism (ESTP) markers are transferable between species and between genera (Brown et al., 2001).

Although many QTLs about bolting have been isolated and characterized with molecular markers, the report of QTL and the markers based on sequence-tagged Polymerase chain reaction (PCR) mapped in purple cai-tai is limited (Canjie et al., 2019), especially those which may provide anchors to the genome of *B. rapa* and are readily transferable to other populations. Thus, the objective of this research was to identify QTLs controlling bolting in two years. Our results should be useful to understand the genetic mechanism about the bolting in purple cai-tai, and contribute to breeders for designing effective strategies for better cultivar.

## MATERIALS AND METHODS

### Plant materials and DNA isolation

Double haploid (DH) population consists of 140 individual DH lines was employed for trait assay and genetic mapping. The population was developed from microspore culture of F<sub>1</sub> buds of the cross between 040-3, a cultivar with early bolting which was derived from 040 and 4-1, a high inbred line with late bolting, which was obtained by seven generations of self-pollination of cultivar Daguzi. The plants of parents, F<sub>1</sub> and 140 individual DH lines were cultivated in an open field at the Institute of Economic Crops of Hubei Academy of Agricultural Science, Wuhan, China (30.57°N, 114.3°E) from September of 2012 to April of 2013, and September of 2013 to April of 2014. The bolting time (that is, days after sowing to appearance of macroscopic floral bud) was judged by the observation recorded every third day (Wang et al., 2018) in 2013, 2014 spring.

### Detection of DNA polymorphism

DNA was isolated from fresh and young leaves of the parental and 140 DH lines according to the protocol published by Guillemaut and Laurence (1992). 106 SSR markers, and 4 ESTP markers and 652 SRAP markers were used to filtrate the polymorphism of the two parents and F<sub>1</sub>. The experiment of SRAP was carried out following the procedure reported by Li and Quiros (2001), with minor modifications. SSR and ESTP markers were obtained as described

by Choi et al. (2007) and HyeRan et al. (2009) (Supplementary Table 1). PCR was performed in a 10 µl reaction mixture containing 2 µl DNA template (40 ng), 1 µl 10 × PCR buffer (MgCl<sub>2</sub>), 0.2 µl forward primer (10 µM), 0.2 µl reverse primer (10 µM), 0.8 µl dNTPs (10 mM), 0.2 µl TaqDNA polymerase (2.0 U/µl), and 5.6 µl ddH<sub>2</sub>O (Biomed Tec Co., Beijing, China). PCR conditions were as the follows: an initial denaturation step at 94°C for 4 min, followed by 35 cycles of DNA amplification (94°C for 30 s, 60°C for 30 s, and 72°C for 60 s), with a final 7 min extension at 72°C (Mastercycler nexus, Eppendorf, German). The PCR products were separated by electrophoresis on 9% polyacrylamide gels (acrylamide/bisacrylamide = 29:1) and screened with silver staining (Choi et al. 2007).

### Linkage analysis, map construction and QTL analysis

A scoring system was applied for the reproducibly polymorphic makers among the parent lines in the DH population. Linkage assay and the construction of maps were carried out by JoinMap Version3.0 (Stam, 1993; Van Ooijen and Voorrips, 2001). SSR and ESTP markers previously mapped (Yan et al., 2011) were utilized for the identification of LGs in the LOD groups with a threshold range of 3.0–8.0. The annotation of LGs was identical with the second generation of referenced LGs in *B. rapa* (A1–A10). A composite interval mapping (CIM) reported by Zeng (1994) was employed for the analysis of QTLs for bolting time by a QTL Cartographer (version 2.5) (Basten et al., 2002). In order to estimate the appropriate significance threshold of a logarithm of odds (LOD), a test of 1,000-permutation was carried out via the QTL Cartographer.

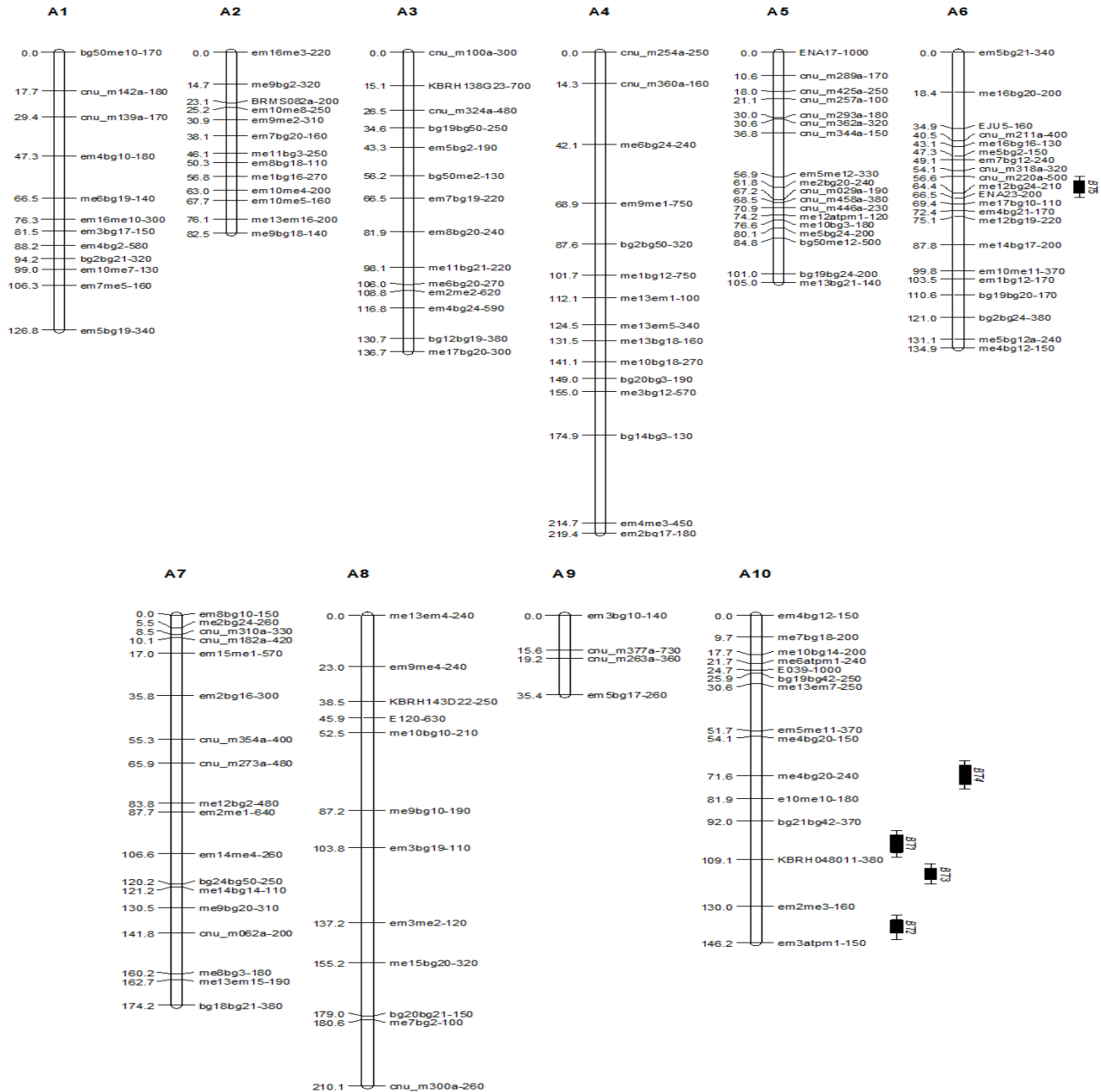
## RESULTS

### Polymorphism screening of primers between parents

In order to construct the genetic linkage map, the two parents and F<sub>1</sub> were filtrated for polymorphism with 652 SRAP markers and 106 SSR markers, and 4 ESTP markers. In total, 183 (24.02%) out of 762 primers (or primer combinations, abbreviated as PCs), including 128 SRAP PCs, 42 SSR PCs, and 3 ESTP, produced polymorphic loci. A total of 129 polymorphic loci were selected with the help of 128 polymorphic SRAP primer combinations. Meanwhile, 42 SSR and 3 ESTP polymorphic loci were obtained. All these obtained polymorphic markers were employed for the assay of DH population (Supplementary Table 2).

### Construction of genetic linkage map

A total of 140 DH individuals from F<sub>1</sub> progenies of two purple cai-tai “4-1” and “040-3” were used for genotyping and linkage analysis. There were 25.0% of 184 polymorphic markers not assigned. As shown in Figure 1, a total of 138 markers were anchored to 10 LGs which spanned 1253.1 cM of map distance with an average distance of 9.08 cM. The location of 10 LGs on their corresponding chromosomes (A1-A10) was confirmed via 32 SSR and 2 ESTP markers of which the map positions were already known on the reference maps of *B. rapa*.



**Figure 1.** Genetic linkage map and localization of QTLs of bolting time traits on a population of 140 DH lines of purple cai-tai.

The length range of individual LGs varied from 35.4 cM (A9) to 219.4 cM (A4).

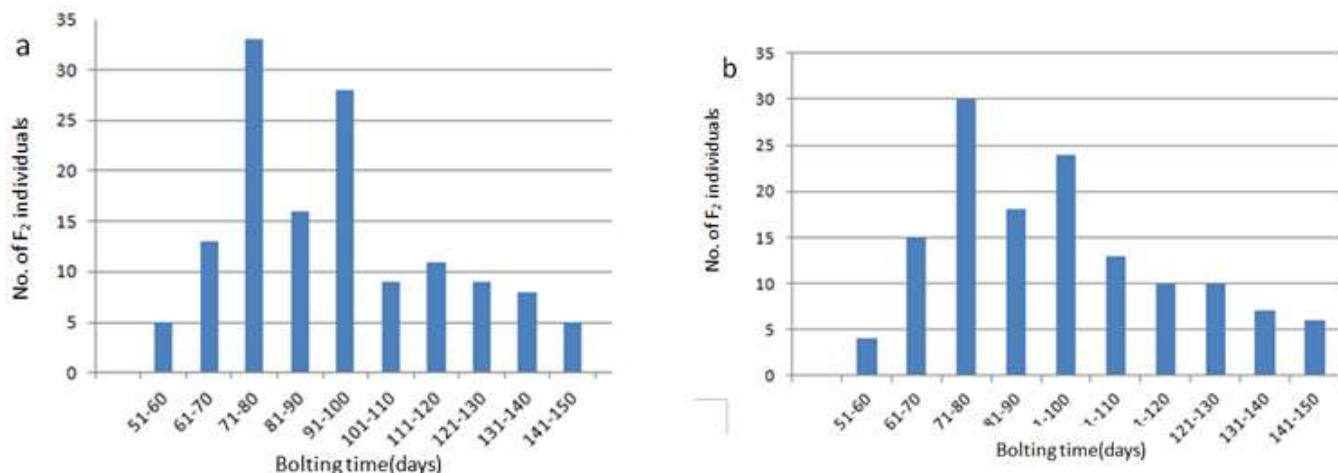
### QTL analysis for bolting time

As shown in Table 1, the parental lines, 040-3 and 4-1, revealed significant difference of the bolting time. All seven plants of 040-3 exhibited stably bolting time at 40 DAS in 2013 spring and 41 DAS in 2014 spring, respectively. For 4-1 of late bolting parent line, bolting

time was detected between narrow ranges from 127 to 129 DAS in 2013 spring, and 128 to 132 DAS in 2014 spring, respectively. These results indicated that the genetic background of these two purple cai-tai lines is nearly homozygous with little environmental effect. Further, the average bolting time of the  $F_1$  showed  $96 \pm 2.2$ ,  $97 \pm 3.2$  DAS which is slightly larger than that of the mid-parent ( $84 \pm 1$  DAS,  $85.5 \pm 1.4$  DAS) in 2013 and 2014 spring, respectively. The bolting time of 137 out of the 140  $F_2$  DH progenies were checked that revealed a continuous distribution from 57 to 141 DAS in 2013

**Table 1.** Variation in bolting and flowering time of parents and F2 population.

Environment	Generations	No. of plants	Bolting time (days)
Spring-2013	040-3	7	40
	4-1	10	128±1
	F1	10	96±2.4
	Mean of F2 population	137	94.7±3.5
	Range of F2 population	137	57-141
Spring-2014	040-3	10	41
	4-1	10	130±1.4
	F1	10	97±3.2
	Mean of F2 population	137	93.1±2.8
	Range of F2 population	137	55-140

**Figure 2.** Frequency distribution of bolting time in the F<sub>2</sub> population. (a) 2013 spring, (b) 2014 spring.

spring, and from 55 to 140 DAS in 2014 spring. The other 3 of the 140 F<sub>2</sub> DH progenies died before bolting. These results suggested that the bolting time in purple cai-tai probably be controlled by quantitative trait locus.

The frequency distributions of bolting time in the F<sub>2</sub> populations revealed continuous distribution, also showing that bolting time are quantitative traits controlled by polygenes (Figure 2). QTL analysis was performed individually for each of the 2013 and 2014 tests. Five QTLs for bolting were detected in A6 and A10 (four regions). The largest QTL effect (LOD of 11.73) on bolting time, named as BT1, was detected between the loci KBRH048O11-380 and bg21bg42-370 on A10, which explained approximately 44.2% phenotypic variation. Other four QTLs, named as BT2, BT3, BT4 and BT5, were mapped in A10 and A6 chromosome explaining 42.7, 41.6, 34.2, and 17.7% phenotypic variation, respectively. Remarkably, BT1, BT2, BT4 were detected twice in 2013 and 2014, but BT3 only in 2013 and BT5 only in 2014 (Table 2).

## DISCUSSION

A genetic linkage map was constructed via a segregating population of 140 purple cai-tai DH lines. This linkage map contains 104 SRAP, 32 SSR, and 2 ESTP markers which were grouped on 10 LGs, and each LG was anchored to the corresponding chromosome of the *B. rapa* reference map based on the common SSR and ESTP (Yan et al., 2011; HyeRan et al., 2009; Su Ryun Choi et al., 2007). It indicates that this map can be integrated into other genetic linkage map of *B. rapa* and be useful for other researchers. Covered with a total genetic distance of 1253.1 cM, the linkage map in the present study is comparable to the published sequenced BAC anchored reference genetic map which is 1,123.3 cM (HyeRan et al., 2009) and the sequence-based genetic linkage map which is 1234.2 cM illustrated by Yan et al. (2011). The genetic map lengths differences among various reports are attributed to the scoring errors for the most parts. In addition, the differences have also

**Table 2.** QTL detected for the bolting time traits based on CIM (composite interval mapping) analysis.

QTLs	Years	Marker interval	Group	QTL position	Peak LOD <sup>a</sup>	Addition effect
BT1	2013	bg21bg42-370--KBRH048011-380	A10	102.049	11.73	15.42
BT1	2014	bg21bg42-370--KBRH048011-380	A10	102.049	11.24	14.53
BT2	2013	em2me3-160--em3atpm1-150	A10	138.992	10.77	15.69
BT2	2014	em2me3-160--em3atpm1-150	A10	138.992	10.54	15.23
BT3	2013	KBRH048011-380--em2me3-160	A10	116.12	11.06	15.24
BT4	2013	me4bg20-150--e10me10-180	A10	72.605	10.64	13.82
BT4	2014	me4bg20-150--e10me10-180	A10	72.605	9.93	12.51
BT5	2014	cnu_m220a-500--ENA23-200	A6	61.628	5.0	10.12

been reported to be caused by the type of markers, number of individuals, number of markers, recombination frequency, LOD values, and the software employed (Gosselin et al., 2002). The density of marks in the linkage map in the present study is more lower than the maps of Xiaowu et al. (2011) and Lei et al. (2018), so it needs to add marks to this linkage map for further research.

In total, five QTL affecting bolting time were identified in this study. The QTL BT5 near the marker cnu\_m220a in A6 is similar with the qFT6.1 in *B. rapa* L. (Yating et al., 2016), it is a new QTL or the same QTL, need further verification. There is no similarity of the other four QTLs BT1, BT2, BT3 and BT4 with the previous studies (Jonathan et al., 1995; Hidetoshi et al., 2001; Yating et al., 2016), they may be new QTLs and subject to be further verify. The number of loci influencing bolting is different with previous genetic analyses (Jonathan H et al., 1995; Hidetoshi et al., 2001), that mostly attributable to different population, number of individuals, number of markers, and so on. In the future, a common linkage map will be employed to comparatively assay these QTLs for the elucidation of the genetics of bolting in *Brassica* crops. Moreover, it might make a contribution to the breeding of novel cultivars with controlled bolting.

## CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

## ACKNOWLEDGMENTS

The authors gratefully appreciate the financial support provided by the National Key Research and Development Program of China (2017YFD0101806), the Technology Innovation Project of Hubei Province (2016ABA095 (Project code: 31100876)).

## REFERENCES

Acher V, Favre-Rampant P, Jeandroz S, Besnard G, Markussen T,

- Aragones A, Fladung M, Ritter E, Favre JM (2004). A full saturated linkage map of *Picea abies* including AFLP, SSR, ESTP, 5S rDNA and morphological markers. *Theoretical and Applied Genetics* 108:1602-1613.
- Brown GR, Kadel EE, Bassoni DL, Kiehne KL, Temesgen B, van Buijtenen JP, Sewell MM, Marshall KA, Neale DB (2001). Anchored reference loci in Loblolly pine (*Pinus taeda* L.) for integrating pine genomics. *Genetics* 159:799-809.
- Bouché F, Lobet G, Tocquin P, Périlleux C (2016). FLOR-ID: an interactive database of flowering-time gene networks in *Arabidopsis thaliana*. *Nucleic Acids Research* 44(Database issue):D1167-D1171
- Canjie W, Honglian Li, Yixiao Li, Qiufeng Meng, Fei Xie, Yuejin Xu, Zhengjie Wan (2019). Genetic characterization and fine mapping BrCER4 in involved cuticular wax formation in purple cai-tai (*Brassica rapa* L. var. *purpurea*). *Molecular Breeding* 39:12.
- Choi SR, Teakle GR, Plaha P, Kim JH, Allender CJ, Beynon E, Piao ZY, Soengas P, Han TH, King GJ, Barker GC, Hand P, Lydiate DJ, Batley J, Edwards D, Koo DH, Bang JW, Park BS, Lim YP (2007). The reference genetic linkage map for the multinational *Brassica rapa* genome sequencing project. *Theoretical and Applied Genetics* 115:777-792.
- Gosselin I, Zhou Y, Bousquet J, Isabel N (2002). Megagametophyte-derived linkage maps of white spruce (*Picea glauca*) based on RAPD, SCAR and ESTP markers. *Theoretical and Applied Genetics* 104:987-997.
- Guillemaut P, Laurence MD (1992). Isolation of plant DNA: a fast, inexpensive and reliable method. *Plant Molecular Biology Reporter* 10:60-65.
- Haidong L, Dezhi Du, Shaomin G, Lu X, Zhigang Z, Zhi Z, Xiaorong X, Guoyong T, Liang X, Zhong F, Yanmei Y, Robert WD (2016). QTL analysis and the development of closely linked markers for days to flowering in spring oilseed rape (*Brassica napus* L.). *Molecular Breeding* 36:52.
- Hidetoshi A, Yasuhisa K, Susumu Y, Suelo E, Masashi Hirai (2001). Identification and mapping of a quantitative trait locus controlling extreme bolting in Chinese cabbage (*Brassica rapa* L. ssp. *Pekinensis* syn. *campestris* L.) using bulked segregant analysis. *Euphytica* 118:75-81.
- Honghao L, Qingbiao W, Yangyong Z, Limei Y, Zhiyuan F, Xiaowu W, Yumei L, Mu Z, Yan L, Hailong Y, Bo L (2014). Linkage map construction using In Del and SSR markers and QTL analysis of heading traits in *Brassica oleracea* var. *capitata* L. *Molecular Breeding* 34:87-98.
- HyeRan K, Su Ryun C, Jina B, Chang PH, Seo YL, Md Jamil H, Dan VN, Mina J, Beom SP, Jea-Wook B, Ian B, Yong PL (2009). Sequenced BAC anchored reference genetic map that reconciles the ten individual chromosomes of *Brassica rapa*. *BMC Genomics* 10:432.
- Jonathan HC, Richard M, James KMB, Caroline D (1995). QTL analysis of flowering time in *Arabidopsis thaliana*. *Molecular and*

- General Genetics 248:278-286.
- Kakizaki T, Kato T, Fukino N, Ishida M, Hatakeyama K, Matsumoto S (2011). Identification of quantitative trait loci controlling late bolting in Chinese cabbage (*Brassica rapa* L.) parental line Nou 6 gou. *Breeding Science* 61:151-159.
- Kim JS, Chung TY, King GJ, Jin M, Yang T, Jin Y, Kim H, Park B (2006). A sequence-tagged linkage map of *Brassica rapa*. *Genetics* 174:29-39.
- Kitamoto N, Yui S, Nishikawa K, Takahata Y, Yokoi S (2014). A naturally occurring long insertion in the first intron in the *Brassica rapa* FLC2 gene causes delayed bolting. *Euphytica* 196:213-223.
- Lei Zhang, Xu C, Jian Wu, Min L, Stefan G, Feng C, Jianli L, Chengcheng C, Zhiyuan L, Bo L, Fan W, Song L, Fuyan L, Xuming L, Lin C, Wencai Y, Mai-he L, Ueli G, Hongkun Z, Xiaowu W (2018). Improved *Brassica rapa* reference genome by single-molecule sequencing and chromosome conformation capture technologies. *Horticulture Research* 5:50.
- Li F, Kitashiba H, Inaba K, Nishio T (2009). A *Brassica rapa* linkage map of EST-based SNP markers for identification of candidate genes controlling flowering time and leaf morphological traits. *DNA Research* 16:311-323.
- Li G, Quiros CF (2001). Sequence-related ampliWed polymorphism (SRAP), a new marker system based on a simple PCR reaction: its application to mapping and gene tagging in Brassica. *Theoretical and applied genetics* 103:455-461.
- Li XN, Wang WK, Wang Z, Li KN, Lim YP, Piao ZY (2015). Construction of chromosome segment substitution lines enables QTL mapping for flowering and morphological traits in *Brassica rapa*. *Frontiers of Plant Science* 6:432.
- Lou P, Zhao JJ, Kim JS, Shen S, Del Carpio DP, Song X, Jin M, Vreugdenhil D, Wang X, Koornneef M, Bonnema G (2007). Quantitative trait loci for flowering time and morphological traits in multiple populations of *Brassica rapa*. *Journal of Experimental Botany* 58:4005-4016.
- Lou P, Xie Q, Xu X, Edwards CE, Brock MT, Weinig C, McClung CR (2011). Genetic architecture of the circadian clock and flowering time in *Brassica rapa*. *Theoretical and applied genetics* 123:397-409.
- Matsumoto S (2006). Simple sequence repeat-based comparative genomics between *Brassica rapa* and *Arabidopsis thaliana*: The genetic origin of clubroot resistance. *Genetics* 173:309-319.
- Nishioka M, Tamura K, Hayashi M, Fujimori Y, Ohkawa Y, Kuginuki Y, Harada K (2005). Mapping of QTL for bolting time in *Brassica rapa* (*syn.campestris*) under different environmental conditions. *Breeding Science* 55:127-133.
- Soengas P, Hand P, Vicente JG, Pole JM, Pink DAC (2007). Identification of quantitative trait loci for resistance to *Xanthomonas campestris* pv. *Campestris* in *Brassica rapa*. *Theoretical and applied genetics* 114:637-645.
- Stam P (1993). Construction of integrated genetic linkage maps by means of a new computer package: Join Map. *Plant Journal* 3:739-744.
- Su Ryon C, Graham RT, Prikshit P, Jeong HK, Charlotte J, Allender EB, Zhong YP, Pilar S, Tae HH, Graham J, King GC, Barker PH, Derek J, Lydiate JB, David E, Dal HK, Jae WB, Beom-Seok P, Yong PL (2007). The reference genetic linkage map for the multinational *Brassica rapa* genome sequencing project. *Theoretical and applied genetics* 115:777-792.
- Suwabe K, Tsukazaki H, Iketani H, Hatakeyama K, Kondo M, Fujimura M, Nunome T, Fukuoka H, Hirai M, Van Ooijen JW, Voorrips RE (2001). JoinMap ® Version 3.0: software for the calculation of genetic linkage maps. CPRO-DLO, Wageningen.
- Uzun A, Yesiloglu T, Aka-Kacar Y, Tuzcu O (2009). Genetic diversity and relationships within Citrus and related genera based on sequence related amplified polymorphism markers (SRAPs). *Science Horticulture* 121:306-312.
- Xiaowu W, Hanzhong W, Jun W, Rifei S, Jian W, Shengyi L, Yinqi B, Jeong-Hwan, Mun IB, Feng C, Sanwen H, Xixiang L, Wei H, Junyi W, Xiyin W, Michael F, Chris P, Andrew H, Paterson BC, Bo W, Alice H, Andrew GS, Beom-Seok P, Bernd W, Binghang L, Bo L, Bo L, Chaobo T, Chi S, Christopher D, Chunfang P, Chunyu G, Chushin K, Chuyu L, David E, Desheng M, Di S, Eleni S, Fei L, Fiona F, Gavin C, Gilles L, Graham JK, Guusje B, Haibao T, Haiping W, Harry B, Helling Z, Hideki H, Hiroshi A, Hui G, Hui W, Huizhe J, Isobel AP, Jacqueline B, Jeong-Sun K, Jérémy J, Jianwen L, Jiaohui X, Jie D, Jin AK, Jingping L, Jingyin Y, Jinling M, Jinpeng W, Jiumeng M, Julie P, Jun W, Katsunori H, Kui W, Li W, Lu F, Martin T, Matthew GL, Meixia Z, Mina J, Nirala R, Nizar D, Paul JB, Qingle C, Quanfei H, Ruiqiang L, Satoshi T, Shifeng C, Shu Z, Shujiang Z, Shunmou H, Shusei S, Silong S, Soo-Jin K, Su-Ryun C, Tae-Ho L, Wei F, Xiang Z, Xu T, Xun X, Yan W, Yang Q, Ye Y, Yingrui L, Yongchen D, Yongcui L, Yongpyo L, Yoshihiro N, Yupeng W, Zhenyi W, Zhenyu L, Zhiwen W, Zhiyong X, Zhonghua Z (2011). The genome of the mesopolyploid crop species *Brassica rapa*. *Nature genetics* 43(10):1035-1039.
- Yan C, Jianfeng G, Jingyi Z, Qian W, Qingyu B, Xilin H (2009). The construction of a genetic linkage map of non-heading Chinese cabbage (*B. campestris* ssp. *chinensis* Makino). *Journal of Genetics and Genomics* 36:501-508.
- Yan W, Silong S, Bo L, Hui W, Jie D, Yongcui L, Qian W, Feng C, Xiaowu W, Jian W (2011). A sequence-based genetic linkage map as a reference for *Brassica rapa* pseudochromosome assembly. *BMC Genomics* 12:239.
- Yating L, Chengyu L, Xingxing S, Hui F, Yugang W (2016). Identification of QTLs with additive, epistatic, and QTL 3 environment interaction effects for the bolting trait in *Brassica rapa* L. *Euphytica* 210:427-439.
- Wang QB, Zhang YJ, Zhang L (2018). A naturally occurring insertion in the RsFLC2 gene associated with late-bolting trait in radish (*Raphanus sativus* L.). *Molecular Breeding* 38:137.

**Supplementary Table 1.** Primers for SSR, ESTP and SRAP marker assays.

Marker name	assay	Assay type	Forward primer	Reverse primer
KBRH138G23		SSR	TTTGACATCGTGCAATGCTA	TTGGGCTGGTCCTGAAGATA
KBRH139B23		SSR	ATTCATGGTTGGTTCACCG	ATTTCCAAAACACACACGCA
KBRH143D22		SSR	GATGTGATACTTTGGCGACGG	TGAAGGATAATATGGTCTTGCC
KBRH143F19		SSR	GCATGCAAGCTTGGAACTGAT	CAGTCACGCTTTCTGACGAAAA
KBRH143H15		SSR	TCTGCATCAAAATGCTAAAATGA	TGATCTTTTAGAAACAAAGATCGAG
KBRH143K20		SSR	CAAATGTCTCAAGACACATAAACCA	CTAAAGCAGCAATTGGGTGTTC
KBRH048O11		SSR	GCCTCTACCTGGCTTCAGCA	TCATTTGGCGCATACTTCCA
EJU3		SSR	CCTCTTTAATTCAAACAAGAAATCA	TTCGGACAATGGCAGTGATA
EJU5		SSR	GGCACGTACATGGAGGATTC	TGTTGGTCGAGCTGTTTCAG
ENA17		SSR	CAGTTATTTTCGCCTCGTCT	TATTTGTGGTCTGTTATTGGA
ENA18		SSR	TAAAATGAAACCCACCCGA	TGTTGGGCAACATCCATTTA
ENA23		SSR	GCTGTGCCAGTTCCTCTTTC	TCATTCCAAATGGCCTTACC
ENA28		SSR	GGAGTCCGAGCGTTATGAAT	CTTCATCGACCCACCTTGTT
ENA4		SSR	ACTTCTCTTTATTCACTTCCCA	GAGGGTGGTTGGTTCATT
ENA6		SSR	CTCGTCTTCTCACCTACAAC	CTGACATCTTCTCACCCAC
cnu_m008a		SSR	GTTGCTGGGCTTGCAGTTAT	GAGCGTACCAGCAACCTCTC
cnu_m016a		SSR	GGTGAATGGAATCTTGTCTTGA	CCCAACAATCCAGAAACAC
cnu_m020a		SSR	GGCTCTCCTCATCGTCAAAA	AATTCCGATTGCGACAAAAC
cnu_m029a		SSR	TACCCATTGGTGTCTCCAG	TCGTTCTCGAATGTGAATTGTC
cnu_m030a		SSR	GAAACAAATTTAAAAATCAGACCA	TGGAACAATCCGTAAAACATGC
cnu_m034a		SSR	TCACCGCCATAATTTGATCC	CCCTCTCAACAAGGTATGCAA
cnu_m037a		SSR	CCTAGTTCCTTGCACTCATGC	TTGTCTTTCAGATTGAAAACCTCG
cnu_m038a		SSR	GGCATGTGTCAATGAGTTGG	CTCCCACTCCTCCATTCAAC
cnu_m044a		SSR	TGTTTTGATCTTTACTGTTTTTGA	AATGTTTTTATATCACTATTGCCAAAT
cnu_m046a		SSR	GCTAAAGGTTTAGTCCAAATAGGATTC	GCAAAATGATGCCCCATAAA
cnu_m050a		SSR	AGCCCAAGCTCGTATTCTT	AAAATCGGGACAACCACCTA
cnu_m052a		SSR	GGAATCCTACGGAAGAGCAA	AAGGTAACGGTGGCAGTGAG
cnu_m062a		SSR	ATCGGCGCTGGTTATGTCA	CTAGGCTGCCCTTTCGATT
cnu_m068a		SSR	CCATATGACTAATTGACACTTTTGAA	TTCCCGAAAGTCTTCTTGG
cnu_m073a		SSR	TGGCATTGACAGAGCTAGTA	TTTATTTAGTTTCATACCCT
cnu_m090a		SSR	GCAAAGATCGGCGAAGAAGA	TGCAGACACATTGCAACAACA
cnu_m098a		SSR	TGCGACCCAAGTAGGTGAAAC	TGTCTCTCGCTCATTATCCAA
cnu_m100a		SSR	AAAGTTCACACAAATGATTTTGATATT	TTTTCTAGGAATGGTCCAACTT
cnu_m114a		SSR	AGTCGGAGGAAACGCGAAATTA	CGAAATAAAGACAGACAGAGACATCCA
cnu_m119a		SSR	ACACCTACTTGTTCATCCAAAT	CGGGTATTTGCGTTGTTTCC
cnu_m132a		SSR	CCATGGCCTCTCGTATTGCT	CCAACGGAGTGTCCCAATC
cnu_m139a		SSR	TCAAGCGCAACAAACATTGG	TGGTGTAGGGTTTAAAGGTTGTGG
cnu_m142a		SSR	GACCTTCGGTTCAGGGTATGG	CTGAACGGTCAATTTGTTTGG
cnu_m146a		SSR	TCATACCAACGTGCTTTGAAGA	GTGTGGCCGGATCTGATCTA
cnu_m148a		SSR	CACAAGCATTCTACCATAGCAAAGTC	TGCACATATGGCATGTTGTTTG
cnu_m149a		SSR	GGAAGCCTCTGTGCGAAAAA	TGCCGACGATTTGATAGAGGA
cnu_m157a		SSR	CCGCAGTTGATCCATTAGCC	ACGCTGCATCCACATGAAAC
cnu_m172a		SSR	GGAATGGAACACCGGATTAGC	TCGGATCTGATTTGTCGGATTT
cnu_m173a		SSR	TGTATTCCATTATTTCCGACTAACCT	CCGCATTTTAAAAACGTGAGAAA
cnu_m179a		SSR	TGGTTACACCTAGTTCCTTGCACTC	GGCCTTTGCCCGTTTAGTTTTA
cnu_m182a		SSR	TTCATCACCGTCTTATGTTGTGC	GGCAGGTGGAATATGTGGAAAT
cnu_m207a		SSR	GGACCCGGAATACCTCAAAGA	CATCAATAGCTCCGACACAATCC
cnu_m211a		SSR	TGTAAAGTTGTGCAAGGATTGTG	TGGTTTTGTGAAAATATGGTGAAA
cnu_m215a		SSR	CCAACCATTTCTGTTAGTCAACC	TTACGCATGTACCTGCACTAAAAA
cnu_m220a		SSR	ATCAGAACCGAATCCGACCA	CAATGGTTGCAATGTTATTTGGA

Supplementary Table 1. Contd.

cnu_m225a	SSR	TTGCGTTTTCTCGTCGTC	CCCCGAGATAAATGGCACAC
cnu_m241a	SSR	AATGCTGTGTCCATGACCAA	CGGGCATCCACCTAATTTGT
cnu_m246a	SSR	AAAGCCATCCATCCATCAAGC	GATGCAACATTTGACTGTGTTAGAGC
cnu_m252a	SSR	TGAAAATCAACACGAACACACAGA	CTCGTGGGGGAATGAGTGAG
cnu_m254a	SSR	AAGCTTGAGCTTCCAGCCTTC	ATCAGTGCCGGCCTTGAATA
cnu_m256a	SSR	TTGAAATACATGATACCCCAACCA	CCGTTTTCAGGGCACAGTTT
cnu_m257a	SSR	TGCATGATGTTTCATGTCTTGAAA	TCCTTCTGTAAACCGGTTGTAATTT
cnu_m263a	SSR	GAGGAAGTACGGCAAGAAACCA	AGGACACATGTCCACATGAAAA
cnu_m268a	SSR	TCATTGGTGAAGAACCACAAA	GCGACCATAAAAAGAGAGTGAGAA
cnu_m273a	SSR	ATAAGGGCATCGCCTCAACA	TGCACGCATCCACATAAACA
cnu_m277a	SSR	GCCATGAGCATTGCGTTAGG	TGAACTCTGGTTGGATTGACGA
cnu_m280a	SSR	TGTTACCACAGGAACCGTTCAA	CTTGGGCACACCATCATCTG
cnu_m284a	SSR	TCGGTTAAATCGAGTAACGATG	TTTCAGGACCTAGACGTTACCAA
cnu_m286a	SSR	AGTTGCCCTATTCATGCAC	AATGCGTTCATGTGGGGATA
cnu_m288a	SSR	GCGTTTTCGTCCTCTTCTCAC	TTACCCACCTTGGCTTCATC
cnu_m289a	SSR	CCCCTGGACTCCGTTTATCT	GATCTACGACGATCGGATGC
cnu_m293a	SSR	AAAAAGAAATGGATATTGTGTGAAA	CCTGGATCAAGACCACGAAG
cnu_m295a	SSR	GCTGCCTAATAGGGTGCTTG	AGAGCGCATTCAAGTCTGGT
cnu_m296a	SSR	TCTCGTCGCTCTGAATTGTG	TTGTGAAATCAAAGCAAAAAGG
cnu_m300a	SSR	AATTAGCGCGATAACATAAATAAAAA	AAATTGCTTAACTATTAATAACTGCAAA
cnu_m308a	SSR	GTTTGGGCCATCATGAAAA	TGGTTGCAAAATGTCACAGAA
cnu_m310a	SSR	GGCAGGTGGAATATGTGGAA	GCACTATCATCATCAAACAGAACA
cnu_m316a	SSR	TCAAGCATGTCCTTAAACTCTGA	GCGTTCACGTTTCCCATATC
cnu_m318a	SSR	TTATCAACATATTTTCAATCATTCCA	GCTTTGGACTATGCTTCTAAAGTACG
cnu_m320a	SSR	TTTTTCCTTTGGCTTAAACTGA	GCCAAAGCCACAAGATAACAT
cnu_m321a	SSR	TTGAATAATGACCCCAATATCA	TCAATAGGTATTAACCAATTCTACCG
cnu_m324a	SSR	TTTTCAACTCCACATGCAC	TGGGTATGTGCCAAATTGTTT
cnu_m327a	SSR	TTCTTGACCAAAAAGATCATGG	CTAACACGGGGAAAAGCAGA
cnu_m332a	SSR	TCGAACCGAAGTAAATAACGGACT	TTTCGCCCACTGACGCTATT
cnu_m338a	SSR	GCAACGATGAATCCCTAAACGA	AAATCCTCCCACTGTTTCCGAT
cnu_m344a	SSR	CCCAAATACGAAAACAAAGTTTGAC	AGGATCTCATCCGCTTTCCA
cnu_m354a	SSR	AAAGAAAACAAAGTGTCAATTGTCTCA	TCTACCGTTGAACCAGAGTTTTT
cnu_m356a	SSR	CGCATTTTCGCCGTCATTA	ACATCAGGCCGTCCTACTAA
cnu_m360a	SSR	ATCAGTGCCGGCCTTGAATA	AAGCTTGAGCTTCCAGCCTTC
cnu_m362a	SSR	CCTCTGCTGAAGGAGGCAAA	AGGTGGCTCTAGCGGAAGGT
cnu_m364a	SSR	ACCTGCCACCCTGTCAAAAC	GCACTAACCCTCCCTCTCCTC
cnu_m371a	SSR	TTTTTGGGTTTCTTCTCAAATGC	ACTCCAGCGAATTTGGCTTT
cnu_m372a	SSR	CCAGTGGCCAATACGAAACC	TGATGGAGAGTGGGTTGTGC
cnu_m377a	SSR	TCAGTTGTCGGATCGTCTATG	CACTTATCTTTCTTTGAAGTTGTTG
cnu_m379a	SSR	ACACCACTAAAACATTGCCATA	ACCGAAGGAGACTGCAAAGA
cnu_m384a	SSR	TGAAGGTGATGATGACGATGA	TCATGGTCTACAAAGACATACGG
cnu_m396a	SSR	TCATCATTAAAATGAGTAAAATTGCG	TTTTGGTGATCTTTTCTAAATTTTC
cnu_m397a	SSR	TCTTCAAGTCAAATACTCACATTCA	AAACGACAAATACATATGACAGTTTTA
cnu_m398a	SSR	TGACATTGCGATCAGATTTGT	TTGGGCTTCACGCATAAGAT
cnu_m400a	SSR	CGAGTTTTTGTGTGTACGTATAGTAAT	CCAAAGTGCGTAAAGGAAGG
cnu_m402a	SSR	GCCGACTCCTAGTGAGGAAA	TGTGTTTTGGGCTCAAAGGT
cnu_m409a	SSR	TTCCGGTCACTTCTAGCTTCA	TTTTGGTGGTTAGTATGTCGCTAT
cnu_m416a	SSR	TGGTGGGTCGTAACAGATGA	GCTCGCTTCCCAAATATGAA
cnu_m425a	SSR	TCGTTTGACCAACCGTACAA	CTTGCCAGCGTTGATACAGA
cnu_m439a	SSR	CCCTACGGACGGATGAGTAA	TCTGAGTGGCACCAGCATTAA
cnu_m442a	SSR	CGATTTGGACAATGACTAGTGG	AACGCCATGGAACAGAAAC
cnu_m446a	SSR	CACGTACGTCTTGGATGAATAAA	ATCTCACGTGGAGCACCATT



Supplementary Table 1. Contd.

cnu_m457a	SSR	CTGCTCCTTCACGTTTTTCATCA	ACGGACAGCAACAACAACAAGA
cnu_m458a	SSR	GGGGTGAATCTTGGATGAGG	CTGACGGATTCCCAACGAAT
cnu_m459a	SSR	CAAAGCCGGATTTCTTTTAGCA	TTTAAAAGTATTCTAACAAATCCGTTG
E039	ESTP	CTTGAGTGCTCAGGTCAAAGC	GAACCCTTACCCCAAGACTAC
E120	ESTP	ATCATAACCCTCAGGTTTGACATC	ACATCAAGCTCCTCTCTGGGTA
E129	ESTP	AGATGGTAAAAGAGCACAAAGCC	TTCAAGCTACCGATCCAAC TG
E138	ESTP	TGCTATCACAGTAGGGATTGCTT	CACTCCCACTCCTCCTAGTCC
atpm1	SRAP	CTCTTGGTGATTCAGCCAC	
bg10	SRAP	CGTTTCTTCTCGCATTCTC	
bg12	SRAP	TCTAAGACCTCCACAGTAAG	
bg14	SRAP	GCGTGGAAGCTGGAAGTCAAC	
bg16	SRAP	TGATACCACTTGCGATACCA	
bg17	SRAP	TGGTATCGCAAGTGGTATCA	
bg18	SRAP	GCAAGTCTCTCAGGTTATTC	
bg19	SRAP	GCTCTTCATCAGTTCTTGGT	
bg2	SRAP	GACCAAATATAAAACACTAACTA	
bg20	SRAP	TCCTCTCCACTTTTGTCTTC	
bg21	SRAP	AACTCGCTTGCTTAGATATG	
bg24	SRAP	CACCTTTTCTACTCCTATC	
bg3	SRAP	GGAACACTTAATGGTACGGT	
bg42	SRAP	ACACATAATCTTCTACAAATAC	
bg50	SRAP	AAGTCGTTGTAGTATAGTGG	
me1	SRAP	TGAGTCCAAACCGGATA	
me2	SRAP	TGAGTCCAAACCGGAGC	
me3	SRAP	TGAGTCCAAACCGGAAT	
me4	SRAP	TGAGTCCAAACCGGACC	
me5	SRAP	TGAGTCCAAACCGGAAG	
me6	SRAP	TGAGTCCAAACCGGACA	
me7	SRAP	TGAGTCCTTTCCGGTAA	
me8	SRAP	TGAGTCCAAACCGGACG	
me9	SRAP	TGAGTCCTTTCCGGTCC	
me10	SRAP	TGAGTCCAAACCGGACT	
me11	SRAP	TGAGTCCTTTCCGGTGC	
me12	SRAP	TGAGTCCAAACCGGTAG	
me13	SRAP	TGAGTCCAAACCGGTCA	
me14	SRAP	TGAGTCCAAACCGGTAA	
me15	SRAP	TGAGTCCAAACCGGTGC	
me16	SRAP	TGAGTCCAAACCGGAAC	
me17	SRAP	TGAGTCCAAACCGGCAT	
em1	SRAP	GACTGCGTACGAATTAAT	
em2	SRAP	GACTGCGTACGAATTTGC	
em3	SRAP	GACTGCGTACGAATTGAC	
em4	SRAP	GACTGCGTACGAATTTGA	
em5	SRAP	GACTGCGTACGAATTAAC	
em6	SRAP	GACTGCGTACGAATTGCA	
em7	SRAP	GACTGCGTACGAATTCAA	
em8	SRAP	GACTGCGTACGAATTCAC	
em9	SRAP	GACTGCGTACGAATTACG	
em10	SRAP	GACTGCGTACGAATTGAT	
em11	SRAP	GACTGCGTACGAATTATG	
em12	SRAP	GACTGCGTACGAATTCGA	
em13	SRAP	GACTGCGTACGAATTTAG	

**Supplementary Table 1.** Contd.

em14	SRAP	GACTGCGTACGAATTTTCG
em15	SRAP	GACTGCGTACGAATTGTC
em16	SRAP	GACTGCGTACGAATTGGT

**Supplementary Table 2.** Detail of the primers for detection of DNA polymorphism and assigned linkage group.

Marker assay name	Assay type	Polymorphism	Assigned linkage group
KBRH138G23-700	SSR	O	A3
KBRH139B23	SSR	x	x
KBRH143D22	SSR	O	x
KBRH143F19	SSR	x	x
KBRH143H15	SSR	O	x
KBRH143K20-320	SSR	O	A7
KBRH048O11	SSR	O	x
EJU3	SSR	x	x
EJU5-130	SSR	O	A6
ENA17	SSR	O	x
ENA18	SSR	x	x
ENA23-200	SSR	O	A6
ENA28	SSR	x	x
ENA4	SSR	O	x
ENA6	SSR	x	x
cnu_m008a	SSR	x	x
cnu_m016a	SSR	x	x
cnu_m020a	SSR	O	x
cnu_m029a-190	SSR	O	A5
cnu_m030a	SSR	x	x
cnu_m034a	SSR	x	x
cnu_m037a	SSR	x	x
cnu_m038a	SSR	x	x
cnu_m044a	SSR	x	x
cnu_m046a	SSR	x	x
cnu_m050a	SSR	x	x
cnu_m052a	SSR	x	x
cnu_m062a-200	SSR	O	A7
cnu_m068a	SSR	x	x
cnu_m073a	SSR	O	x
cnu_m090a	SSR	O	x
cnu_m098a	SSR	x	x
cnu_m100a-300	SSR	O	A3
cnu_m114a	SSR	x	x
cnu_m119a	SSR	x	x
cnu_m132a	SSR	x	x
cnu_m139a	SSR	O	x
cnu_m142a-180	SSR	O	A1
cnu_m146a	SSR	O	x
cnu_m148a	SSR	x	x
cnu_m149a	SSR	x	x
cnu_m157a	SSR	x	x
cnu_m172a	SSR	x	x

Supplementary Table 2. Contd.

cnu_m173a	SSR	x	x
cnu_m179a	SSR	x	x
cnu_m182a-420	SSR	O	A7
cnu_m207a	SSR	x	x
cnu_m211a	SSR	O	x
cnu_m215a	SSR	x	x
cnu_m220a-500	SSR	O	A6
cnu_m225a	SSR	x	x
cnu_m241a	SSR	x	x
cnu_m246a	SSR	x	x
cnu_m252a	SSR	x	x
cnu_m254a	SSR	O	x
cnu_m256a	SSR	x	x
cnu_m257a-100	SSR	O	A5
cnu_m263a-360	SSR	O	A9
cnu_m268a	SSR	x	x
cnu_m273a	SSR	O	x
cnu_m277a	SSR	x	x
cnu_m280a	SSR	x	x
cnu_m284a	SSR	x	x
cnu_m286a	SSR	x	x
cnu_m288a	SSR	x	x
cnu_m289a-170	SSR	O	A5
cnu_m293a-180	SSR	O	A5
cnu_m295a	SSR	x	x
cnu_m296a	SSR	x	x
cnu_m300a-260	SSR	O	A8
cnu_m308a	SSR	x	x
cnu_m310a	SSR	O	x
cnu_m316a	SSR	x	x
cnu_m318a-320	SSR	O	A6
cnu_m320a	SSR	x	x
cnu_m321a	SSR	x	x
cnu_m324a	SSR	O	x
cnu_m327a	SSR	O	x
cnu_m332a	SSR	x	x
cnu_m338a	SSR	x	x
cnu_m344a-150	SSR	O	A5
cnu_m354a-400	SSR	O	A7
cnu_m356a	SSR	x	x
cnu_m360a	SSR	O	x
cnu_m362a-320	SSR	O	A5
cnu_m364a	SSR	x	x
cnu_m371a	SSR	x	x
cnu_m372a	SSR	x	x
cnu_m377a-730	SSR	O	A9
cnu_m379a	SSR	x	x
cnu_m384a	SSR	O	x
cnu_m396a	SSR	x	x
cnu_m397a	SSR	x	x
cnu_m398a	SSR	x	x
cnu_m400a	SSR	x	x

Supplementary Table 2. Contd.

cnu_m402a	SSR	x	x
cnu_m409a	SSR	x	x
cnu_m416a	SSR	O	x
cnu_m425a-250	SSR	O	A5
cnu_m439a	SSR	x	x
cnu_m442a	SSR	x	x
cnu_m446a-230	SSR	O	A5
cnu_m457a	SSR	x	x
cnu_m458a-380	SSR	O	A5
cnu_m459a	SSR	x	x
hri_mBRMS082a-200	SSR	O	A2
E039-1000	ESTP	O	A10
E120-630	ESTP	O	A8
E129	ESTP	x	x
E138	ESTP	O	x
bg50me10-170	SRAP	O	A1
em4bg10-180	SRAP	O	A1
me6bg19-140	SRAP	O	A1
em16me10-300	SRAP	O	A1
em3bg17-150	SRAP	O	A1
em4bg2-580	SRAP	O	A1
bg2bg21-320	SRAP	O	A1
em10me7-130	SRAP	O	A1
em7me5-160	SRAP	O	A1
em5bg19-340	SRAP	O	A1
em16me3-220	SRAP	O	A2
me9bg2-320	SRAP	O	A2
em10me8-250	SRAP	O	A2
em9me2-310	SRAP	O	A2
em7bg20-160	SRAP	O	A2
me11bg3-250	SRAP	O	A2
em8bg18-110	SRAP	O	A2
me1bg16-270	SRAP	O	A2
em10me4-200	SRAP	O	A2
em10me5-160	SRAP	O	A2
me13em16-200	SRAP	O	A2
me9bg18-140	SRAP	O	A2
bg19bg50-250	SRAP	O	A3
em5bg2-190	SRAP	O	A3
bg50me2-130	SRAP	O	A3
em7bg19-220	SRAP	O	A3
em8bg20-240	SRAP	O	A3
me11bg21-220	SRAP	O	A3
me6bg20-270	SRAP	O	A3
em2me2-620	SRAP	O	A3
em4bg24-590	SRAP	O	A3
bg12bg19-380	SRAP	O	A3
me17bg20-300	SRAP	O	A3
me6bg24-240	SRAP	O	A4
em9me1-750	SRAP	O	A4
bg2bg50-320	SRAP	O	A4
me1bg12-750	SRAP	O	A4

Supplementary Table 2. Contd.

me13em1-100	SRAP	O	A4
me13em5-340	SRAP	O	A4
me13bg18-160	SRAP	O	A4
me10bg18-270	SRAP	O	A4
bg20bg3-190	SRAP	O	A4
me3bg12-570	SRAP	O	A4
bg14bg3-130	SRAP	O	A4
em4me3-450	SRAP	O	A4
em2bg17-180	SRAP	O	A4
em5me12-330	SRAP	O	A5
me2bg20-240	SRAP	O	A5
me12atpm1-120	SRAP	O	A5
me10bg3-180	SRAP	O	A5
me5bg24-200	SRAP	O	A5
bg50me12-500	SRAP	O	A5
bg19bg24-200	SRAP	O	A5
me13bg21-140	SRAP	O	A5
em5bg21-340	SRAP	O	A6
me16bg20-200	SRAP	O	A6
me16bg16-130	SRAP	O	A6
me5bg2-150	SRAP	O	A6
em7bg12-240	SRAP	O	A6
me12bg24-210	SRAP	O	A6
me17bg10-110	SRAP	O	A6
em4bg21-170	SRAP	O	A6
me12bg19-220	SRAP	O	A6
me14bg17-200	SRAP	O	A6
em10me11-370	SRAP	O	A6
em1bg12-170	SRAP	O	A6
bg19bg20-170	SRAP	O	A6
bg2bg24-380	SRAP	O	A6
me5bg12-240	SRAP	O	A6
me4bg12-150	SRAP	O	A6
em8bg10-150	SRAP	O	A7
me2bg24-260	SRAP	O	A7
em15me1-570	SRAP	O	A7
em2bg16-300	SRAP	O	A7
me12bg2-480	SRAP	O	A7
em2me1-640	SRAP	O	A7
em14me4-260	SRAP	O	A7
bg24bg50-250	SRAP	O	A7
me14bg14-110	SRAP	O	A7
me9bg20-310	SRAP	O	A7
me8bg3-180	SRAP	O	A7
me13em15-190	SRAP	O	A7
bg18bg21-380	SRAP	O	A7
me13em4-240	SRAP	O	A8
em9me4-240	SRAP	O	A8
me10bg10-210	SRAP	O	A8
me9bg10-190	SRAP	O	A8
em3bg19-110	SRAP	O	A8
em3me2-120	SRAP	O	A8

**Supplementary Table 2.** Contd.

me15bg20-320	SRAP	O	A8
bg20bg21-150	SRAP	O	A8
me7bg2-100	SRAP	O	A8
em3bg10-140	SRAP	O	A9
em5bg17-260	SRAP	O	A9
em4bg12-150	SRAP	O	A10
me7bg18-200	SRAP	O	A10
me10bg14-200	SRAP	O	A10
me6atpm1-240	SRAP	O	A10
bg19bg42-250	SRAP	O	A10
me13em7-250	SRAP	O	A10
em5me11-370	SRAP	O	A10
me4bg20-150	SRAP	O	A10
me4bg20-240	SRAP	O	A10
e10me10-180	SRAP	O	A10
bg21bg42-370	SRAP	O	A10
em2me3-160	SRAP	O	A10
em3atpm1-150	SRAP	O	A10
em8bg19-130	SRAP	O	x
me12bg10-240	SRAP	O	x
em1bg10-260	SRAP	O	x
bg12bg50-370	SRAP	O	x
me14bg10-240	SRAP	O	x
me17bg24-330	SRAP	O	x
me5bg12-270	SRAP	O	x
bg18bg20-120	SRAP	O	x
me3bg18-140	SRAP	O	x
me14bg21-190	SRAP	O	x
bg2bg42-270	SRAP	O	x
me1atpm1-160	SRAP	O	x
me11bg18-330	SRAP	O	x